SYNTHESIS and characterizations of new Mn(II), Co(II), Ni(II), Cr(III) and Fe(III) melatonin drug complexes have been studied. The melatonin ligand act as a monodentate ligand through nitrogen of the deprotonated –NH pyrrole ring. This was confirmed by infrared spectra and microanalytical analysis. The magnetic moments data approved the complexity of Mn(II), Co(III), Ni(III) as a square planar and Cr(III), Fe(III) as an octahedral geometry, respectively. The biological activity of the melatonin free ligand is lower than that of the respected metal complexes. Thus, this indicates that the complexity is more effective than the antimicrobial activities of melatonin ligand.

Keywords: Melatonin drug, Synthesis, Antimicrobial, Complexes, Ligand.
two techniques, IR and differential scanning calorimetry [53]. The studies reveal that the physical properties of the DPPC bilayers changes by melatonin Asymmetric double bond vibrations. Furthermore, there are more than one spectrum is produced at large melatonin concentrations, which refers that melatonin- afforded a phase separation in the DPPC membranes.

The prevention of the lead toxicity is of a high international public health priority [54, 55]. The evaluation of the melatonin effects, the antioxidant and the free radical scavenging activities, on lead formed neurotoxicity and oxidative stress. Previous results suggested that melatonin treatment can improve the oxidative stress of impotence and disability by protecting the brain from lead toxicity. Due to the essential biological activities of this molecule, melatonin has been subjected of several structural and spectroscopic studies.

Melatonin and glutathione binding has been investigated by using the titration calorimetry and by using UV–vis absorption spectroscopy, Fourier transform infrared spectroscopy [56]. Thermodynamic investigations with addition of the daily doses of 0.2% Pb(CH₃COO)₂ to drinking with the distilled water and continues till the weaning. Melatonin was administrated once daily. After 21 days, Multi antioxidant enzyme activities as superoxide dismutase and glutathione peroxidase were evaluated. Lipid peroxides levels were measured as a marker of lipid peroxidation confirmed that melatonin/glutathione correlates to HSA which is driven by favorable entropy. The major forces are hydrogen bond and van der Waals. The interaction for glutathione is recognized by a high number of the binding sites, which informs that the binding takes place by adsorption surface which leads to the protein surface coating. Regarding the melatonin, one molecule of melatonin conjugates with one molecule of HSA and no binding occurs between more melatonin with HSA in this study. The UV–vis analysis, IR, and spectroscopy suggested that melatonin and glutathione may afford a micro environmental changes of HAS. The experimental evidences suggested that the melatonin is responsible for influencing the diabetic complications [57] by reducing the unnecessary reactive oxygen production and protection of the beta cells, as they have low antioxidant capacity and normalize the oxidative status in the cells. Nitric oxide is known to induce a lot of nephrotic diseases that afforded by the lead toxicity and the current works have shown the role of the antioxidants in reducing the lead toxic effects. Melatonin is known for its direct and indirect antioxidant activities. Additionally, it is produced by natural way in mammalians. Additionally, [58] reported that melatonin alleviates the lead toxic effects in the kidneys and with no alterations of NO metabolites. The synthesis of a new fluorescent coumarin-that containing analogues of melatonin was presented [59].

Nano system types, namely as lecithin/chitosan nanoparticles and some types of chitosan micelles, have been prepared and characterized [60] for their potential in melatonin delivery, which is known to have hypotensive effect. The melatonin particle size and properties were studied as a function with presence of the nanosystem. Supplementation with melatonin are widely used in Europe and in the USA to attenuate delay of sleep disorders [61]. The European Commission approved mainly two health claims for the food that containing a melatonin. The method of determination of melatonin dose in food that was marketed in Europe by confirming a liquid chromatography with diode array detection method. Contaminants that are present in melatonin supplements were estimated by using high- liquid performance chromatography and mass spectrometer.

<Fig. 1. Melatonin structure.>
Experimental

Chemicals, reagents and instrumentals

The chemicals that were used from (Merck which is the high purest grade. The drug melatonin was purchased from The Egyptian Pharmaceutical Company (EIPICo.). C and H amounts were detected using a Perkin-Elmer. The metal amount was measured by converting the complexity into their analogous stable oxide forms. Infrared spectra were estimated by Bruker FT-IR spectrophotometer (ranged from 4000–400 cm⁻¹) in KBr pellets. Molar conductivities of freshly prepared DMSO solutions were estimated by using Jenway. The X-ray diffraction patterns for the solid complexes were recorded on X’Pert PRO PANanalytical X-ray powder diffraction, target copper with secondary monochromate. Scanning electron microscopy (SEM) images were taken in Quanta FEG 250 equipment. The mass susceptibility (Xg) of the solid paramagnetic Chromium(III), Manganese (II), Iron(III), Cobalt(II), and Nickel(II) complexity was detected using Gouy’s method at the room temperature by using a magnetic balance at central lab at Cairo university. The effective magnetic moment (μ_eff) values were obtained by using the following equations (1, 2 and 3) [62].

\[
X_g = \frac{C_{Bal} L (R - R_o)}{10^9 M} \quad \ldots \ldots \ldots \ldots (1)
\]

Where:
- \(R_o\) = Empty tube reading
- \(L\) = Length of the sample \((\text{cm})\)
- \(M\) = Mass of the sample \((\text{gm})\)
- \(R\) = The reading for tubes with samples
- \(C_{Bal}\) = Constant of calibration balance = 2.086
- \(X_g = X \times M\). \(W_t\) \ldots \ldots (2)

The amounts of \(X_g\) were estimated from equation (2) and were corrected by using the Pascal’s constants, and were applied in the Curie’s equation (3).

\[
\mu_{eff} = 2.84 \sqrt{X_g M} \times T \quad \ldots \ldots \ldots \ldots \ldots \ldots \ldots \ldots \ldots \ldots \ldots \ldots \ldots (3)
\]

Where \(T = t \ (°C) + 273\)

Thermogravimetric Analysis (TGA) and Differential Thermal Analysis (DTA) were estimated by special thermal analyzers at Cairo university. All the experiments were carried out by a single loose top loading platinum sample pan under \(N_2\) atmosphere at a flow level of 30 ml/min and a 10°C/ min with using temperature range 25-800 °C.

Antibacterial and anti-fungal evaluation

Regarding Gupta et al., [63] hole-well methods that were applied. The examined bacterial isolates were seeded in tubes with the nutrient media. The seeded NB was homogenized with the melted agar media. The suspensions were poured in the Petri dishes. The inhibition zone diameter that was over than 7 mm indicated that the tested compounds were active against the bacterial activity. The antibacterial activities were tested against Escherichia coli, Bacillus subtilis and anti-fungal (Asperagillus oryzae, niger and Flavus).

Synthesis of melatonin complexes

The Chromium(III), Manganese (II), Iron(III), Cobalt(II), and Nickel(II) complexity were synthesized by reactions \(\text{CrCl}_3\), \(\text{MnCl}_2\), \(\text{H}_2\text{O}, \text{FeCl}_3\), \(\text{2H}_2\text{O}, \text{CoCl}_3\), \(\text{6H}_2\text{O}\) and \(\text{NiCl}_2\), \(\text{6H}_2\text{O}\) metals \((1 \ \text{mmol}; 20 \ \text{mL} \ \text{distilled water})\) to melatonin \((2 \ \text{mmol}; 20 \ \text{ml} \ 99\% \ \text{Methanal})\) with molar ratio 1:2 for all complexity except \(\text{Cr(III)}\) and \(\text{Fe(III)}\) is 1:3 ratio. Concentration of hydrogen for melatonin metal ions complexity were adjusted between 7-9 by using 5% Ammonium hydroxide/Methanol. The obtained solutions were stirred and refluxed on a hot plate at 60 °C for about 1 h. The products were taken, filtered and were washed with mixing of the distilled water with \(\text{CH}_3\text{OH}\) then were dried at 70 °C and remains under vacuum over the anhydrous \(\text{CaCl}_2\).

Results and Discussions

Physical properties

MLT ligand physical data and their complexity with Chromium(III), Manganese (II), Iron(III), Cobalt(II), and Nickel(II) are in Table 1. The product complexity are stable in the air, and with high melting points. Water and most organic solvents insoluble except in DMSO and in DMF. The ligand character of MLT with some metal ions were investigated by the infrared spectra, thermogravimetric analyses and molar conductance. The measuring of \((\text{C}, \text{H}, \text{N})\) of the complexity show 1:2 or 1:3 (metal: MLT) stoichiometry for Manganese (II), Cobalt(II), Nickel(II) (Chromium(III) and Iron(III)) complexity, respectively. The amount of \(\text{C}, \text{H}, \text{N}\) were detected at room temperature for Manganese(II), Cobalt(II), Nickel(II), Chromium(III) and Iron(III) MLT complexity and have paramagnetic characters. The molar conductance data of the complexity which is low refers to non-electrolytic nature [64]; this proves the suggestion for Cl anions absence which is low refers to non-electrolytic nature [64];
Infrared spectra

IR spectra of melatonin and its complexity were measured in Fig. 3 and Table 2. The IR spectra of melatonin complexity are correlated; this may be due to the coordination site toward Cr(III), Mn(II), Fe(III), Co(II) and Ni(II) ions which are the same. The most importance property of the IR spectra of the complexity is the disappearance of a band at 3360 cm\(^{-1}\). The strong band at 3300 cm\(^{-1}\) can be attributed to NH (amide) which is not affected; this indicates that the NH group is not participated in the coordination mechanism. Spectrum at 1621 cm\(^{-1}\), due to C=O stretching vibration, is not affected in the complexity and has low intensity. Spectra at 1315 and 1214 cm\(^{-1}\) in MLT do not shift in complexity. Nakamoto [66] attributed to the spectra at 1315 and 1214 cm\(^{-1}\) in the metal oxamido complexes to the Carbon-Nitrogen vibration.

The fact is difficult to reconcile with the proposed structures by Fazakerley et al. [65]. Strong bands at 3300 cm\(^{-1}\) can be attributed to NH (amide) which is not affected; this indicates that the NH group is not participated in the coordination mechanism. Spectrum at 1621 cm\(^{-1}\), due to C=O stretching vibration, is not affected in the complexity and has low intensity. Spectra at 1315 and 1214 cm\(^{-1}\) in MLT do not shift in complexity. Nakamoto [66] attributed to the spectra at 1315 and 1214 cm\(^{-1}\) in the metal oxamido complexes to the Carbon-Nitrogen vibration. If we consider that the 1315 and 1214 cm\(^{-1}\) bands were due to

![Diagram](image-url)
C-N vibration, we deduce that the NHCO group was not participated in coordination process. The free MLT has a band in C=O region, at 1170 cm⁻¹. After the complexity, the maxima is the same without changes, we conclude that the C-O association probably did not share the complexity, the presence of a medium to weak bands at (523-622)cm⁻¹ can be owned to Metal-Nitrogen stretching vibration motion [67].

Magnetic measurements

Magnetic measurements were obtained by Gouy method [72] and the observed values for the iron(III) and chromium(III) complexes are 5.26 and 3.64 BM in the accepted scale with an octahedral geometry [68, 69]. The Mn(II) complex shows a magnetic moment (5.92 B.M) which are appreciably close to the calculated spin-only value for five unpaired electrons, and reveal a high spin state, therefore, the structure of the prepared Mn(II) complex. This indicates a square planer geometry around Mn(II) ion [70, 71]. The magnetic data of the [Co(MLT),(H₂O)₃]₂H₂O complex is agreement with to that reported for related cobalt(II) complexes [70]. This geometry is confirmed by the values of the effective magnetic moment (1.81 BM) suggests a square-planar geometry. The square planar Ni (II) complexes are a diamagnetic (0.0 B.M.) meanwhile, the tetrahedral complexes have moments in (3.20–4.10 B.M), and octahedral complexes should have moments between 2.90–3.30 B.M. [72-74]. The Ni(II) complex gave a moment of 0.0 B.M. and hence attributed as a square planer.

Thermal analysis of MLT complexes

The thermo gravimetric analysis occurs at 10°C/min at N₂ atmosphere. The weight loss was measured up to 136 °C. The thermal products are shown in Table 3 and shown in Fig. 4. The losses in weight for each complexity were estimated in the ranges of the correlating temperatures.

[Cr(MLT),(H₂O)₃]·H₂O

The thermal degradation of [Cr(MLT),(H₂O)₃]·H₂O complex takes place at 3 steps. The 1st decomposition step occurs in temperature range (50-230°C) and this refers to the loss of (H₂O), weight loss (Obs.=2.46%, calc = 2.20%). The 2nd step occurs (230-417°C) that is attributed to the 3H₂O+C₆H₂₅OH loss (Organic moiety) (obs. = 40.06%, calc = 40.02%). The last degradation step takes place at temperature range 430-490°C, while the weight loss is (Obs.=48.32%, calc= 48.47%), which are attributed to C₅H₃NO₂⁻ loss. The last residue CrO₂.₅ remains stable till 800 °C.

[Mn(MLT),(H₂O)₃]·H₂O

The Mn(II) complex is degraded in 2 steps. The 1st step is occurred at 50-300°C and corresponding to the loss of 2H₂O+C₁₈H₁₂O₂ (Organic moiety) (obs. = 55.06%, calc. = 55.33%). The 2nd step occurs in the range (300–500°C) and can be attributed due to C₆H₅NO loss (obs. = 52.02%, calc. = 52.34%). The last residue was produced at 800°C is MnO+C.

[Fe (MLT),(H₂O)₃]·H₂O

Fe (III) complex decomposed thermally at 4 steps. The 1st step occurs in (40-80 °C) and they were attributed to the loss of 1.5 H₂O molecule (obs. = 3.20%, calc. = 3.28%). The 2nd step is (80-160 °C) which is attributed to the loss of 2.5 H₂O with a weight loss (obs. = 5.79%, calc. = 5.48%). The 3rd step occurs within the temperature (160-280°C) can be assigned to loss C₆H₉NO₃ with representing weight loss (obs. = 35.84%, calc. = 35.08%). The last degradation step in (280-450 °C) is joined by (calc. = 28.86%), which are attributed to the loss of C₆H₅N₂O₄. The FeO1½+12C are the last product that remains stable till the temperature 800 °C.

[Co(MLT),(H₂O)₃]·H₂O

Cobalt complexity is degraded in 3 steps. The 1st step was obtained at 50-100°C and was corresponding to 4(H₂O) molecules evolution, that represent (Obs.=12.31%, Calc= 12.14%). While, the 2nd decomposition step is carried out at 100-277°C and attributed to the loss of C₁₈H₂₅NO (Obs= 23.61%, Calc=23.60%). The last degradation step takes at 277-490°C and was attributed to C₆H₅NO₃ loss with (Obs= 33.62% and Calc= 33.89%). The residue resulted at 800°C which is CoO.

[Ni(MLT),(H₂O)₃]·2H₂O

Ni(II) compound is degraded in 3 steps. The 1st step is carried out at 50-105°C and refers to 4(H₂O) loss, that represent the weight loss as follows (obs. =12.17%, calc= 12.14%). The 2nd step occurs in (105-280°C) and can be attributed to C₆H₆NO loss (obs. = 23.58%, calc=23.62%).The last step is occurring at (280-500 °C) and attributed to C₆H₅NO₃ loss that represent a weight loss of 33.12 %, (Calc= 33.40%). The final residues were obtained at 800°C are NiO+9C.

TABLE 2. Infrared spectral bands and assignments of (MLT) complexes.

<table>
<thead>
<tr>
<th>Compound</th>
<th>$\nu$(N-H) Indole</th>
<th>$\nu$(N-H) amide</th>
<th>$\nu$(C=O) amide</th>
<th>$\nu$(C-N)</th>
<th>$\nu$(C-O)</th>
<th>$\nu$(M-N)</th>
</tr>
</thead>
<tbody>
<tr>
<td>MLT</td>
<td>3360</td>
<td>3300</td>
<td>1620</td>
<td>1315, 1213</td>
<td>1170</td>
<td>-----</td>
</tr>
<tr>
<td>Cr</td>
<td>-----</td>
<td>3298</td>
<td>1622</td>
<td>1315, 1213</td>
<td>1171</td>
<td>621, 592, 564</td>
</tr>
<tr>
<td>Mn</td>
<td>-----</td>
<td>3299</td>
<td>1621</td>
<td>1315, 1213</td>
<td>1170</td>
<td>622, 592, 564</td>
</tr>
<tr>
<td>Fe</td>
<td>-----</td>
<td>3298</td>
<td>1622</td>
<td>1315, 1213</td>
<td>1171</td>
<td>622, 592, 539</td>
</tr>
<tr>
<td>Co</td>
<td>-----</td>
<td>3299</td>
<td>1623</td>
<td>1315, 1213</td>
<td>1171</td>
<td>626, 593, 523</td>
</tr>
<tr>
<td>Ni</td>
<td>-----</td>
<td>3297</td>
<td>1621</td>
<td>1315, 1213</td>
<td>1172</td>
<td>626, 590</td>
</tr>
</tbody>
</table>

(a) (b)

Fig. (3a). IR of Mn /MLT complex; (3b): IR of Fe /MLT complex.

Fig. 4. TG/DTG of Cr /MLT complex.
TABLE 3. Thermogravimetric data of the MLT complexes.

<table>
<thead>
<tr>
<th>Compound</th>
<th>Steps</th>
<th>DTG peak (˚C)</th>
<th>TG Weight loss (%)</th>
<th>Assignments</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td>Calc.</td>
<td>Found</td>
</tr>
<tr>
<td>Cr(III)</td>
<td>1st</td>
<td>230</td>
<td>2.2</td>
<td>2.46</td>
</tr>
<tr>
<td></td>
<td>2nd</td>
<td>417</td>
<td>40.02</td>
<td>40.06</td>
</tr>
<tr>
<td></td>
<td>3rd</td>
<td>490</td>
<td>48.47</td>
<td>48.32</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mn(II)</td>
<td>1st</td>
<td>300</td>
<td>55.33</td>
<td>55.06</td>
</tr>
<tr>
<td></td>
<td>2nd</td>
<td>500</td>
<td>29.65</td>
<td>30.08</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Fe(III)</td>
<td>1st</td>
<td>80</td>
<td>3.28</td>
<td>3.20</td>
</tr>
<tr>
<td></td>
<td>2nd</td>
<td>160</td>
<td>5.48</td>
<td>5.79</td>
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<tr>
<td></td>
<td>3rd</td>
<td>280</td>
<td>35.08</td>
<td>35.84</td>
</tr>
<tr>
<td></td>
<td>4th</td>
<td>450</td>
<td>28.86</td>
<td>28.90</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Co(II)</td>
<td>1st</td>
<td>100</td>
<td>12.14</td>
<td>12.31</td>
</tr>
<tr>
<td></td>
<td>2nd</td>
<td>277</td>
<td>23.60</td>
<td>23.61</td>
</tr>
<tr>
<td></td>
<td>3rd</td>
<td>490</td>
<td>33.89</td>
<td>33.62</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Ni(II)</td>
<td>1st</td>
<td>105</td>
<td>12.14</td>
<td>12.17</td>
</tr>
<tr>
<td></td>
<td>2nd</td>
<td>280</td>
<td>23.62</td>
<td>23.58</td>
</tr>
<tr>
<td></td>
<td>3rd</td>
<td>500</td>
<td>33.40</td>
<td>33.12</td>
</tr>
<tr>
<td></td>
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<td></td>
</tr>
</tbody>
</table>

**XRD and SEM investigations**

XRD and SEM analysis showed the crystalline nature of the metal complexes. The samples of melatonin solid complexes were characterized at the room temperature by the X-ray diffraction by using the Cu Kα radiation. The X-ray diffraction patterns of the synthesized MLT complexes are crystalline in nature. The diffraction characterization of the synthesized MLT complexes recorded in between 4° to 90°. The crystalline size of synthesized complexes is calculated using the Scherrer formula [63] D=kλ/ bCosθ, where k is a constant and equal 0.94, λ the wavelength of X-ray used (0.154 nm) and b is a full in width at half maxima peak of XRD pattern. The crystalline size was found for cobalt and nickel complexes 50 nm and 30 nm, respectively. It is observed that crystalline size is different for both the complexes, due to change in the metal ions. The XRD patterns are shown in Fig. 5.

The SEM image of the MLT complexes are shown in Fig. 6. From this figure, it can be seen that the average length of the grains for the cobalt(II) and nickel(II) complexes are 50-100 μm, respectively. The surface morphology changes with change in metal ions, both the images having large number of irregular shaped and some having regular grains.

**Antimicrobial activity**

As shown in (Fig 5 and Table 4), The antimicrobial activities were elevated in this order: Mn⁺²(MLT(1.10-2.70 cm) > Fe⁺³ MLT(1.50-2.50 cm) > Ni⁺² MLT(1.00-2.00 cm) > Co⁺² MLT(0.50-1.50 cm) > Cr⁺³ MLT(1.00-1.30 cm).Cr⁺³ MLT was the least active being active against five bacteria and fungi, Escherichia coli (Gram−ve), Bacillus subtilis (Gram +ve) and anti-fungal (Asperagillus oryzae, niger, Flavus). The high MLT complexes sensitivity have been assigned to hyper-conjugation of the coordinated Lewis bases, which elevates all the electrons density on the metal ions coordinated form with highest antimicrobial activity [64].

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**Conflict of Interest**

The authors declare no potential conflicts of interest with respect to the research, authorship, and publication of this article.
Fig. 5. XRD spectrum of Co(II) complex; (b) XRD spectrum of Ni(II) complex.

Fig. 6. SEM spectrum of Co(II) complex; (b) SEM spectrum of Ni(II) complex.

Fig. 7. Inhibition zone diameter (mm/mg) sample of DMSO (Control) and MLT complexes.

TABLE 4. The inhibition zone diameter (mm/mg sample) of complexes against some kind of bacteria and fungi.

<table>
<thead>
<tr>
<th>Complexes</th>
<th>E. Coli</th>
<th>B. subtilis</th>
<th>Asperagillus oryzae</th>
<th>Asperagillus niger</th>
<th>Asperagillus Flavus</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>0.0</td>
<td>0.0</td>
<td>0.0</td>
<td>0.0</td>
<td>0.0</td>
</tr>
<tr>
<td>Cr\textsuperscript{3}MLT</td>
<td>0.0</td>
<td>1.3</td>
<td>1.0</td>
<td>1.1</td>
<td>1.5</td>
</tr>
<tr>
<td>Mn\textsuperscript{2}MLT</td>
<td>2.7</td>
<td>1.5</td>
<td>0.0</td>
<td>1.1</td>
<td>1.5</td>
</tr>
<tr>
<td>Fe\textsuperscript{3}MLT</td>
<td>2.5</td>
<td>2.3</td>
<td>0.0</td>
<td>1.5</td>
<td>1.5</td>
</tr>
<tr>
<td>Co\textsuperscript{2}MLT</td>
<td>1.5</td>
<td>0.5</td>
<td>0.0</td>
<td>1.0</td>
<td>1.5</td>
</tr>
<tr>
<td>Ni\textsuperscript{2}MLT</td>
<td>2.0</td>
<td>2.0</td>
<td>0.0</td>
<td>1.5</td>
<td>1.0</td>
</tr>
</tbody>
</table>

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