



Gene Downregulation by Natural Products for Alleviating Dyslipidemia

Miral O. Sabry^{a,*}, Mohamed S. Sedeek^a, Marwa Y. Issa^a, Soheir M. El Zalabani^a



CrossMark

^aPharmacognosy Department, Faculty of Pharmacy, Cairo University, Kasr El-Ainy Street, Cairo 11562, Egypt

Abstract

The levels of lipids in our bodies are regulated through regulatory proteins known as sterol regulatory element binding proteins (SREBPs). Dietary triglycerides are transported from the liver and intestine to body tissues by the microsomal triglyceride transfer protein (MTP). The role of the dysregulation of lipid components in metabolic diseases and cancer is reviewed herein. The current survey provides a comprehensive overview of published data spanning from 1997 to 2023, sourced from Scopus, Google Scholar, PubMed, Web of Science, and ScienceDirect, concerning dyslipidemia and the downregulation of SREBP-1, SREBP-2, and MTP gene expression by natural products. Natural products, mainly phenolic compounds such as hesperidin, luteolin, xanthohumol, silymarin, curcumin, and quercetin, triterpenes such as betulin, in addition to alkaloids as berberine, and fatty acids like oleic acid and α -linolenic acid were found to downregulate SREBP-1 and SERBP-2. Taxifolin, piperine, and ellagic acid inhibited MTP gene expression. Natural products regulating triglycerides and cholesterol-related gene expression could be key lead drugs to new medicines used to treat hyperlipidemia, hypercholesterolemia, liver cancer, breast cancer, and metabolic disorders.

Keywords: cytotoxicity; gene expression; herbal medicines; hyperlipidemia; MTP; SREBP.

Introduction

Dyslipidemia is a collective term used to describe abnormal levels of triglycerides, cholesterol, and/or high-density lipoproteins. It is characterized by a complex interplay of various lipid components, including elevated triglycerides (TG) and TG-rich lipoproteins, along with increased postprandial TG levels [1]. The imbalance is made worse by lower levels of Apolipoprotein A-I (Apo A-I) and high-density lipoprotein cholesterol (HDL-C) [2]. Dyslipidemia is marked by an increase in Apolipoprotein B (Apo B), reflecting higher levels of atherogenic lipoproteins [3]. Notably, there is an elevation in low-density lipoprotein cholesterol (LDL-C) particles number, especially the small dense LDL particles, which raises the risk of cardiovascular disease [4]. It was found that individuals with type 2 diabetes and metabolic syndrome commonly have dyslipidemia [5].

Dyslipidemia can be mediated by genetic factors, diet (such as unhealthy and ultraprocessed foods), lifestyle (such as sedentary lifestyle and smoking), and certain types of drugs [6]. Dyslipidemia can be a primary disease condition, or it can be secondary to certain disease conditions such as diabetes mellitus. Dyslipidemia contributes as a risk factor in fatal cardiovascular complications leading to the increase in risk of mortality [6–9].

Moreover, dyslipidemia in diabetic individuals is characterized by an increase in oxidized and glycated lipids, emphasizing the role of oxidative stress and glycation in lipid metabolism abnormalities associated with type 2 diabetes and metabolic syndrome [10].

Excessive accumulation of liver lipids contributes to inflammation and lipid metabolism disorders, causing non-alcoholic fatty liver disease (NAFLD). This condition is considered as a major cause of hepatic cancer [11]. Therefore, controlling lipid levels would be a promising strategy for combating NAFLD and liver cancer.

Currently, achieving an effective control of dyslipidemia is usually carried out with multi-drug treatment, preferably accompanied with lifestyle and dietary adjustments and restrictions. However, patients may have low compliance in following lifestyle changes [12]. In addition, long-term use of lipid-lowering medications can cause serious side effects. Therefore, finding alternatives from natural source and thus fewer side effects can be of great merit [7, 9, 13].

Nature serves as a significant and ongoing source of inspiration for researchers and scientists due to its abundance of bioactive compounds [14]. Natural products have long been utilized in traditional and modern medicine due to their diverse pharmacological properties, which include antimicrobial, anti-inflammatory, antioxidant, antidiabetic, and anticancer,

*Corresponding author e-mail: miral.om.mohamed@std.pharma.cu.edu.eg; (Miral O. Sabry).

Received date 23 May 2024; Revised date 26 June 2024; Accepted date 07 July 2024

DOI: 10.21608/ejchem.2024.292003.9756

©2025 National Information and Documentation Center (NIDOC)

antihypertensive, activities, where the therapeutic efficacy of natural products is attributed to their complex structures and the specific mechanisms through which they interact with biological targets [15–22].

Transcription factors are proteins involved in the process of converting, or transcribing, DNA into RNA [23]. Those concerned with lipid metabolism are the Sterol Regulatory Element-Binding Proteins (SREBPs) and the Microsomal Triglyceride Transfer Proteins (MTP) [24, 25]. Several phenolic compounds were found to be effective in regulating lipid metabolism. For example flavonoids can target lipid metabolism by inhibiting sterol regulatory element-binding protein-1 (SREBP-1), leading to inhibition of fatty acid synthase and associated enzymes [11], specially rutin, which was found to downregulate the transcription of SREBP-1c in hepatic cell line, thus decreasing the levels of triglycerides and cholesterol. Taxifolin, as well, downregulated the transcription of SREBP-1c in mice fed with high fat-diet [11].

SREBPs are a class of proteins that were initially identified by Brown and Goldstein nearly thirty years ago. These proteins regulate the transcriptional level of body lipids, including triglycerides and cholesterol. According to Wen et al., (2018), SREBPs are the transcription factors that control the expression of the genes that govern the synthesis of fatty acids, cholesterol, and triglycerides [30]. Three isoforms of SREBPs—SREBP-1a, SREBP-1c, and SREBP-2—are recognized and have been found to regulate different pathways that slightly overlap [24, 30, 31].

The regulatory protein involved in the synthesis of cholesterol and fatty acids is SREBP-1a, and that involved in the synthesis of fatty acids is SREBP-1c. The gene encoding SREBP-1 is located on chromosome 17 [30]. The SREBP-1 gene uses a different transcription start site to encode each of SREBP-1a and SREBP-1c [30]. Following gene transcription, their mRNA is translated into the inactive SREBP-1 protein, which *via* regulated cellular transport, is activated at the Golgi apparatus through proteolysis. The active SREBP-1 protein finally moves into the cell nucleus, where it activates the fatty acid biosynthesis by upregulating the genetic expression of the involved enzymes, such as acetyl-CoA carboxylase (ACC) and fatty acid synthase (FAS). When the blood levels of fatty acids and triglycerides reach normal healthy levels, SREBP-1 is suppressed through end-product feedback inhibition. At the same time, the SREBP transcriptional activity is depressed. However, this step fails to occur in hyperlipidemic individuals [29, 32].

A similar process takes place in regulating cholesterol biosynthesis in the body. The involved protein is SREBP-2, which is carried on chromosome 22. SREBP-2 controls cholesterol synthesis in insulin-responsive tissues such as the liver. In case of cellular demand for cholesterol, or when the blood levels of glucose or insulin increase, SREBP-2 is activated through proteolysis at the Golgi apparatus, resulting in activation of the cholesterol biosynthesis pathway including all its enzymes. When cholesterol blood levels reach normal healthy levels, SREBP-2 is suppressed through end-product feedback inhibition, and at the same time, the SREBP transcriptional activity is depressed. However, this step fails to occur in hypercholesterolemic individuals [29, 32, 33].

On the other hand, MTP protein is responsible for activation of the assembly of VLDL (Very Low-Density Lipoprotein) and synthesis of chylomicrons in the liver and intestine [34]. In addition, VLDL is the lipoprotein responsible for the transport of lipids from the liver to body tissues, thus raising lipid levels in the body, while chylomicrons are responsible for the absorption of dietary lipids in the small intestine. Therefore, downregulating the gene expression of MTP protein is expected to decrease the lipid levels in blood [35–37].

MTP is mainly produced in hepatocytes and enterocytes, and inhibition of its production was found to decrease the secretion and synthesis of VLDL in the liver and to reduce the plasma levels of triglycerides by decreasing the fat absorbed through chylomicrons [36, 38].

The term hyperlipidemia designates a group of medical conditions in which blood levels of triglycerides and cholesterol are increased above normal levels. This condition was found to be directly linked to fatal heart and blood vessel diseases [28]. In addition, high blood levels of triglycerides and cholesterol were recorded in certain types of cancer, such as hepatic and breast cancers. Besides, its pathogenesis was found to be associated with upregulated SREBP-1, SREBP-2, and MTP genetic expression [30, 35, 39, 40].

Although statins are currently considered a cornerstone in treating dyslipidemia because they inhibit the essential enzyme HMG-CoA reductase which is involved in the cholesterol biosynthesis pathway, their reputed drawbacks would preferably be avoided [36]. In addition to causing myopathy, rhabdomyolysis, and liver damage [28, 41, 42], statins inhibit HMG-CoA reductase, the rate-limiting enzyme in the cholesterol biosynthesis pathway, leading to a decrease in intracellular cholesterol levels. As a compensatory response, this inhibition triggers upregulation of the entire cholesterol biosynthetic pathway, including an increase in the expression of HMG-CoA reductase and other enzymes involved in the pathway [43]. Optimal treatment with statins still leaves a 60-80% residual cardiovascular risk [36, 44]. This residual risk is attributed to various factors, including persistent LDL cholesterol levels, elevated triglycerides, low HDL cholesterol, and other non-lipid-related risk factors, requiring further strategies such as combination therapies and lifestyle modifications to address this residual risk [44, 45]. Therefore, searching for natural products that can treat dyslipidemia by downregulating the gene expression of SREBPs and MTP, and thus suppressing the biosynthesis and absorption of lipids in the body from the very beginning, would be preferable.

Other than just hyperlipidemia, SREBPs were found to contribute to the pathogenesis of several diseases and disorders. These include obesity, cancer, non-alcoholic fatty liver, diabetes mellitus, atherosclerosis, chronic kidney, and neurodegenerative diseases [40, 46, 47]. SREBPs contribute to these diseases through several pathways that generate reactive oxygen species (ROS), increase endoplasmic reticulum stress, and cause inflammation, autophagy, and apoptosis [29, 48]. Among natural products, polyphenols have been extensively studied for their antioxidant properties [49, 50]. For instance, the polyphenol resveratrol is known to specifically reduce ROS generation by upregulating antioxidant enzymes such as superoxide dismutase and catalase, and by directly scavenging free radicals [51–53]. This action helps to mitigate oxidative stress and inflammation, which are critical factors in the pathogenesis of cardiovascular and metabolic diseases [54, 55].

Targeting SREBPs and MTP gene expression might be a promising approach in treating liver and breast cancers [56]. Cancer

cells satisfy their high lipid and cholesterol needs by increasing the uptake of dietary lipids as well as their endogenous synthesis [57]. Increased de-novo lipid biosynthesis is a necessary hallmark of cancer progression and metastasis. Part of this effect was found to be due to the increased expression of SREBP-1 [58].

Targeting De-novo lipid biosynthesis could be considered as a promising strategy in combating cancer since normal tissues depend on circulating lipids; meanwhile, cancer cells depend on de-novo synthesized lipids [58]. Blood levels of lipids are directly associated with hepatic and breast cancers. Several studies have proved that increased SREBPs activation is associated with cancer development [39, 40], tumor metastasis, as well as poor prognosis in breast cancer [30]. Moreover, in various types of cancers, it was found that the expression of SREBPs target genes are elevated above normal levels [59, 60].

Likewise, it was observed that down-regulating the gene expression of SREBPs altered the metabolic pathways in cancer cells *in-vitro* [30]. In fact, downregulating the expression of MTP protein in obese mice was found to treat dyslipidemia and to shrink tumor volume by 50% *via* alteration of circulating lipids [61]. The mevalonate pathway, through which lipids are synthesized and controlled by SREBPs, was upregulated in hepatic and breast cancers. A possible explanation was the mutations in sterol-regulatory related genes such as SREBPs [29].

AMPK (AMP-activated protein kinase) is a cellular energy sensor that becomes activated in response to low energy conditions [62]. Once activated, AMPK exerts profound effects on lipid and cholesterol metabolism [63]. AMPK activation inhibits SREBPs, particularly SREBP-1c, which reduces the production of fatty acids and cholesterol precursors in the liver and other tissues [64]. Many natural products, such as glabridin from licorice root, polyphenols from black nightshade, and components from okra, have been shown to activate AMPK [65–67].

The mechanism of action of natural products downregulating SREBP-1, SREBP-2, and MTP gene expression is represented in **figure 1**.

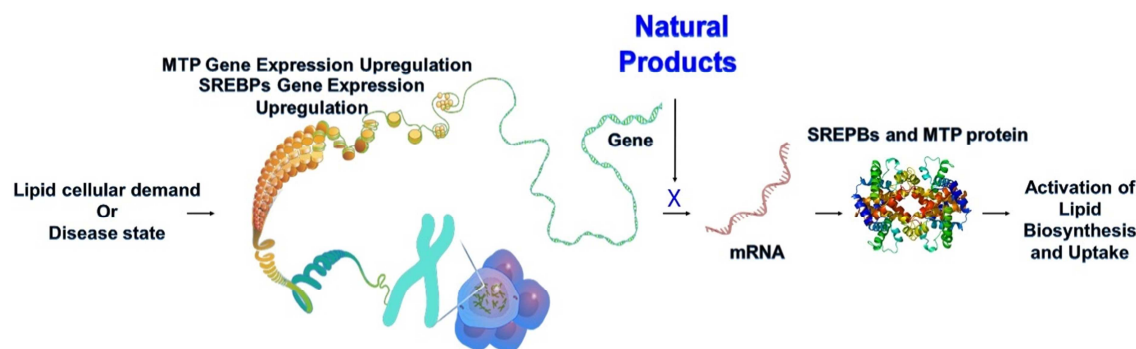


Figure 1: Mechanism of action of natural products downregulating SREBP-1, SREBP-2, and MTP gene expression.

The following survey intends to shed light on the nature of plant metabolites downregulating SREBP-1, SREBP-2, and MTP gene expression, their respective sources, and mechanisms of action to facilitate further incorporation in pharmaceutical formulations. Structural formulae of natural products isolated and/or identified in plants and proved to exert anti-hyperlipidemic and/or anti-hypercholesteremic activities are represented in **figure 2**. Recently, these compounds were found to act through downregulating SREBPs and/or MTP gene expression. It can be observed that the majority of the mentioned examples are mainly of phenolic or terpenoid nature.

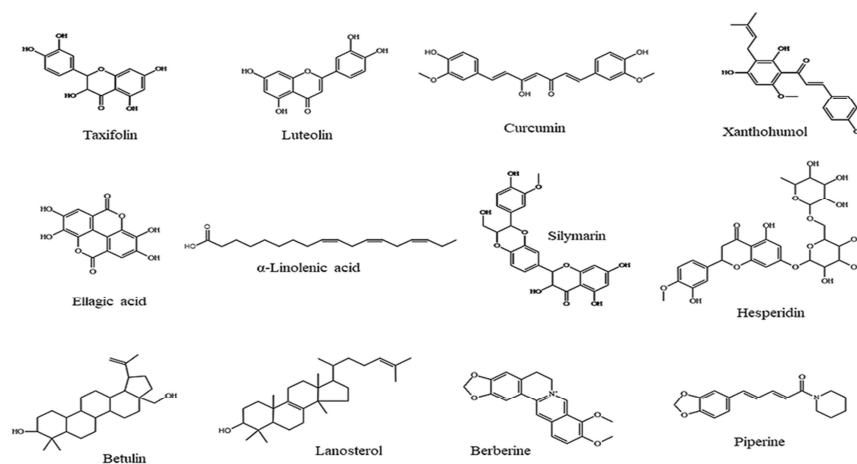


Figure 2: Representative examples of natural products downregulating SREBP-1, SREBP-2 and MTP gene expression.

1. Phenolic Compounds and Flavonoids

1.1. Quercetin flavonoid

Quercetin is a flavonoid widely distributed in plants. It belongs to the flavonols subfamily and acts as a pigment and is present in black tea, onions, citrus fruits, grapes, berries, cherries, apples, buckwheat, kale, tomatoes, and broccoli [68–71]. It is well recognized to possess anti-inflammatory, antioxidant, and neuroprotective potentials and is capable of protecting against aging-related diseases [71–74]. Quercetin reduces plasma LDL levels and prevents LDL oxidation, contributing to a decreased risk of cardiovascular diseases [71]. According to recent studies, quercetin quickly reduced the synthesis of fatty acids and cholesterol. Additionally, these investigations indicated that quercetin lowered the expression of the SREBP-1 and SREBP-2 genes [75–77].

1.2. Citrus fruits flavonoids

For instance, hesperidin (hesperitin-7-rutinoside), a flavanone glycoside commonly present in citrus fruits (*Citrus* L. species, family Rutaceae) [78], successfully prevented the buildup of fat in mice receiving a high-fat diet, and in HepG2 cell lines exposed to oleic acid, besides exerting several pharmacological activities including anti-inflammatory and antioxidant [79–81]. In a clinical study [80], hesperidin treatment improved lipid and glucose metabolisms in non-alcoholic fatty liver patients besides reducing inflammation and liver steatosis. Furthermore, Chen et al. (2022) discovered that hesperidin reduced the accumulation of lipids in the liver through a number of mechanisms, including downregulation of SREBP-1C expression [80].

Naringenin, another citrus fruits flavanone, was found to inhibit the secretion of Apolipoprotein B100 by downregulating the gene expression of microsomal triglyceride transfer protein (MTP) in HepG2 cells *via* a mechanism similar to that of insulin [82].

1.3. Celery and parsley flavonoids

Luteolin (3',4',5,7-tetrahydroxyflavone), a flavonoid present in celery and parsley (*Petroselinum crispum* Mill. and *Apium graveolens* L., family Apiaceae), was found effective against many types of tumors. These include stomach, prostate, and lung cancers [83]. Luteolin significantly downregulated SREBP-2 and SREBP-1 expression and subsequently lowered fatty acid and cholesterol levels in the blood [83, 84].

1.4. Hops prenylated flavonoids

Hops (*Humulus lupulus* L., family Cannabinaceae) is a renowned anti-hyperlipidemic. The major constituent of its fruit is the prenylated flavonoid xanthohumol that exerts anti-oxidative, chemopreventive, and anti-inflammatory actions. A recent study [85] revealed that xanthohumol reduced the gene expression of SREBP-1 in mice liver, besides suppressing the development of obesity. Moreover, xanthohumol prevented the activation of SREBP by blocking its endoplasmic reticulum to Golgi apparatus transportation. Likewise, it was found to reduce cholesterol and triglyceride levels, thus preventing obesity, hepatic steatosis, and atherosclerotic plaque development [29, 85, 86].

1.5. Tamarind and pine flavonoids

The flavonoid taxifolin present in tamarind (*Tamarindus indica* L, family Fabaceae) and Pine (*Pinus* species Lindl., family Pinaceae) was found to downregulate the gene expression of MTP protein [87, 88]. Furthermore, tamarind fruit pulp reduced total cholesterol and LDL-C levels in human subjects to a significant extent [89] while pine bark extract reduced LDL-C and increased HDL-C in human subjects [90].

1.6. Licorice root flavonoids

Glabridin is one of the prenylated isoflavanes extracted from the roots of licorice (*Glycyrrhiza glabra* L., family Fabaceae). Glabridin was claimed to exert numerous bioactivities, including antitumor, cardiovascular and hepatic protection, anti-obesity, and anti-diabetes [91]. In fact, the glabridin-rich acetone extract of licorice inhibited adipogenesis in 3T3-L1 adipocytes *in-vitro*, in addition to downregulating the expression of the lipogenic genes, stearoyl-CoA desaturase, fatty acid synthase, and SREBP-1 [92]. Moreover, in obese mice receiving a high-fat diet, AMPK (AMP-activated protein kinase) was activated by glabridin, and thus suppressing the expression of SREBP-1c [65].

1.7. Milk thistle fruit flavonolignans

Silymarin, the active component of dried milk thistle (*Silybum marianum* L., family Asteraceae) fruits extract consisting of a mixture of flavonolignans is reputed for its well-established hepatoprotective, antioxidant, antifibrotic, and anti-inflammatory properties besides being safe and clinically well tolerated. Recently, silymarin was found to suppress the expression of SREBP-1c mRNA [93–95].

1.8. Red soybeans isoflavones and amino acids

Several studies demonstrated the efficacy of soy beans protein in decreasing serum lipid levels [96]. The consumption of red soybean (*Glycine* species Willd., family Fabaceae) by mice fed a high-fat diet was found to decrease the expression of SREBP-1, this was referred to its isoflavone and amino acid contents [97].

1.9. Lizard's tail lignans

Sauchinone, a lignan present in lizard's tail (*Saururus chinensis* (Lour.) Baill., family Saururaceae), is well-known for its antioxidant, hepatoprotective, anti-inflammatory, and anti-steatosis properties [98, 99]. Both *in-vivo* and *in-vitro*, it was found to inhibit SREBP-1c through activation of AMPK (AMP-activated protein kinase) and inhibition of liver-X-receptor- α [100]. Another study proved that sauchinone is capable of downregulating both SREBP-1 and SREBP-2 in HepG2 cells [99].

1.10. Turmeric herb polyphenolics

Curcumin, the main active polyphenolic compound of turmeric (*Curcuma longa* L., family Zingiberaceae), has demonstrated a wide range of biological and pharmacological activities, including antimicrobial, anti-inflammatory, anticancer, anti-diabetic, anti-malarial, antiprotozoal, and antioxidant effects, making it a promising scaffold for drug design and development despite its low bioavailability challenges [101]. It was also found to promote weight loss in clinical trials and to reduce the incidence of obesity-related metabolic disorders [102]. Published data supported that, among natural products, curcumin is reported to significantly inhibit SREBPs signaling, and, consequently to manage dyslipidemia and decrease lipid accumulation through downregulation of SREBPs [103].

1.11. Mulberry leaf phenolics

Mulberry (*Morus* species L., family Moraceae) leaf aqueous extract, a traditional Chinese remedy for treatment of obesity, is rich in phenolics and flavonoids. A dose-dependent decrease was observed in the level of triglycerides, preventing aggregation of lipid globules and downregulating the gene expression of SREBP-1c [104].

1.12. Black nightshade whole plant polyphenols

Black nightshade (*Solanum nigrum* L., family Solanaceae) is known to be rich in polyphenols [66]. The aqueous extract of the whole plant was reported to exhibit lipid-lowering activity, reducing blood triglycerides and cholesterol levels in mice *via* downregulation of SREBP gene expression through AMPK-dependent phosphorylation of SREBP [66, 105, 106].

1.13. Oak bark and Pomegranate rind polyphenols

Ellagic acid, a polyphenolic constituent of oak bark (*Quercus* species L., family Fagaceae) and pomegranate (*Punica granatum* L., family Lythraceae), was found to regulate the metabolism of cholesterol in hepatocytes through down-regulation of MTP mRNA [107]. Ellagic acid supplementation in mice enhances hepatic lipid metabolism and antioxidant capacity, improving cholesterol profiles and increasing specific enzyme abundances to promote liver health [108].

1.14. Red grapes polyphenolic stilbenoids

Resveratrol is a polyphenolic stilbene present in peanuts (*Arachis hypogaea* L., family Fabaceae) and red grapes (genus *Vitis* L., family Vitaceae) [109]. Studies revealed that resveratrol prevents fat accumulation *in-vivo* in the liver of mice receiving a high-fat diet and *in-vitro* in HepG2 cells [109]. In addition, it exerts anti-neoplastic effects against different types of cancers [110]. Besides resveratrol was claimed to downregulate the gene expression of SREBP-1 and to inhibit its activity in HepG2 cells. This was suggested to be mediated by the Sirt1-FOXO1 (Sirtuin 1-Forkhead box protein O1) signaling pathway [109].

Besides, resveratrol was found effective against xenograft oral cancer by significantly reducing lipogenesis, mediated through downregulation of the gene expression of SREBP-1 and epidermal fatty acid-binding protein (E-FABP). In addition, it initiated autophagy in oral cancer cells, and this was suggested to be through inhibition of SREBP-1-mediated cell survival signaling [110]. Moreover, resveratrol was found to sensitize pancreatic cancer cells to the anticancer drug gemcitabine through inhibition of SREBP [111].

Many strategies have been studied to enhance the bioavailability of resveratrol. For instance, resveratrol-based coumarins, synthesized and characterized for their enhanced biomedical effects, showed improvements in pharmacokinetic properties and various therapeutic activities compared to resveratrol alone [112].

2. Triterpenoids

2.1. Birch bark lupane triterpenoids

Betulin is a lupane triterpenoid found in Birch bark (*Betula* species L., family Betulaceae). It downregulates cholesterol and fatty acid synthesis and reduces blood lipid levels while increasing insulin sensitivity and inhibiting SREBP maturation [29, 113].

2.2. Chaga lanostane triterpenoids

Chaga (*Inonotus obliquus* Pilát, family Hymenochaetaceae), a mushroom traditionally used in Chinese Medicine to treat various gastrointestinal diseases, was found to possess significant anticancer, anti-inflammatory and hypoglycemic activities [114]. Its active constituents *viz.* the lanostane triterpenoids inotodiol, lanosterol, and trametenolic acid, significantly decreased hepatic lipid accumulation through the downregulation of SREBP-1c [114].

2.3 Okra fruits triterpenoids

The alcohol extract of okra (*Abelmoschus esculentus* L., family Malvaceae) fruit, rich in quercetin glucosides and pentacyclic triterpene ester, was found to be effective in stimulating AMP-activated protein kinase (AMPK) leading to phosphorylation of SREBP-1c and consecutively its downregulation [67, 106]. Its water extract, rich in polysaccharides, was relatively more stable in decreasing the expression of SREBP-1c [115].

3. Miscellaneous Compounds

3.1. Chinese goldthread alkaloids

Berberine is an isoquinoline alkaloid isolated from several plants of family Berberidaceae, especially *Coptis* species. Among these, *Coptis chinensis* F. (Chinese goldthread) rhizome is considered one of the main sources of berberine [116]. Berberine was found to be effective in treating a variety of cancers, including liver, colon, breast, and lung cancers. Moreover, its lipid-lowering, immunomodulatory, antioxidative, cardioprotective, hepatoprotective, and renoprotective activities were reported [116, 117]. Besides, a clinical trial demonstrated that berberine can be used in the treatment of dyslipidemia alone or together with simvastatin [116]. Recently, berberine was proven to inhibit SREBP-1 activation and expression of its transporting protein SREBP and cleavage-activating protein (SCAP) [118, 119].

3.2. Black and white pepper alkaloids

Piperine is the major pungent alkaloid in *Piper nigrum* L., family Piperaceae. It occurs in both black and white pepper and was found to lower plasma cholesterol levels through several mechanisms, including downregulation of the gene expression of intestinal MTP [120].

3.3. Unsaturated fatty acids

Unsaturated fatty acids such as α -Linolenic acid (ALA) were found to downregulate SREBP-2, SREBP-1a, and SREBP-1c expression [121–124]. Moreover, PUFAs and oleic acid were reported to inhibit the proteolysis of SREBP-1 and to downregulate its expression. Thus, they could help in the treatment of hyperlipidemia and other metabolic diseases, taking into consideration that SREBP-1 activation was found to produce lipotoxicity, which contributes to several lipidemic disorders [29, 122, 123, 125].

3.4. Citrus pectin

Pectin is a complex and soluble polysaccharide found in the cell walls of citrus fruits [126]. Pectin has long been recognized for its impact on lipid metabolism, proven by a clinical trial conducted in 1977 which involved administering 40–50 g/day of pectin for two weeks to nine normolipidemic and hyperlipidemic patients. The results demonstrated that while pectin supplementation significantly decreased serum total and unesterified cholesterol levels in hypercholesterolemic subjects [127]. Recently, Wang *et al.* (2022) reported that citrus pectin lowered cholesterol and triglyceride levels in HepG2 cells through downregulation of fatty acid synthetase and SREBP-1c [128].

3.5. Garlic bulbs

Garlic (*Allium sativum* L., family Amyrallidaceae) is known to possess cardiovascular benefits and anti-hyperlipidemic activity [129]. Many human studies have proven that garlic can reduce the blood levels of total cholesterol and LDL-C in patients with hypercholesterolemia [130–132]. Garlic extract was recently reported to reduce the gene expression of SREBP-1c, leading to a decrease in its target genes [105].

3.6. Chinese toad bufadienolides

Cinobufotalin is one of the bufadienolides extracted from the venomous skin secretions of Chinese toads (*Bufo bufo gargarizans* Cantor, family Bufonidae). The dried skin secretion of *Bufo bufo gargarizans* has been used in Traditional Chinese Medicine in treatment of several types of cancers [133]. Many studies reported the powerful anticancer activity of cinobufotalin against hepatocellular carcinoma, human lymphoma, lung cancer, and colon adenocarcinoma through numerous mechanisms [134–138]. Among the various mechanisms cinobufotalin targeted cancer through inhibiting lipogenesis in hepatocellular carcinoma specifically by inhibiting SREBP-1 expression [136].

3.7. Fenugreek steroidal saponin

Fenugreek (*Trigonella foenum-graecum* L., family Fabaceae) is known to decrease serum triglycerides and cholesterol, as well as hepatic lipids [139–141]. Diosgenin is a saponin phytosterol effective against lipid metabolism disorders and is extracted from fenugreek [142–144]. When mice on a high-fat diet were given diosgenin, their weight gain was considerably reduced and their lipid profile was improved. Furthermore, diosgenin suppressed their two-fold increase in the gene expression of SREBP-1c and its downstream gene, fatty acid synthase [142]. *In-vitro*, diosgenin inhibited the accumulation of triglycerides and downregulated the lipogenic genes' expression, including SREBP-1c, in HepG2 cells. This activity was through inhibiting the transactivation of liver-X-receptor- α , which is the transcription factor responsible for stimulating the gene expression of SREBP-1c [140, 145].

Applications and Prospects

Microsomal triglyceride transfer protein (MTP) regulates the absorption and transportation of lipids by playing a role in the assembly and secretion of Apolipoproteins as chylomicrons and VLDL in the intestine and liver [146]. A few MTP inhibitors have been identified, and they were proven to reduce the levels of plasma lipids. However, inhibiting MTP resulted in accumulation of lipids in the liver, a significant drawback that stopped previously identified MTP inhibitors from reaching clinical use [88]. For instance, Lomitapide is known to inhibit MTP activity and reduce lipid levels. However, its use is limited due to hepatotoxicity [147, 148]. Therefore, it appears essential to search for intestine-specific MTP inhibitors or to find lead drugs capable of regulating the expression of MTP instead of inhibiting it, to serve as starting points for further

optimization and development into potential therapeutic agents [146]. This mechanism offers a safer and potentially more effective strategy for managing hyperlipidemia. Therefore, natural products that downregulate the gene expression of MTP instead of inhibiting its activity can be considered as a better approach to lower plasma lipids.

Using SREBP inhibitors in combination with fatty acid synthase (FAS) inhibitors in the treatment of cancer might be considered as a promising strategy in the treatment of primary cutaneous T-cell lymphomas (CTCL) [149]. FAS, the enzyme that catalyzes the biosynthesis of fatty acids, is highly expressed in CTCL and contributes to oncogenicity in other malignancies. However, inhibition of FAS was found to increase FAS expression due to feedback inhibition. Adding an SREBP inhibitor partially reduced the upregulation of FAS caused by FAS inhibitors. This is explained by the role of SREBP in upregulating FAS gene expression [149].

Recently, a novel stilbene resveratrol derivative (BF175) containing boron and two chlorine groups was synthesized. Adding boron to small molecules enhanced their binding to target molecules, such as proteins, DNA or RNA. BF175 inhibited the target gene expression of SREBP, while its non-chlorine-containing analogue showed no effects. BF175 repressed the expression of SREBP-1c only in HepG2 cells while decreasing the transcription of both SREBP-1 and SREBP-2 genes *in-vivo* in mice liver [150]. Since Natural polyphenolics, such as resveratrol, are hindered by their relatively low bioavailability, using a similar strategy for chemical modifications can be developed for the improvement of their bioavailability.

2. Conclusions

In conclusion, sterol regulatory element-binding proteins (SREBPs) and microsomal triglyceride transfer protein (MTP) are responsible for regulating the process of endogenous lipids synthesis and the uptake of exogenous lipids. However, this process is disrupted in many metabolic diseases such as dyslipidemia, obesity, cancer, non-alcoholic fatty liver, diabetes mellitus, atherosclerosis, chronic kidney diseases, and neurodegenerative diseases. Targeting the gene expression of these proteins is a promising strategy for the treatment of many metabolic diseases. Strikingly, a large number of natural products downregulate the gene expression of SREBPs and MTP proteins, and thus could be considered as potential therapeutic agents in the treatment of hyperlipidemic conditions.

Natural products reported to downregulate SREBPs are mainly of polyphenolic and terpenoid nature, including hesperidin, luteolin, quercetin, xanthohumol, curcumin, silymarin, and ellagic acid. Besides, the alkaloids berberine and piperine, citrus pectin, chaga, mulberry, black nightshade, garlic, red soya bean, and okra extracts were also active. Meanwhile, gene expression of MTP protein was reported to be downregulated by taxifolin, piperine, and ellagic acid.

3. Conflicts of interest

There are no conflicts to declare.

4. References

- Borén J, Matikainen N, Adiels M, Taskinen M-R (2014) Postprandial hypertriglyceridemia as a coronary risk factor. *Clinica Chimica Acta* 431:131–142. <https://doi.org/10.1016/j.cca.2014.01.015>
- Ng DS (2013) Diabetic Dyslipidemia: From Evolving Pathophysiological Insight to Emerging Therapeutic Targets. *Canadian Journal of Diabetes* 37:319–326. <https://doi.org/10.1016/j.cjcd.2013.07.062>
- Manjunath CN, Rawal JR, Irani PM, Madhu K (2013) Atherogenic dyslipidemia. *Indian Journal of Endocrinology and Metabolism* 17:969. <https://doi.org/10.4103/2230-8210.122600>
- Duran EK, Aday AW, Cook NR, Buring JE, Ridker PM, Pradhan AD (2020) Triglyceride-Rich Lipoprotein Cholesterol, Small Dense LDL Cholesterol, and Incident Cardiovascular Disease. *Journal of the American College of Cardiology* 75:2122–2135. <https://doi.org/10.1016/j.jacc.2020.02.059>
- Kopin L, Lowenstein C (2017) Dyslipidemia. *Annals of Internal Medicine*
- Santos HO, Bueno AA, Mota JF (2018) The effect of artichoke on lipid profile: A review of possible mechanisms of action. *Pharmacological Research* 137:170–178. <https://doi.org/10.1016/j.phrs.2018.10.007>
- He N, Ye H (2020) Exercise and Hyperlipidemia. In: Xiao J (ed) *Physical Exercise for Human Health*. Springer, Singapore, pp 79–90
- Nikpayam O, Faghfour AH, Tavakoli-Rouzbehani OM, Jalali S-M, Najafi M, Sohrab G (2020) The effect of green coffee extract supplementation on lipid profile: A systematic review of clinical trial and in-vivo studies. *Diabetes & Metabolic Syndrome: Clinical Research & Reviews* 14:1521–1528. <https://doi.org/10.1016/j.dsx.2020.07.043>
- Shahinfar H, Bazshahi E, Amini MR, Payandeh N, Pourreza S, Noruzi Z, Shab-Bidar S (2021) Effects of artichoke leaf extract supplementation or artichoke juice consumption on lipid profile: A systematic review and dose–response meta-analysis of randomized controlled trials. *Phytotherapy Research* 35:6607–6623. <https://doi.org/10.1002/ptr.7247>
- Yadav MK, Kumar P, Sharma P, Mohapatra TK (2020) Evaluation of Dyslipidemia and Oxidative Stress in Type II Diabetes Patients. *Journal of Datta Meghe Institute of Medical Sciences University* 15:448. https://doi.org/10.4103/jdmimsu.jdmimsu_366_20
- Li L, Qin Y, Xin X, Wang S, Liu Z, Feng X (2023) The great potential of flavonoids as candidate drugs for NAFLD. *Biomedicine & Pharmacotherapy* 164:114991. <https://doi.org/10.1016/j.biopha.2023.114991>
- Cicero AFG, Fogacci F, Stoian AP, Vrablik M, Al Rasadi K, Banach M, Toth PP, Rizzo M (2021) Nutraceuticals in the Management of Dyslipidemia: Which, When, and for Whom? Could Nutraceuticals Help Low-Risk Individuals with Non-optimal Lipid Levels? *Curr Atheroscler Rep* 23:57. <https://doi.org/10.1007/s11883-021-00955-y>

13. Hussein WA, Salem AA-E, Fahmy HA, Mounair SM, Soliman AS, Abbas MS (2022) Effect of Carob, Doum, and Cinnamon Powder on Blood Lipid Profile in Diabetic Rats. *Egyptian Journal of Chemistry* 65:317–328. <https://doi.org/10.21608/ejchem.2022.114446.5202>
14. Jebir RM, Mustafa YF (2022) Watermelon Allsweet: A Promising Natural Source of Bioactive Products. *Journal of Medicinal and Chemical Sciences* 5:652–666. <https://doi.org/10.26655/JMCHEMSCI.2022.5.1>
15. Khalil RR, Mohammed ET, Mustafa YF (2022) Evaluation of In vitro Antioxidant and Antidiabetic Properties of Cydonia Oblonga Seeds' Extracts. *Journal of Medicinal and Chemical Sciences* 5:1048–1058. <https://doi.org/10.26655/JMCHEMSCI.2022.6.18>
16. Mustafa YF, Ismael RN, Jebir RM (2024) Natural coumarins from two cultivars of watermelon seeds as biosafe anticancer agents, an algorithm for their isolation and evaluation. *Journal of Molecular Structure* 1295:136644. <https://doi.org/10.1016/j.molstruc.2023.136644>
17. Al-Madhagy SomaiaA, Gad SS, Mostafa ES, Angeloni S, Saad MA, Sabry OM, Caprioli G, El-Hawary SS (2023) A new firewall in the fight against breast cancer: in-vitro and in-silico studies correlating chemistry to apoptotic activity of *Otostegia fruticosa*. *Natural Product Research* 37:2770–2775. <https://doi.org/10.1080/14786419.2022.2130306>
18. Khaled A, Ahmed E, Mamdouh M, Saad H, Mohamed A, Sobhy M, Piatti D, Sabry M, Saad MA, Sabry OM, Caprioli G (2023) Natural angiotensin converting enzyme inhibitors: A safeguard against hypertension, respiratory distress syndrome, and chronic kidney diseases. *Phytotherapy Research* 37:5464–5472. <https://doi.org/10.1002/ptr.7987>
19. Ahmed H, Abdelraheem A, Salem M, Sabry M, Fekry N, Mohamed F, Saber A, Piatti D, Sabry M, Sabry O, Caprioli G (2024) Suppression of breast cancer: modulation of estrogen receptor and downregulation of gene expression using natural products. *Natural Product Research* 38:1997–2006. <https://doi.org/10.1080/14786419.2023.2232926>
20. Abouelwafa E, Zaki A, M. Sabry O, Caprioli G, Abdel-Sattar E (2023) *Dolomiaea costus*: an untapped mine of sesquiterpene lactones with wide magnificent biological activities. *Natural Product Research* 37:4069–4079. <https://doi.org/10.1080/14786419.2022.2164577>
21. Eldin AB, Ezzat M, Afifi M, Sabry O, Caprioli G (2023) Herbal medicine: the magic way crouching microbial resistance. *Natural Product Research* 37:4280–4289. <https://doi.org/10.1080/14786419.2023.2172009>
22. Sabry MO, Sedek M, Issa MY, Elzalabani S (2024) Plants Effective in the Control of Hyperlipidemia and Hypercholesterolemia: A Review. *Egyptian Journal of Chemistry* 67:33–41. <https://doi.org/10.21608/ejchem.2023.227181.8364>
23. Lambert SA, Jolma A, Campitelli LF, Das PK, Yin Y, Albu M, Chen X, Taipale J, Hughes TR, Weirauch MT (2018) The Human Transcription Factors. *Cell* 172:650–665. <https://doi.org/10.1016/j.cell.2018.01.029>
24. Shimano H (2001) Sterol regulatory element-binding proteins (SREBPs): transcriptional regulators of lipid synthetic genes. *Prog Lipid Res* 40:439–452. [https://doi.org/10.1016/s0163-7827\(01\)00010-8](https://doi.org/10.1016/s0163-7827(01)00010-8)
25. Iqbal J, Jahangir Z, Al-Qarni AA (2020) Microsomal Triglyceride Transfer Protein: From Lipid Metabolism to Metabolic Diseases. In: Jiang X-C (ed) *Lipid Transfer in Lipoprotein Metabolism and Cardiovascular Disease*. Springer, Singapore, pp 37–52
26. Pe O, Ma F, Ao A Hyperlipidemia: Etiology and Possible Control. 8
27. Brown MS, Goldstein JL (1997) The SREBP Pathway: Regulation of Cholesterol Metabolism by Proteolysis of a Membrane-Bound Transcription Factor. *Cell* 89:331–340. [https://doi.org/10.1016/S0092-8674\(00\)80213-5](https://doi.org/10.1016/S0092-8674(00)80213-5)
28. Pe O, Cs A-O, Okorochoa A, Ao A (2015) Hyperlipidemia: Etiology and Possible Control. *IOSR Journal of Dental and Medical Sciences* 14:93–100. <https://doi.org/10.9790/0853-1410693100>
29. Shimano H, Sato R (2017) SREBP-regulated lipid metabolism: convergent physiology — divergent pathophysiology. *Nat Rev Endocrinol* 13:710–730. <https://doi.org/10.1038/nrendo.2017.91>
30. Wen Y-A, Xiong X, Zaytseva YY, Napier DL, Vallee E, Li AT, Wang C, Weiss HL, Evers BM, Gao T (2018) Downregulation of SREBP inhibits tumor growth and initiation by altering cellular metabolism in colon cancer. *Cell Death Dis* 9:265. <https://doi.org/10.1038/s41419-018-0330-6>
31. DeBose-Boyd RA, Ye J (2018) SREBPs in Lipid Metabolism, Insulin Signaling, and Beyond. *Trends in Biochemical Sciences* 43:358–368. <https://doi.org/10.1016/j.tibs.2018.01.005>
32. Hannah VC, Ou J, Luong A, Goldstein JL, Brown MS (2001) Unsaturated fatty acids down-regulate srebp isoforms 1a and 1c by two mechanisms in HEK-293 cells. *J Biol Chem* 276:4365–4372. <https://doi.org/10.1074/jbc.M007273200>
33. Madison BB (2016) Srebp2: A master regulator of sterol and fatty acid synthesis I. *Journal of Lipid Research* 57:333–335. <https://doi.org/10.1194/jlr.C066712>
34. Nielsen LB, Véniant M, Borén J, Raabe M, Wong JS, Tam C, Flynn L, Vanni-Reyes T, Gunn MD, Goldberg IJ, Hamilton RL, Young SG (1998) Genes for apolipoprotein B and microsomal triglyceride transfer protein are expressed in the heart: evidence that the heart has the capacity to synthesize and secrete lipoproteins. *Circulation* 98:13–16. <https://doi.org/10.1161/01.cir.98.1.13>
35. Hussain MM, Rava P, Walsh M, Rana M, Iqbal J (2012) Multiple functions of microsomal triglyceride transfer protein. *Nutrition & Metabolism* 9:14. <https://doi.org/10.1186/1743-7075-9-14>
36. Ahn CH, Choi SH (2015) New Drugs for Treating Dyslipidemia: Beyond Statins. *Diabetes Metab J* 39:87. <https://doi.org/10.4093/dmj.2015.39.2.87>
37. Prasomthong J, Limpeanchob N, Daodee S, Chonpathompikunlert P, Tunsophon S (2022) Hibiscus sabdariffa extract improves hepatic steatosis, partially through IRS-1/Akt and Nrf2 signaling pathways in rats fed a high fat diet. *Sci Rep* 12:7022. <https://doi.org/10.1038/s41598-022-11027-9>

38. Hu H, Tan L, Li X, Li J, Fan C, Huang F, Zhuo Z, Hou K, Xu Y, Wang Q, Yang Y, Cheng J (2022) Betaine Reduces Lipid Anabolism and Promotes Lipid Transport in Mice Fed a High-Fat Diet by Influencing Intestinal Protein Expression. *Foods* 11:2421. <https://doi.org/10.3390/foods11162421>
39. Lo AK-F, Lung RW-M, Dawson CW, Young LS, Ko C-W, Yeung WW, Kang W, To K-F, Lo K-W (2018) Activation of sterol regulatory element-binding protein 1 (SREBP1)-mediated lipogenesis by the Epstein-Barr virus-encoded latent membrane protein 1 (LMP1) promotes cell proliferation and progression of nasopharyngeal carcinoma. *The Journal of Pathology* 246:180–190. <https://doi.org/10.1002/path.5130>
40. Kanagasabai T, Li G, Shen TH, Gladoun N, Castillo-Martin M, Celada SI, Xie Y, Brown LK, Mark ZA, Ochieng J, Ballard BR, Cordon-Cardo C, Adunyah SE, Jin R, Matusik RJ, Chen Z (2022) MicroRNA-21 deficiency suppresses prostate cancer progression through downregulation of the IRS1-SREBP-1 signaling pathway. *Cancer Letters* 525:46–54. <https://doi.org/10.1016/j.canlet.2021.09.041>
41. Last AR, Ference JD, Falleroni J (2011) Pharmacologic Treatment of Hyperlipidemia. 84:8
42. Last AR (2017) Hyperlipidemia: Drugs for Cardiovascular Risk Reduction in Adults. 95:12
43. Istvan ES, Deisenhofer J (2001) Structural mechanism for statin inhibition of HMG-CoA reductase. *Science* 292:1160–1164. <https://doi.org/10.1126/science.1059344>
44. Lim S, Park YM, Sakuma I, Koh KK (2013) How to control residual cardiovascular risk despite statin treatment: Focusing on HDL-cholesterol. *International Journal of Cardiology* 166:8–14. <https://doi.org/10.1016/j.ijcard.2012.03.127>
45. Stone NJ, Robinson JG, Lichtenstein AH, Bairey Merz CN, Blum CB, Eckel RH, Goldberg AC, Gordon D, Levy D, Lloyd-Jones DM, McBride P, Schwartz JS, Shero ST, Smith SC, Watson K, Wilson PWF, American College of Cardiology/American Heart Association Task Force on Practice Guidelines (2014) 2013 ACC/AHA guideline on the treatment of blood cholesterol to reduce atherosclerotic cardiovascular risk in adults: a report of the American College of Cardiology/American Heart Association Task Force on Practice Guidelines. *J Am Coll Cardiol* 63:2889–2934. <https://doi.org/10.1016/j.jacc.2013.11.002>
46. Ma D-B, Liu X-Y, Jia H, Zhang Y, Jiang Q, Sun H, Li X, Sun F, Chai Y, Feng F, Liu L (2022) A Novel Small-Molecule Inhibitor of SREBP-1 Based on Natural Product Monomers Upregulates the Sensitivity of Lung Squamous Cell Carcinoma Cells to Antitumor Drugs. *Frontiers in Pharmacology* 13: <https://doi.org/10.3389/fphar.2022.895744>
47. Raof A, Fawzy G (2022) Anti-Obesity Potential of Natural Products. *Egyptian Journal of Chemistry* 65:329–358. <https://doi.org/10.21608/ejchem.2022.118996.5354>
48. Lhoták S, Sood S, Brimble E, Carlisle RE, Colgan SM, Mazzetti A, Dickhout JG, Ingram AJ, Austin RC (2012) ER stress contributes to renal proximal tubule injury by increasing SREBP-2-mediated lipid accumulation and apoptotic cell death. *Am J Physiol Renal Physiol* 303:F266–278. <https://doi.org/10.1152/ajprenal.00482.2011>
49. Scalbert A, Johnson IT, Saltmarsh M (2005) Polyphenols: antioxidants and beyond2. *The American Journal of Clinical Nutrition* 81:215S–217S. <https://doi.org/10.1093/ajcn/81.1.215S>
50. Abd-elfattah M, Maina N, Kareu PG, El-Shemy HA (2023) Antioxidant Potential of Eight selected Kenyan Medicinal plants. *Egyptian Journal of Chemistry* 66:545–553. <https://doi.org/10.21608/ejchem.2022.127658.5666>
51. Pervaiz S, Holme AL (2009) Resveratrol: Its Biologic Targets and Functional Activity. *Antioxidants & Redox Signaling* 11:2851–2897. <https://doi.org/10.1089/ars.2008.2412>
52. Konyalioglu S, Armagan G, Yalcin A, Atalayin C, Dageci T (2013) Effects of resveratrol on hydrogen peroxide-induced oxidative stress in embryonic neural stem cells. *Neural Regen Res* 8:485–495. <https://doi.org/10.3969/j.issn.1673-5374.2013.06.001>
53. Cosín-Tomás M, Senserrich J, Arumí-Planas M, Alquézar C, Pallàs M, Martín-Requero Á, Suñol C, Kaliman P, Sanfeliu C (2019) Role of Resveratrol and Selenium on Oxidative Stress and Expression of Antioxidant and Anti-Aging Genes in Immortalized Lymphocytes from Alzheimer's Disease Patients. *Nutrients* 11:1764. <https://doi.org/10.3390/nu11081764>
54. Xia N, Daiber A, Förstermann U, Li H (2017) Antioxidant effects of resveratrol in the cardiovascular system. *British Journal of Pharmacology* 174:1633–1646. <https://doi.org/10.1111/bph.13492>
55. Jomova K, Raptova R, Alomar SY, Alwasel SH, Nepovimova E, Kuca K, Valko M (2023) Reactive oxygen species, toxicity, oxidative stress, and antioxidants: chronic diseases and aging. *Arch Toxicol* 97:2499–2574. <https://doi.org/10.1007/s00204-023-03562-9>
56. Xiao X, Song B-L (2013) SREBP: a novel therapeutic target. *Acta Biochim Biophys Sin (Shanghai)* 45:2–10. <https://doi.org/10.1093/abbs/gms112>
57. Beloribi-Djefaflija S, Vasseur S, Guillaumond F (2016) Lipid metabolic reprogramming in cancer cells. *Oncogenesis* 5:e189–e189. <https://doi.org/10.1038/oncsis.2015.49>
58. Mounier C, Bouraoui L, Rassart E (2014) Lipogenesis in cancer progression (Review). *International Journal of Oncology* 45:485–492. <https://doi.org/10.3892/ijo.2014.2441>
59. Sun Y, He W, Luo M, Zhou Y, Chang G, Ren W, Wu K, Li X, Shen J, Zhao X, Hu Y (2015) SREBP1 regulates tumorigenesis and prognosis of pancreatic cancer through targeting lipid metabolism. *Tumor Biol* 36:4133–4141. <https://doi.org/10.1007/s13277-015-3047-5>
60. Lewis CA, Braut C, Peck B, Bensaad K, Griffiths B, Mitter R, Chakravarty P, East P, Dankworth B, Alibhai D, Harris AL, Schulze A (2015) SREBP maintains lipid biosynthesis and viability of cancer cells under lipid- and oxygen-deprived conditions and defines a gene signature associated with poor survival in glioblastoma multiforme. *Oncogene* 34:5128–5140. <https://doi.org/10.1038/onc.2014.439>
61. Hussain A, Lian J, Watts R, Gutiérrez T, Nelson R, Goping IS, Lehner R (2022) Attenuation of obesity-induced hyperlipidemia reduces tumor growth. *Biochimica et Biophysica Acta (BBA) - Molecular and Cell Biology of Lipids* 1867:159124. <https://doi.org/10.1016/j.bbalip.2022.159124>

62. Hardie DG, Ross FA, Hawley SA (2012) AMPK - a nutrient and energy sensor that maintains energy homeostasis. *Nat Rev Mol Cell Biol* 13:251–262. <https://doi.org/10.1038/nrm3311>
63. Viollet B, Horman S, Leclerc J, Lantier L, Foretz M, Billaud M, Giri S, Andreelli F (2010) AMPK inhibition in health and disease. *Crit Rev Biochem Mol Biol* 45:276–295. <https://doi.org/10.3109/10409238.2010.488215>
64. Moslehi A, Hamidi-zad Z (2018) Role of SREBPs in Liver Diseases: A Mini-review. *J Clin Transl Hepatol* 6:332–338. <https://doi.org/10.14218/JCTH.2017.00061>
65. Lee J-W, Choe SS, Jang H, Kim J, Jeong HW, Jo H, Jeong K-H, Tadi S, Park MG, Kwak TH, Kim JM, Hyun D-H, Kim JB (2012) AMPK activation with glabridin ameliorates adiposity and lipid dysregulation in obesity. *Journal of Lipid Research* 53:1277–1286. <https://doi.org/10.1194/jlr.M022897>
66. Chang J-J, Chung D-J, Lee Y-J, Wen B-H, Jao H-Y, Wang C-J (2017) Solanum nigrum Polyphenol Extracts Inhibit Hepatic Inflammation, Oxidative Stress, and Lipogenesis in High-Fat-Diet-Treated Mice. *J Agric Food Chem* 65:9255–9265. <https://doi.org/10.1021/acs.jafc.7b03578>
67. Nasrollahi Z, ShahaniPour K, Monajemi R, Ahadi AM (2022) Effect of quercetin and *Abelmoschus esculentus* (L.) Moench on lipids metabolism and blood glucose through AMPK- α in diabetic rats (HFD/STZ). *J Food Biochem* 46:e14506. <https://doi.org/10.1111/jfbc.14506>
68. Singh P, Arif Y, Bajguz A, Hayat S (2021) The role of quercetin in plants. *Plant Physiology and Biochemistry* 166:10–19. <https://doi.org/10.1016/j.plaphy.2021.05.023>
69. Salehi B, Machin L, Monzote L, Sharifi-Rad J, Ezzat SM, Salem MA, Merghany RM, El Mahdy NM, Kılıç CS, Sytar O, Sharifi-Rad M, Sharopov F, Martins N, Martorell M, Cho WC (2020) Therapeutic Potential of Quercetin: New Insights and Perspectives for Human Health. *ACS Omega* 5:11849–11872. <https://doi.org/10.1021/acsomega.0c01818>
70. Anand David AV, Arulmoli R, Parasuraman S (2016) Overviews of Biological Importance of Quercetin: A Bioactive Flavonoid. *Pharmacogn Rev* 10:84–89. <https://doi.org/10.4103/0973-7847.194044>
71. AL-shakarchi W, Abdulaziz NT, Mustafa YF (2022) A review of the chemical, pharmacokinetic, and pharmacological aspects of quercetin. *Eurasian Chemical Communications* 4:645–656. <https://doi.org/10.22034/ecc.2022.335451.1393>
72. Izzularab BM, Beltagy AM, Meged M, Tousson E, Beltagy DM (2022) Beneficial Roles of Quercetin Nanoparticles Extracted from *Euphorbia Helioscopia* Against Lung Damage in Mice. *Egyptian Journal of Chemistry* 65:1487–1494. <https://doi.org/10.21608/ejchem.2022.137018.6043>
73. Elraey M, Elgamal A, Gaara AH (2022) Quercetin -3-O- β -D-glucuronide butyl ester from *Vitis vinifera* leaves of potent anti-helicobacter pylori activity and impact of its combination with clarithromycin. *Egyptian Journal of Chemistry* 65:609–615. <https://doi.org/10.21608/ejchem.2022.106639.4896>
74. Mahmoud AA, Elfiky AM, Elreedy HA, Salaheldin K (2022) Quercetin attenuates testicular dysfunction induced by aluminum chloride in male Wistar rats. *Egyptian Journal of Chemistry* 65:665–676. <https://doi.org/10.21608/ejchem.2021.102711.4760>
75. Damiano F, Giannotti L, Gnoni GV, Siculella L, Gnoni A (2019) Quercetin inhibition of SREBPs and ChREBP expression results in reduced cholesterol and fatty acid synthesis in C6 glioma cells. *Int J Biochem Cell Biol* 117:105618. <https://doi.org/10.1016/j.biocel.2019.105618>
76. Saleh Al-Maamari JN, Rahmadi M, Panggono SM, Prameswari DA, Pratiwi ED, Ardianto C, Balan SS, Suprpti B (2021) The effects of quercetin on the expression of SREBP-1c mRNA in high-fat diet-induced NAFLD in mice. *J Basic Clin Physiol Pharmacol* 32:637–644. <https://doi.org/10.1515/jbcp-2020-0423>
77. Gnoni A, Di Chiara Stanca B, Giannotti L, Gnoni GV, Siculella L, Damiano F (2022) Quercetin Reduces Lipid Accumulation in a Cell Model of NAFLD by Inhibiting De Novo Fatty Acid Synthesis through the Acetyl-CoA Carboxylase 1/AMPK/PP2A Axis. *International Journal of Molecular Sciences* 23:1044. <https://doi.org/10.3390/ijms23031044>
78. Sabbagh GM, Awwad P, Alwassouf S (2023) In-vitro and In-silico Evaluation of Citrus Peel Flavonoids as Potential Antibacterial Agents against *Streptococcus pneumoniae* Isolated from Pneumonia Patients. *Egyptian Journal of Chemistry* 66:363–376. <https://doi.org/10.21608/ejchem.2023.185639.7427>
79. Banjerdpongchai R, Wudtiwai B, Khaw-on P, Rachakhom W, Duangnil N, Kongtawelert P (2015) Hesperidin from Citrus seed induces human hepatocellular carcinoma HepG2 cell apoptosis via both mitochondrial and death receptor pathways. *Tumour Biol* 37:227–237. <https://doi.org/10.1007/s13277-015-3774-7>
80. Chen H, Nie T, Zhang P, Ma J, Shan A (2022) Hesperidin attenuates hepatic lipid accumulation in mice fed high-fat diet and oleic acid induced HepG2 via AMPK activation. *Life Sciences* 296:120428. <https://doi.org/10.1016/j.lfs.2022.120428>
81. Morshedzadeh N, Ramezani Ahmadi A, Behrouz V, Mir E (2023) A narrative review on the role of hesperidin on metabolic parameters, liver enzymes, and inflammatory markers in nonalcoholic fatty liver disease. *Food Science & Nutrition* 11:7523–7533. <https://doi.org/10.1002/fsn3.3729>
82. Allister EM, Borradaile NM, Edwards JY, Huff MW (2005) Inhibition of Microsomal Triglyceride Transfer Protein Expression and Apolipoprotein B100 Secretion by the Citrus Flavonoid Naringenin and by Insulin Involves Activation of the Mitogen-Activated Protein Kinase Pathway in Hepatocytes. *Diabetes* 54:1676–1683. <https://doi.org/10.2337/diabetes.54.6.1676>
83. Lim W, Yang C, Bazer FW, Song G (2016) Luteolin Inhibits Proliferation and Induces Apoptosis of Human Placental Choriocarcinoma Cells by Blocking the PI3K/AKT Pathway and Regulating Sterol Regulatory Element Binding Protein Activity. *Biol Reprod* 95:82. <https://doi.org/10.1095/biolreprod.116.141556>
84. Wong TY, Lin S, Leung LK (2015) The Flavone Luteolin Suppresses SREBP-2 Expression and Post-Translational Activation in Hepatic Cells. *PLoS One* 10:e0135637. <https://doi.org/10.1371/journal.pone.0135637>

85. Miyata S, Inoue J, Shimizu M, Sato R (2015) Xanthohumol Improves Diet-induced Obesity and Fatty Liver by Suppressing Sterol Regulatory Element-binding Protein (SREBP) Activation. *Journal of Biological Chemistry* 290:20565–20579. <https://doi.org/10.1074/jbc.M115.656975>
86. Doddapattar P, Radović B, Patankar JV, Obrowsky S, Jandl K, Nushold C, Kolb D, Vujić N, Doshi L, Chandak PG, Goeritzer M, Ahammer H, Hoefler G, Sattler W, Kratky D (2013) Xanthohumol ameliorates atherosclerotic plaque formation, hypercholesterolemia, and hepatic steatosis in ApoE-deficient mice. *Molecular Nutrition & Food Research* 57:1718–1728. <https://doi.org/10.1002/mnfr.201200794>
87. Casaschi A, Rubio BK, Maiyoh GK, Theriault AG (2004) Inhibitory activity of diacylglycerol acyltransferase (DGAT) and microsomal triglyceride transfer protein (MTP) by the flavonoid, taxifolin, in HepG2 cells: potential role in the regulation of apolipoprotein B secretion. *Atherosclerosis* 176:247–253. <https://doi.org/10.1016/j.atherosclerosis.2004.05.020>
88. Hussain MM, Bakillah A (2008) New approaches to target microsomal triglyceride transfer protein. *Current opinion in lipidology* 19:572. <https://doi.org/10.1097/MOL.0b013e328312707c>
89. Iftekhhar ASMM, Rayhan I, Quadir MA, Akhteruzzaman S, Hasnat A (2006) Effect of Tamarindus indica fruits on blood pressure and lipid-profile in human model: an in vivo approach. *Pak J Pharm Sci* 19:125–129
90. Devaraj S, Vega-López S, Kaul N, Schönlaue F, Rohdewald P, Jialal I (2002) Supplementation with a pine bark extract rich in polyphenols increases plasma antioxidant capacity and alters the plasma lipoprotein profile. *Lipids* 37:931–934. <https://doi.org/10.1007/s11745-006-0982-3>
91. Zhang J, Wu X, Zhong B, Liao Q, Wang X, Xie Y, He X (2023) Review on the Diverse Biological Effects of Glabridin. *Drug Design, Development and Therapy* 17:15. <https://doi.org/10.2147/DDDT.S385981>
92. Lee M-H, Kim H-M, Chung H-C, Lee J-H (2020) Licorice extract suppresses adipogenesis through regulation of mitotic clonal expansion and adenosine monophosphate-activated protein kinase in 3T3-L1 cells. *Journal of Food Biochemistry* 44:e13528. <https://doi.org/10.1111/jfbc.13528>
93. Kheiripour N, J K, I K, H T, Mt G, M H (2018) Silymarin prevents lipid accumulation in the liver of rats with type 2 diabetes via sirtuin1 and SREBP-1c. *Journal of basic and clinical physiology and pharmacology* 29:. <https://doi.org/10.1515/jbcpp-2017-0122>
94. Gillessen A, Schmidt HH-J (2020) Silymarin as Supportive Treatment in Liver Diseases: A Narrative Review. *Adv Ther* 37:1279–1301. <https://doi.org/10.1007/s12325-020-01251-y>
95. Aghemo A, Alekseeva OP, Angelico F, Bakulin IG, Bakulina NV, Bordin D, Bueverov AO, Drapkina OM, Gillessen A, Kagarmanova EM, Korochanskaya NV, Kucheryavii UA, Lazebnik LB, Livzan MA, Maev IV, Martynov AI, Osipenko MF, Sas EI, Starodubova A, Uspensky YP, Vinnitskaya EV, Yakovenko EP, Yakovlev AA (2022) Role of silymarin as antioxidant in clinical management of chronic liver diseases: a narrative review. *Annals of Medicine* 54:1548–1560. <https://doi.org/10.1080/07853890.2022.2069854>
96. Morelli V, Zoorob RJ (2000) Alternative Therapies: Part II. Congestive Heart Failure and Hypercholesterolemia. *afp* 62:1325–1330
97. Noriega-López, Tovar, Gonzalez-Granillo, Hernández-Pando, Escalante, Santillán-Doherty, Torres (2007) Pancreatic insulin secretion in rats fed a soy protein high fat diet depends on the interaction between the amino acid pattern and isoflavones. *The Journal of biological chemistry* 282:. <https://doi.org/10.1074/jbc.M701045200>
98. Yw K, Ym K, Ym Y, Th K, Sj H, Jr L, Sc K, Sg K (2010) Inhibition of SREBP-1c-mediated hepatic steatosis and oxidative stress by sauchinone, an AMPK-activating lignan in Saururus chinensis. *Free radical biology & medicine* 48:. <https://doi.org/10.1016/j.freeradbiomed.2009.12.006>
99. Chae H-S, You BH, Kim D-Y, Lee H, Ko HW, Ko H-J, Choi YH, Choi SS, Chin Y-W (2018) Sauchinone controls hepatic cholesterol homeostasis by the negative regulation of PCSK9 transcriptional network. *Scientific Reports* 8:. <https://doi.org/10.1038/s41598-018-24935-6>
100. Kim YW, Kim YM, Yang YM, Kim TH, Hwang SJ, Lee JR, Kim SC, Kim SG (2010) Inhibition of SREBP-1c-mediated hepatic steatosis and oxidative stress by sauchinone, an AMPK-activating lignan in Saururus chinensis. *Free Radical Biology and Medicine* 48:567–578. <https://doi.org/10.1016/j.freeradbiomed.2009.12.006>
101. Mustafa YF (2016) Synthesis, Antioxidant, and Preliminary Antitumor Activities of New Curcumin Analogues. *Journal of Global Pharma Technology*
102. Akbari M, Lankarani KB, Tabrizi R, Ghayour-Mobarhan M, Peymani P, Ferns G, Ghaderi A, Asemi Z (2019) The Effects of Curcumin on Weight Loss Among Patients With Metabolic Syndrome and Related Disorders: A Systematic Review and Meta-Analysis of Randomized Controlled Trials. *Front Pharmacol* 10:. <https://doi.org/10.3389/fphar.2019.00649>
103. Ding L, Li J, Song B, Xiao X, Zhang B, Qi M, Huang W, Yang L, Wang Z (2016) Curcumin rescues high fat diet-induced obesity and insulin sensitivity in mice through regulating SREBP pathway. *Toxicol Appl Pharmacol* 304:99–109. <https://doi.org/10.1016/j.taap.2016.05.011>
104. Du Y, Li D, Lu D, Zhang R, Zhong Q, Zhao Y, Zheng X, Ji S, Wang L, Tang D-Q (2022) Amelioration of lipid accumulations and metabolism disorders in differentiation and development of 3T3-L1 adipocytes through mulberry leaf water extract. *Phytomedicine* 98:153959. <https://doi.org/10.1016/j.phymed.2022.153959>
105. El-Tantawy WH, Temraz A (2019) Natural products for controlling hyperlipidemia: review. *Archives of Physiology and Biochemistry* 125:128–135. <https://doi.org/10.1080/13813455.2018.1441315>
106. Li Y, Xu S, Mihaylova M, Zheng B, Hou X, Jiang B, Park O, Luo Z, Lefai E, Shyy JY-J, Gao B, Wierzbicki M, Verbeuren TJ, Shaw RJ, Cohen RA, Zang M (2011) AMPK Phosphorylates and Inhibits SREBP Activity to Attenuate

- Hepatic Steatosis and Atherosclerosis in Diet-induced Insulin Resistant Mice. *Cell Metab* 13:376–388. <https://doi.org/10.1016/j.cmet.2011.03.009>
107. Kubota S, Tanaka Y, Nagaoka S (2019) Ellagic acid affects mRNA expression levels of genes that regulate cholesterol metabolism in HepG2 cells. *Bioscience, Biotechnology, and Biochemistry* 83:952–959. <https://doi.org/10.1080/09168451.2019.1576498>
108. Xu Q, Li S, Tang W, Yan J, Wei X, Zhou M, Diao H (2021) The Effect of Ellagic Acid on Hepatic Lipid Metabolism and Antioxidant Activity in Mice. *Front Physiol* 12. <https://doi.org/10.3389/fphys.2021.751501>
109. Wang G-L, Fu Y-C, Xu W-C, Feng Y-Q, Fang S-R, Zhou X-H (2009) Resveratrol inhibits the expression of SREBP1 in cell model of steatosis via Sirt1–FOXO1 signaling pathway. *Biochemical and Biophysical Research Communications* 380:644–649. <https://doi.org/10.1016/j.bbrc.2009.01.163>
110. Fukuda M, Ogasawara Y, Hayashi H, Inoue K, Sakashita H (2022) Resveratrol Inhibits Proliferation and Induces Autophagy by Blocking SREBP1 Expression in Oral Cancer Cells. *Molecules* 27:8250. <https://doi.org/10.3390/molecules27238250>
111. Zhou C, Qian W, Ma J, Cheng L, Jiang Z, Yan B, Li J, Duan W, Sun L, Cao J, Wang F, Wu E, Wu Z, Ma Q, Li X (2019) Resveratrol enhances the chemotherapeutic response and reverses the stemness induced by gemcitabine in pancreatic cancer cells via targeting SREBP1. *Cell Proliferation* 52:e12514. <https://doi.org/10.1111/cpr.12514>
112. Mustafa YF (2023) Synthesis, in silico analysis, and biomedical effects of coumarins derived from resveratrol. *Phytomedicine Plus* 3:100501
113. Tang J-J, Li J-G, Qi W, Qiu W-W, Li P-S, Li B-L, Song B-L (2011) Inhibition of SREBP by a small molecule, betulin, improves hyperlipidemia and insulin resistance and reduces atherosclerotic plaques. *Cell Metab* 13:44–56. <https://doi.org/10.1016/j.cmet.2010.12.004>
114. Peng A, Liu S, Fang L, Zhu Z, Zhou Y, Yue S, Ma Z, Liu X, Xue S, Qiu Y, Qi R (2022) Inonotus obliquus and its bioactive compounds alleviate non-alcoholic fatty liver disease via regulating FXR/SHP/SREBP-1c axis. *European Journal of Pharmacology* 921:174841. <https://doi.org/10.1016/j.ejphar.2022.174841>
115. Peng C-H, Ker Y-B, Li H-H, Tsou S-H, Lin C-L, Huang C-N (2022) Abelmoschus esculentus subfractions ameliorate hepatic lipogenesis and lipid uptake via regulating dipeptidyl peptidase-4—With improving insulin resistance. *PLoS ONE* 17:e0265444. <https://doi.org/10.1371/journal.pone.0265444>
116. Kong W-J, Wei J, Zuo Z-Y, Wang Y-M, Song D-Q, You X-F, Zhao L-X, Pan H-N, Jiang J-D (2008) Combination of simvastatin with berberine improves the lipid-lowering efficacy. *Metabolism - Clinical and Experimental* 57:1029–1037. <https://doi.org/10.1016/j.metabol.2008.01.037>
117. Neag MA, Mocan A, Echeverría J, Pop RM, Bocsan CI, Crişan G, Buzoianu AD (2018) Berberine: Botanical Occurrence, Traditional Uses, Extraction Methods, and Relevance in Cardiovascular, Metabolic, Hepatic, and Renal Disorders. *Front Pharmacol* 9. <https://doi.org/10.3389/fphar.2018.00557>
118. Liu Y, Hua W, Li Y, Xian X, Zhao Z, Liu C, Zou J, Li J, Fang X, Zhu Y (2020) Berberine suppresses colon cancer cell proliferation by inhibiting the SCAP/SREBP-1 signaling pathway-mediated lipogenesis. *Biochemical Pharmacology* 174:113776. <https://doi.org/10.1016/j.bcp.2019.113776>
119. Zhang Y, Jin J, Li P, Yang H, Zheng Z (2022) Recent advances in research on natural product inhibitors of SREBPs. *Tradit Med Res* 7:23. <https://doi.org/10.53388/TMR20220214264>
120. Y, Liu J, Hao W, He Z, Zhu H, Liang N, Ma KY, He W-S, Yang Y, Chen Z-Y (2018) Plasma cholesterol-lowering activity of piperine is mediated by inhibition on cholesterol absorption via down-regulation of intestinal ACAT2 and MTP. *Journal of Functional Foods* 49:465–471. <https://doi.org/10.1016/j.jff.2018.09.014>
121. Fukumitsu S, Villareal MO, Onaga S, Aida K, Han J, Isoda H (2013) α -Linolenic acid suppresses cholesterol and triacylglycerol biosynthesis pathway by suppressing SREBP-2, SREBP-1a and -1c expression. *Cytotechnology* 65:899–907. <https://doi.org/10.1007/s10616-012-9510-x>
122. Jeyapal S, Kona SR, Mullapudi SV, Putcha UK, Gurumurthy P, Ibrahim A (2018) Substitution of linoleic acid with α -linolenic acid or long chain n-3 polyunsaturated fatty acid prevents Western diet induced nonalcoholic steatohepatitis. *Sci Rep* 8:10953. <https://doi.org/10.1038/s41598-018-29222-y>
123. Ou J, Tu H, Shan B, Luk A, DeBose-Boyd RA, Bashmakov Y, Goldstein JL, Brown MS (2001) Unsaturated fatty acids inhibit transcription of the sterol regulatory element-binding protein-1c (SREBP-1c) gene by antagonizing ligand-dependent activation of the LXR. *Proc Natl Acad Sci U S A* 98:6027–6032. <https://doi.org/10.1073/pnas.111138698>
124. Xian M, Oh M, Kwak HJ, Jeong H, Ko H-J, Kim SH (2023) Chemical constituents from the stem bark of *Albizia julibrissin* and their SREBP-1c inhibitory activity. *Journal of Asian Natural Products Research* 0:1–6. <https://doi.org/10.1080/10286020.2022.2163633>
125. Yoshikawa T, Shimano H, Yahagi N, Ide T, Amemiya-Kudo M, Matsuzaka T, Nakakuki M, Tomita S, Okazaki H, Tamura Y, Iizuka Y, Ohashi K, Takahashi A, Sone H, Osuga J, Gotoda T, Ishibashi S, Yamada N (2002) Polyunsaturated Fatty Acids Suppress Sterol Regulatory Element-binding Protein 1c Promoter Activity by Inhibition of Liver X Receptor (LXR) Binding to LXR Response Elements*. *Journal of Biological Chemistry* 277:1705–1711. <https://doi.org/10.1074/jbc.M105711200>
126. Voragen AGJ, Coenen G-J, Verhoef RP, Schols HA (2009) Pectin, a versatile polysaccharide present in plant cell walls. *Struct Chem* 20:263–275. <https://doi.org/10.1007/s11224-009-9442-z>
127. Miettinen TA, Tarpila S (1977) Effect of pectin on serum cholesterol, fecal bile acids and biliary lipids in normolipidemic and hyperlipidemic individuals. *Clin Chim Acta* 79:471–477. [https://doi.org/10.1016/0009-8981\(77\)90444-2](https://doi.org/10.1016/0009-8981(77)90444-2)

128. Wang M-M, Wang F, Li G, Tang M-T, Wang C, Zhou Q-Q, Zhou T, Gu Q (2022) Antioxidant and hypolipidemic activities of pectin isolated from citrus canning processing water. *LWT* 159:113203. <https://doi.org/10.1016/j.lwt.2022.113203>
129. Banerjee SK, Maulik SK (2002) Effect of garlic on cardiovascular disorders: a review. *Nutr J* 1:4. <https://doi.org/10.1186/1475-2891-1-4>
130. Sun Y-E, Wang W, Qin J (2018) Anti-hyperlipidemia of garlic by reducing the level of total cholesterol and low-density lipoprotein. *Medicine (Baltimore)* 97:e0255. <https://doi.org/10.1097/MD.00000000000010255>
131. Isaacsohn JL, Moser M, Stein EA, Dudley K, Davey JA, Liskov E, Black HR (1998) Garlic Powder and Plasma Lipids and Lipoproteins: A Multicenter, Randomized, Placebo-Controlled Trial. *Archives of Internal Medicine* 158:1189–1194. <https://doi.org/10.1001/archinte.158.11.1189>
132. Ashraf R, Amir K, Shaikh A, Ahmed T (2004) Effects of garlic on dyslipidemia in patients with type 2 diabetes mellitus. *Journal of Ayub Medical College, Abbottabad : JAMC* 17:60–4
133. Qi J, Tan CK, Hashimi SM, Zulfiker AHM, Good D, Wei MQ (2014) Toad Glandular Secretions and Skin Extractions as Anti-Inflammatory and Anticancer Agents. *Evidence-Based Complementary and Alternative Medicine* 2014:e312684. <https://doi.org/10.1155/2014/312684>
134. Emam, Qi Z, Y F, A R, K A, M K, T K (2012) Apoptotic cell death by the novel natural compound, cinobufotalin. *Chemico-biological interactions* 199:. <https://doi.org/10.1016/j.cbi.2012.07.005>
135. Han Y, Ma R, Cao G, Liu H, He L, Tang L, Li H, Luo Q (2021) Combined Treatment of Cinobufotalin and Gefitinib Exhibits Potent Efficacy against Lung Cancer. *Evidence-Based Complementary and Alternative Medicine* 2021:e6612365. <https://doi.org/10.1155/2021/6612365>
136. Meng H, Shen M, Li J, Zhang R, Li X, Zhao L, Huang G, Liu J (2021) Novel SREBP1 inhibitor cinobufotalin suppresses proliferation of hepatocellular carcinoma by targeting lipogenesis. *European Journal of Pharmacology* 906:174280. <https://doi.org/10.1016/j.ejphar.2021.174280>
137. Wang J, Chang H, Su M, Zhao H, Qiao Y, Wang Y, Shang L, Shan C, Zhang S (2022) The Potential Mechanisms of Cinobufotalin Treating Colon Adenocarcinoma by Network Pharmacology. *Frontiers in Pharmacology* 13:
138. Li, S P, X Z, D Q, W Z, Y D, R Y, X Y, Z Z, S X, D F, H S (2022) Cinobufotalin inhibits the epithelial-mesenchymal transition of hepatocellular carcinoma cells through down-regulate β -catenin in vitro and in vivo. *European journal of pharmacology* 922:. <https://doi.org/10.1016/j.ejphar.2022.174886>
139. Hannan JMA, Rokeya B, Faruque O, Nahar N, Mosihuzzaman M, Azad Khan AK, Ali L (2003) Effect of soluble dietary fibre fraction of *Trigonella foenum graecum* on glycemic, insulinemic, lipidemic and platelet aggregation status of Type 2 diabetic model rats. *Journal of Ethnopharmacology* 88:73–77. [https://doi.org/10.1016/S0378-8741\(03\)00190-9](https://doi.org/10.1016/S0378-8741(03)00190-9)
140. Uemura T, Goto T, Kang M, Mizoguchi N, Hirai S, Lee J, Nakano Y, Shono J, Hoshino S, Taketani K, Tsuge N, Narukami T, Makishima M, Takahashi N, Kawada T (2011) Diosgenin, the Main Aglycon of Fenugreek, Inhibits LXR α Activity in HepG2 Cells and Decreases Plasma and Hepatic Triglycerides in Obese Diabetic Mice. *The Journal of Nutrition* 141:17–23. <https://doi.org/10.3945/jn.110.125591>
141. Annida B, Stanely Mainzen Prince P (2004) Supplementation of Fenugreek Leaves Lower Lipid Profile in Streptozotocin-Induced Diabetic Rats. *Journal of Medicinal Food* 7:153–156. <https://doi.org/10.1089/1096620041224201>
142. Khateeb S, Albalawi A, Alkhedaide A (2021) Regulatory effect of diosgenin on lipogenic genes expression in high-fat diet-induced obesity in mice. *Saudi Journal of Biological Sciences* 28:1026–1032. <https://doi.org/10.1016/j.sjbs.2020.11.045>
143. Zhang S-Z, Liang P-P, Feng Y-N, Yin G-L, Sun F-C, Ma C-Q, Zhang F-X (2022) Therapeutic potential and research progress of diosgenin for lipid metabolism diseases. *Drug Development Research* 83:1725–1738. <https://doi.org/10.1002/ddr.21991>
144. Wang D, Wang X (2022) Diosgenin and Its Analogs: Potential Protective Agents Against Atherosclerosis. *DDDT* 16:2305–2323. <https://doi.org/10.2147/DDDT.S368836>
145. Watanabe M, Uesugi M (2013) Small-molecule inhibitors of SREBP activation – potential for new treatment of metabolic disorders. *Med Chem Commun* 4:1422–1433. <https://doi.org/10.1039/C3MD00177F>
146. Hussain M, Nijstad N, Franceschini L (2011) Regulation of microsomal triglyceride transfer protein. *Clinical Lipidology* 6:293–303. <https://doi.org/10.2217/clp.11.21>
147. Blom DJ, Raal FJ, Santos RD, Marais AD (2019) Lomitapide and Mipomersen—Inhibiting Microsomal Triglyceride Transfer Protein (MTP) and apoB100 Synthesis. *Curr Atheroscler Rep* 21:48. <https://doi.org/10.1007/s11883-019-0809-3>
148. Gouni-Berthold I, Berthold HK (2015) Mipomersen and lomitapide: Two new drugs for the treatment of homozygous familial hypercholesterolemia. *Atheroscler Suppl* 18:28–34. <https://doi.org/10.1016/j.atherosclerosissup.2015.02.005>
149. Chi C, Harth L, Galera MR, Torrealba MP, Vadivel CK, Geisler C, Bonefeld CM, Nielsen PR, Bzorek M, Becker JC, Gjerdrum LMR, Ødum N, Woetmann A (2022) Concomitant Inhibition of FASN and SREBP Provides a Promising Therapy for CTCL. *Cancers* 14:4491. <https://doi.org/10.3390/cancers14184491>
150. Zhao X, Xiaoli A, Zong H, Aa A, Yang E, Ji J-Y, Pessin J, Das B, Yang F (2014) Inhibition of SREBP Transcriptional Activity by a Boron-Containing Compound Improves Lipid Homeostasis in Diet-Induced Obesity. *Diabetes* 63:. <https://doi.org/10.2337/db13-0835>