



## Effect of Diode Laser Irradiation and Application of Nanoparticle Herbal Endodontic Irrigation Solutions on *Candida Albicans* and *Enterococcus Faecalis* Bacteria



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*In Loving Memory of Late Professor Doctor "Mohamed Refaat Hussein Mahran"*

### Abstract

The present study investigate whether diode laser activation of two nanoparticle herbal irrigants affect *Enterococcus faecalis* (*E.faecalis*) biofilm and *Candida albicans* (*C. albicans*) compared with photodynamic therapy (PDT) utilizing methylene blue as photosensitizer (PS). Material and methods: sixty single rooted teeth had undergo mechanical preparation. Teeth were divided into 3 groups (20 teeth each), G1 : 20 teeth inoculated with *Enterococcus faecalis* half (n= 10) irrigated with nanocurcumin and the other half irrigated with nanoneem both activated by diode laser, G2: 20 teeth inoculated with *Candida albicans* irrigated similar to group 1 and G3: 10 teeth inoculated with *E.faecalis* and 10 with *Candida albicans* undergo PDT using methylene blue as PS. Results: For *E.faecalis* no statistically significant difference neither between nanoneem and nanocurcumin ( $p>0.05$ ) nor between nanocurcumin and PDT ( $p>0.05$ ). While there was a statistically significant difference between nanoneem and PDT ( $p<0.001$ ). While for *C. albicans* no statistically significant difference between nanocurcumin and PDT ( $p>0.05$ ). While the differences were statistically highly significant between nanoneem and PDT ( $p=0.002$ ) and between nanoneem and nanocurcumin ( $p<0.001$ ). Conclusion: nanoneem and nanocurcumin irrigants activated by laser have an effect on root canal microorganisms and could be used as adjunctive endodontic irrigant.

Keywords: Root canal irrigants; antibacterial and antifungal effect; nanocurcumin; nanoneem, photodynamic therapy

### 1. Introduction

Many studies have shown that microbial organisms like *Enterococcus faecalis* (*E. faecalis*) and *Candida albicans* (*C. albicans*) are found in endodontic infections that lead to failure of root canal treatment (RCT) [1, 2]. Moreover, bacteria may reach the outer surface of the root by penetrating the dentinal tubules of the root, causing failure of root canal treatment [3]. Adequate penetration of antimicrobial irrigant solutions into anatomical complexities is essential for good debridement and disinfection [4]. Strategies of disinfection that use lasers or light can be divided into three groups: direct laser irradiation, laser-activated irrigation, and photodynamic therapy [5].

Laser wavelength, its energy, pulse frequency, and the design of the fiber optic tip, together with its position within the root canal, could influence the effect of LAI. Water, present in the endodontic irrigant solutions, blunts the thermal effect of the

laser beam on the root dentin, but at the same time, when it becomes a target chromophore by mid-infrared laser activation, it can work synergistically to clean the canal [6]. Diode laser (DL) activation of irrigants has shown more favorable results, where 810-nm diode laser (DL) decreased the number of *E. faecalis* bacteria through invading dentinal tubules [7]. While different diode laser wavelengths (940- and 980-nm) are more highlighted because these are closely inconsistent with water absorption and are much more absorbed [8], For endodontic applications, activation of the irrigants could be done using a diode laser with parameters of 940-nm wavelength, a 50–60 Hz frequency system, and a maximum output power of 7 W using 200- $\mu$ m plain-ended fiber, which consequently removes debris and the smear layer [9]. Photodynamic therapy (PDT) is based on utilizing light of an appropriate wavelength to activate photosensitizing molecules adhered to the bacterial or fungal membrane.

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It results in exciting its electronic layers to a so-called triplet state [10]. As photosensitizers, methylene blue (MB) and toluidine blue O (TBO) are amphibic; they can be used against both gram-positive and gram-negative bacteria present in root canal infections [11, 12]. PDT could be applied by a diode laser, in which the optical fiber is placed within the root canal near its full length, thus allowing for even light distribution both vertically and horizontally, resulting in better disinfection within the root canal system.

Herbal medicine has been widely used as a therapeutic agent because of its biocompatibility, safety, and cost-effectiveness, which raises the need to explore its application in the dental field [13]. Neem is one of the few herbal extracts that has antimicrobial properties along with other anti-cariogenic and anti-inflammatory activities [14]. It can be used as an effective substitute for sodium hypochlorite due to its high biocompatibility [15]. However, the major disadvantage of neem is its pungent taste, which reduces the patient's acceptance and can be neutralized by adding sweeteners [16]. Although neem has an efficient antimicrobial effect on many microbes, different studies have mentioned that neem has limited action against the main microbes in root canal infections like *E. faecalis* and *candida albicans* [17, 18].

So the use of nanotechnology to modify the particle size of neem extract may be beneficial to improve its antimicrobial action against various microorganisms present in endodontic infections. Curcumin, the main component of turmeric powder, has been used as a photosensitizer (PS) for PDT recently in dentistry [19]. *Curcuma longa* is a yellowish-orange powder that is a natural component of the rhizome, and it has a characteristic antimicrobial effect on bacteria, viruses, and fungi [20]. Gomes-Filho et al. (2016) [21] reported that PDT utilizing curcumin as PS had a benign effect on pulpal cells in addition to not interfering with fibroblast viability. Standard PDT performed with a diode laser and methylene blue performed similar bacterial inactivation as a combination of curcumin with the blue LED light [20]. Some studies showed that the antifungal properties of nanocurcumin were more potent than those of native curcumin [22].

Nanoparticles gained several properties that may improve the treatment of endodontic infections: synergistically increased antibacterial activity, increased reactivity, and the ability to interact with other reactive compounds. This leads to the innovation of various experimental nanoparticle-incorporated obturation materials and sealers that perform a range of favorable physicochemical properties, including enhanced antibacterial

performance and bioactivity. Therefore, this study will investigate the effect of herbal irrigants, especially neem and curcumin, in nanofarm after being activated by a diode laser in comparison with standard PDT using methylene blue.

## 2. Materials and methods

### 2.1. Preparation of nanoneem and nanocurcumin irrigants

The ultrasonic-solvent emulsification technique was used to prepare solid lipid nanoparticles (SLN), also called nanostructured lipid carriers, according to Adel [23]. Two phases were prepared: the oil phase and the water phase.

After volatilizing most of the solvents, the water phase was added to the oil phase. Finally, the impurity materials were filtered from the oil-loaded SLNs suspension and then stored at 4 °C for further bioassays.

Transmission electron microscopy (TEM) analysis was done, confirming their size below 100 nanometers. The effective electric charge on the nanoparticle's surface is measured by a test called Zeta, which quantifies the charges. This measure was done for both liquids showing negative Zeta potential.

### 2.2. Microorganism and Culture Conditions

Qualitative evaluations were carried out in nutrient broth according to Elboraey et al., 2021 [24] and Sultan et al., 2022 [25]. The pathogenic microorganisms used in this study were Gram-positive bacteria [*Enterococcus faecalis* ATCC 29212] and pathogenic yeast [*Candida albicans* ATCC 10231], prepared from fresh overnight broth cultures using nutrient broth medium.

20.0 µL of the above-prepared inoculum of each test strain was separately inoculated into each 2.0 mL-volume cryovial containing one tooth with one central hole filled with 1.0 ml of the sterile nutrient broth medium (NB). The inoculated cryovials with teeth were incubated at 37°C for 21 days for more test strain attachment. After the proper incubation period, the first inoculum was taken from each cryovial separately as a pre-inoculum or control sample.

### 2.3. Preparation of teeth

The extracted single-rooted teeth (n = 60) with one canal and well-developed apices were selected after checking their periapical radiograph. Teeth with caries, fracture, resorption, or malformation are excluded. De-coronation of all teeth was done using a diamond disc (Mani, Inc., Tochigi, Japan). Light cure composite (3M ESPE Z250, USA) was used to seal all the apical foramina of all teeth, which were

coated with two layers of varnish to prevent bacterial leakage.

Endodontic access was done with a 4-round carbide bur (DENTSPLY Maillefer, OK, USA). The working length was confirmed using a radiograph and was kept half a millimeter short of the radiographic apex. Preparation was done using the crown-down technique, where coronal preparation was done using Gates Glidden drills size 1:4. Mechanical preparation was done up to 30 stainless steel hand files, and rotary instrumentation was done using Pro-Taper (DENTSPLY Maillefer, OK, USA) up to F3. All the teeth were autoclaved twice at 121 °C for the elimination of all the microbes from the tooth and to prevent contamination of the study with other microbes.

Sixty tooth samples are divided into three groups and undergo different interventions, as follows:

**G1:** 20 teeth were inoculated with *Enterococcus faecalis*; 10 teeth were irrigated with nanocurcumin; and another 10 teeth were irrigated with nanoneem. All irrigated teeth undergo activation by a diode laser system (Sirona, Bensheim, Germany) at 980 nm, power 0.5 W, with pulse settings ranging from 10 to 10,000 Hz in eight increments. In this intervention, a 200 µm laser tip was placed into the root canal at a preset position 1 mm from the closed end, and the fiber tip moved slowly in an up-and-down motion within the canal for 20 seconds, repeated three times at 10-second intervals. The panel settings were 2.5 W/15 Hz with an average power of 1.2 W for the diode laser.

**G2:** 20 teeth inoculated with *C. albicans*. Irrigated similarly to group 1 and undergo laser activation with the same parameters for an equal period of exposure.

**G3:** 10 teeth inoculated with *E. faecalis* and 10 with *Candida albicans*. For both groups, 1 mL of methylene blue was injected in the root canal using a 25-gauge needle and irradiated with a 200 µm fiber tip using a diode laser of wave length 650 nm (Pioon, Wahan, China) applied with settings of 200 mW and an energy of 60 joules. The photosensitizer is activated for 60 seconds in a circular motion with slight up-and-down movements.

In this step, a serial dilution from each [pre-inoculum or control group] and from each sample treated has been done ( $10^{-1}$ ,  $10^{-4}$ ) separately.

The microbial inhibition was determined using the total viable count technique by counting the colony-forming units (CFU) using the Scan 100 colony counter system (France).

A diode laser is an electrically pumped semiconductor laser in which the active laser medium is formed by a p-n junction of a semiconductor diode. A 980 nm diode laser was used

for its high transmission due to its higher absorption in water. This allows for microbial reduction at a depth of 1.000µm. Siro-Laser Advance Plus is a diode laser device with both an infrared and a red diode. The established infrared 980 nm wavelength covers all indications in the fields of soft-tissue surgery, periodontology, and endodontics. This enables it to cover more than 20 indications.

### 3. Results

Table (1) shows a comparison between nanoneem (median 190.0, IQR 135.3-207.0) and nanocurcumin (median 69.0, IQR 18.5-137.3) activated by laser and PDT using methylene blue (median 4.5, IQR 0-39.5) on *Enterococcus faecalis* bacteria. G1 showed efficient antimicrobial activity against *E. faecalis*, with significant results compared to pretreatment samples. The results revealed that there was a statistically highly significant difference among groups ( $p < 0.01$ ), while comparisons between each group and another showed no statistically significant difference, neither between nanoneem and nanocurcumin ( $p > 0.05$ ) nor between nanocurcumin and PDT ( $p > 0.05$ ). The difference was statistically highly significant between nanoneem and PDT ( $p < 0.001$ ).

Fig. (1) Shows a comparison between nanoneem, nanocurcumin activated by laser, and methylene blue CFU\* $10^{-4}$  of bacterial *Enterococcus faecalis*. There is a statistically highly significant difference between nanoneem and methylene blue ( $p < 0.001$ ). While the differences between nanoneem and nanocurcumin and between nanocurcumin and methylene blue are statistically insignificant ( $p > 0.05$ ).

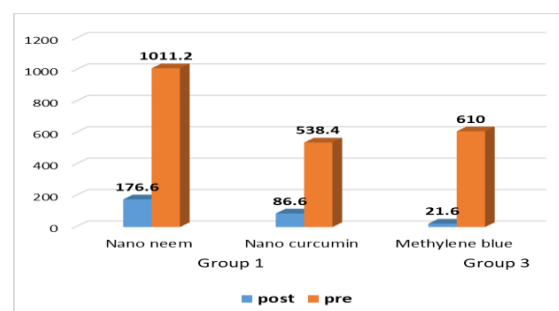


Fig. (1): Comparison between nanoneem and nanocurcumin group1 and MB group 3 samples on *E. faecalis* CFU\* $10^{-4}$

Table (2) shows a comparison between nanoneem (median 1.0, IQR 0-1.0), nanocurcumin (median 307.5, IQR 219.3-508.5) activated by laser, and PDT (median 257.0, IQR 242.3-276.5) on *Candida albicans*. Both nanoherbal irrigants activated by diode laser showed significant results against *Candida albicans* compared to pre-inoculum samples. The results revealed that there was a

statistically highly significant difference among groups ( $p < 0.01$ ), while comparisons between each group and another showed no statistically significant difference between nanocurcumin and methylene blue ( $p > 0.05$ ). The differences were statistically highly significant between nanoneem and methylene blue ( $p = 0.002$ ) and between nanoneem and nanocurcumin ( $p < 0.001$ ).

Fig (2) shows a comparison between the second and third group samples of *Candida albicans* CFU\*10<sup>-4</sup>. There is a statistically highly significant difference between nanoneem and methylene blue ( $p = 0.002$ ) and between nanoneem and nanocurcumin ( $p < 0.001$ ) with the lowest growth in nanoneem by laser group. While the difference between

nanocurcumin and methylene blue is statistically insignificant ( $p > 0.05$ ).

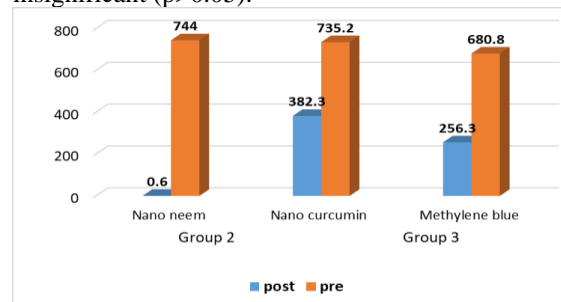


Fig. (2): Comparison between nanoneem and nanocurcumin of group 2 and MB of group 3 samples on *Candida albicans* CFU\*10<sup>-4</sup> (n=10 samples in each group)

Table (1): Statistical comparison between CFU\*10<sup>-4</sup> pre and post treatment of laser activated nanoneem and nanocurcumin irrigants against PDT on *E. faecalis*.

		Group 1 activated by laser		Group 3 PDT using Methylene blue
		<i>nanoNeem</i>	<i>nanoCurcumin</i>	
Post treatment	Mean±SD	176.6±40.8	86.6±82.5	21.6±33.5
	SE	12.9	26.1	10.6
	Median IQR	190 135.3-207.0	69 18.5-137.3	4.5 0-39.5
	Range (min-max)	106.0-224.0	13.0-265.0	0-95.0
pre	Mean±SD	1011.2±358.8	538.4±91.2	610.0±91.9
	SE	160.5	40.8	41.1
	Median IQR	1024 692.0-1324.0	556 454.0-614.0	618.0 522.0-694.0
	Range (min-max)	484.0-1444.0	420.0-664.0	404.0-788.0
	<b>P value</b>	0.006*	<0.001*	<0.001*
			P value	Adjusted significance
	Group 1 nanoneem vs. Group 3		0.001	0.017*
	Group 1 nanocurcumin vs. Group 3		0.06	0.9

Table (2): statistical comparison between CFU\*10<sup>-4</sup> pre and post treatment of activated nanoneem and nanocurcumin irrigants against PDT on *C. albicans*.

		Group 2		Group 3 PDT using Methylene blue
		<i>NanoNeem</i>	<i>NanoCurcumin</i>	
Post treatment	Mean±SD	0.6±0.5	382.3±222.2	256.3±34.8
	SE	0.2	70.3	11.0
	Median (IQR)	1 0-1.0	307.5 219.3-508.5	257.0 242.3-276.5
	Range (min-max)	0-1.0	177.0-870.0	180.0-305.0
pre	Mean±SD	744.0±208.2	735.2±150.7	680.8±158.1
	SE	93.1	67.4	70.7
	Median (IQR)	724.0 558.0-940.0	664.0 608.0-898.0	720.0 560.0-782.0
	Range (min-max)	556.0-1056.0	596.0-904.0	404.0-788.0
	P value	0.001*	0.007*	0.003*
			P value	Adjusted significance
	Group 2 nanoneem vs. Group 3		0.012	0.185
	Group 2 nanocurcumin vs. Group 3		0.635	1.0

#### 4. Discussion

The photodynamic therapy (PDT) principle is effective against bacteria, viruses, fungi, and protozoa [26].

Mature (3 weeks old) bacterial biofilms resist antimicrobial treatments more than young biofilms, so the culture for the microorganisms used in the present study was 21 days old, as Shen et al. (2011 [27]).

In modern clinical trials, toluidine blue O (TBO) and methylene blue (MB) were used as photosensitizers (PSs) with absorption wavelengths ranging from 600–660 nm [28, 29, 30, 31]. Phenothiaziniums, such as MB, attack the plasma membranes of yeast and bacteria and produce bacterial DNA damage [32]. In the present study, MB was used and activated with a diode laser with a wavelength of 650 nm. Methylene blue alone, used at concentrations of 100 µg/mL, could destroy 83% of *E. faecalis* bacteria inside a root canal [33].

A laser was used to activate the MB, in accordance with earlier research by Bago et al. and Rios et al. The irradiation times were 30 and 60 seconds, respectively. There is no agreement in the literature concerning the most reliable PDT parameters for *E. faecalis* destruction inside root canals [34, 35].

A previous study reported that methylene blue in PDT along with the diode laser could result in a 96.7% reduction of *E. coli* (910 nm, 1W at pulsed mode) [36]. The laser parameters used in the present study were similar to those of Pourhajibagher & Bahador et al. (2018 [37]), whose results revealed that the microbial count significantly decreased ( $P < 0.05$ ) when using an output power of 220 mW at 635 nm for 60 s.

The average pre-irradiation time, according to the literature, is 5 to 15 minutes [38]. In this study, PS was left for 5 minutes before irradiation.

Oda et al.'s 2019 comparison of 30 µL of curcumin as PS activated by LED light 430 for 5 min and standard PDT (1% methylene blue activated with diode laser 660nm for 4 min) resulted in no significant difference between both groups. We reported the same results, although nanocurcumin was used as an irrigant, not PS, and with a different laser parameter, 980 nm, for 60 sec at 20 Hz [39].

Nardini et al. reported that MB, when activated at 660 nm, presented similar reactive oxygen species (ROS) production to curcumin photosensitizer [40]. Moreover, a significant reduction in planktonic cultures and biofilms of *E. faecalis* could be achieved by activating curcumin with a red light LED (660 nm). Likewise, nanocurcumin and nanoneem, when activated by laser, produce significant antimicrobial effects compared with pretreatment groups.

In a study investigating irrigation with 2.5% NaOCl in comparison to NaOCl followed by either MB-PDT or curcumin-PDT using a LED lamp emitting a red spectrum with a power peak of 630 nm, there were no significant differences among these groups against *E. faecalis* [41]. Diode laser activation

for nanocurcumin showed comparable results to MB-PDT, but for nanoneem, it showed a significant difference to MB as the latter was better.

Hendi. et al. (2021, compared the activated silver nanoparticle irrigant by diode laser 940 nm for 15 sec at 3 times interval together with 5% NaOCl on *E. faecalis* bacteria and found that NaOCl was significantly more effective [42]. They concluded that laser irradiation could not enhance the effect of silver nanoparticles. On the contrary, the present study showed that diode lasers with the same parameters enhanced the effects of both nanoherbal irrigants compared to pretreatment samples.

Some studies reported that conventional cleaning and irrigation protocols produce a similar reduction in *E. faecalis* counts in infected root canals compared to PDT [43]. Likewise, others use methylene blue (50 mg/mL) as photosensitizers and red light (665 nm), with an irradiation time of five minutes and an output power of one watt, to obtain the same results. [44]. Some studies reported that after mixing methylene blue with 0.5% hydrogen peroxide or poly (lactic-co-glycolic acid) nanoparticles, there was a decrease in the number of colony-forming units compared with conventional PDT [7, 10].

In accordance with the present study's results, on activation of 0.05 mg/mL MB using a 670 nm diode laser for 30 sec on *E. faecalis* biofilm cultures, the results showed a significant decrease in bacterial account in PDT-treated groups [45]. While Soukos et al. applied PDT with longer treatment times (5 minutes), they eliminated nearly 95% of the bacterial biofilm [46].

Diode laser activation of nanocurcumin showed an insignificant difference compared to PDT, similarly to the study that evaluated the efficacy of antimicrobial activity for NaOCl activated by laser for 20 seconds, repeated three times, and PDT applied for 60 seconds using indocyanine green (12 mg/ml) as a photosensitizer with an 810 diode laser and 1.5 output power, showed no difference in both groups, although they used a lower wavelength [47]. Bago et al. activated 2.5% NaOCl by endoactivator, comparing its effect with PAD using diode laser (100 mW, 60 seconds) on the 7-day biofilm of *Enterococcus faecalis*. They concluded that PAD was significantly more effective than activated NaOCl irrigation in reducing CFUs ( $P < 0.05$ ) [34].

All nanoparticle herbal irrigants, together with PDT, were significantly effective compared to pretreatment samples. Literature found that a combination of photodynamic therapy and 2.5% NaOCl was effective in reducing *E. faecalis* biofilms [48, 49]. The combination of PDT with NaOCl offers many advantages, like faster permeation of the drug into the root canal for eliminating bacteria in a short time and the absence of thermal side effects in the nearby tissues [50].

In this study, PDT was performed at 650 nm and 200 mW for 60 sec, which was significantly better than nanoneem against *E. faecalis*. The same parameters were used by Asnaashari M., who reported an 80% decrease in the bacteria inside the canal compared to NaOCl [51].

Photoactivated therapy using 1.2 mg/L tolonium chloride with a 100 mW diode laser for 120 seconds against irrigation with 5 mL of 2.5% NaOCl showed that NaOCl was significantly better than PDT [52, 53, 54]. This may be due to the high susceptibility of the real-time PCR technique for detecting bacteria used in Janani's study, unlike the CFU used in the present study. In the same way, NaOCl was significantly better as an antibacterial agent (98% against *E. faecalis*) compared to PDT, which utilized a high output power of 2 W [55].

Nanoneem was activated for one minute to allow more penetration into dentinal tubules, unlike another study, which reported that 5 minutes of PDT and NaOCl were statistically significant compared with 1 minute of PDT. This may indicate that short irradiation time may be the cause of incomplete elimination of bacteria and consequently less amount of arisen ROS in dentinal tubules, where it is difficult for the photosensitizer agent to diffuse deeper [56]. So, the present results showed no statistically significant difference between nanocurcumin and methylene blue ( $p > 0.05$ ). While the differences were statistically highly significant between (nanoneem, methylene blue) ( $p = 0.002$ ) and between (nanoneem, nanocurcumin) ( $p < 0.001$ )

Nanoneem almost succeeded in eliminating all *C. albicans* from pretreatment groups, unlike nanocurcumin and PDT-MB, which were significantly effective against *C. albicans* but failed to remove all microorganisms from pretreatment groups; similarly, Oliveira et al. (2015, Eldeniz et al. (2015) reached the same results when comparing the effect of PDT with classical endodontic irrigation against *Candida albicans*. They concluded that incomplete microbial removal could be associated with several factors related to PDT, such as the type of chromophore, its concentration, the light source wavelength, the maximum absorption of each chromophore, and the time of irradiation [57, 58]. This coincided with the results of the other studies [59, 60].

No significant difference was found between activated nanocurcumin and PDT; the same results were obtained when comparing 1 and 5 minutes of PDT using photosensitizer phenothiazine chloride at a concentration of 10 mg/ml with an output power of 100 mW/cm<sup>2</sup> at 66 nm with the gold standard of irrigation of 2.5% NaOCl on *Candida albicans* [56].

PDT antifungal efficacy has been reported in the present study, as Wiench et al. evaluated this antifungal effect using a diode laser with a wavelength of 635 and toluidine blue (TB) as PS on selected *Candida* species cultured on an acrylic surface. They

used different parameters for laser (400 mW, 24 J/cm), (300 mW, 18 J/cm<sup>2</sup>), and (200 mW, 12 J/cm<sup>2</sup>) all for 30 sec using 8 mm optical fiber; the reduction in CFUs was statistically significant; using MB on an acrylic plate with parameters 660 nm laser (CW, 137 J/cm<sup>2</sup> and 480 sec.); obtaining the same results [61, 62]. Likewise, Ferreira et al. (2016) who used MB activated with a 660 nm diode laser with 690 mW and an energy of 60 J/cm<sup>2</sup> [63], and Kato et al. (2013) used the same parameter with a 10-minute exposure time, which led to the complete elimination of fungi [64].

Few in vivo studies performed for PDT using Methylene blue at 660 nm and energy 40 J combined with potassium iodide showed almost complete eradication of *C. albicans* [65].

Even when using the same PS and activated light or laser source, the variety of irradiation procedures, including time and light power, in addition to the difference in PS concentration, make comparisons between the literatures difficult [66].

There is a lack of studies about the laser activation of herbal irrigants and their nanoforms in comparison with standard endodontic irrigants.

## 5. Conclusion

Activation of nanoparticle herbal irrigants are effective against *E. faecalis* specially nanocurcumin, while nanoneem is powerfully effective against *C. albicans*. They can be used as adjunctive in cleaning process of endodontic treatment.

## 6. List of abbreviations

- NaOCl-----Sodium hypochlorite
- *E. faecalis*----- *Enterococcus faecalis*
- *C. albicans*----- *Candida albicans*
- SLN-----Solid lipid nanoparticles
- TEM----Transmission Electron Microscopy
- PCR----- Polymerase chain reaction
- *C. longa*----- *Curcuma longa*
- *A. indica*----- *Azadirachta indica*
- CFU----- Colony forming units
- FtsZ: Filamenting temperature-sensitive mutant Z

## 7. Ethics approval and consent to participate

In this study, ethical approval was obtained from the medical research ethics committee at the National Research Center in Egypt for the study titled Effect of photodynamic therapy on *Enterococcus faecalis* and *Candida albicans* compared to diode laser-activated nanoparticle herbal irrigants. Final approval number: (1483042021).

## 8. Competing interests

The authors declare no competing interests.

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## 11. References

1. Arslan S, Ozbilge H, Kaya EG, Er O. In vitro antimicrobial activity of propolis, BioPure MTAD, sodium hypochlorite, and chlorhexidine on *Enterococcus faecalis* and *Candida albicans*. *Saudi Med. J.* 2011;32(5):479-83.
2. Ramani N, Mathew S. Comparative evaluation of antimicrobial efficacy of Chlorhexidine digluconate and Propolis when used as intracanal medicament. *JIOH.* 2012;4 (2):17-23.
3. Love RM, Jenkinson HF (2002) Invasion of dentinal tubules by oral bacteria. *Crit. rev. oral biol. med.*. 2002;13:171-83.
4. Paque F, Boessler C, Zehnder M. Accumulated hard tissue debris levels in mesial roots of mandibular molars after sequential irrigation steps. *Int Endod J.* 2011;44:148-53.
5. Olivi G. Laser use in endodontics: evolution from direct laser irradiation to laser-activated irrigation. *J Laser Dent.* 2013;21:58-71.
6. Haapasalo M, Endal U, Zandi H, Coil JM. Eradication of endodontic infection by instrumentation and irrigation solutions. *Endod Top.* 2005;10(1):77-102.
7. Stojicic S, Amorim H, Shen Y, Haapasalo M. Ex vivo killing of *Enterococcus faecalis* and mixed plaque bacteria in planktonic and biofilm culture by modified photoactivated disinfection. *Int Endod J.* 2013;46:649-59.
8. Gutknecht N, Franzen R, Schippers M, Lampert F. Bactericidal effect of a 980-nm diode laser in the root canal wall dentin of bovine teeth. *J Clin Laser Med Surg.* 2004;22:9-13.
9. Hmud R, Kahler WA, George R, Walsh LJ. Cavitation effects in aqueous endodontic irrigants generated by near-infrared lasers. *J Endod* 2010;36(2):275-78.
10. Pagonis TC, Chen J, Fontana CR et al. Nanoparticle based endodontic antimicrobial photodynamic therapy. *J Endod.* 2010;36:322-8.
11. Plotino G., Grande N. M., Mercade M. Photodynamic therapy in endodontics. *Int. Endod. J.* 2019;52;(6):760-74.
12. Boehm TK, Ciancio SG. Diode laser activated indocyanine green selectively kills bacteria. *J Int Acad Periodontol.* 2011;13:58-63.
13. Susan AC, Bharathraj AR, Praveen M, Kumar NS, Karunakaran JV. Intraradicular smear removal efficacy of Triphala as a final rinse solution in curved canals: A scanning electron microscope study. *J Pharm Bioallied Sci.* 2019;11:S420-8.
14. Lakshmi T, Krishnan V, Rajendran R, et al. *Azadirachta indica*: a herbal panacea in dentistry – an update. *Pharmacogn Rev.* 2015;9(17):41-44.
15. John P, Gopalakrishnan, Dinesh K, Romel J. Herbal root canal irrigants: A review. *J Odontol Res.* 2015;3:9-14.
16. Afshan T, Parwez A, Prasanna PL, et al. Comparison of Antimicrobial Efficacy of Herbal Root Canal Irrigants (*Azadirachta indica*, *Morinda citrifolia*) against *Enterococcus faecalis*. *World J Dent.* 2020;11(3):206-10.
17. Sundaram D, Narayanan K, Vadakkepurayil K. A Comparative evaluation on antimicrobial effect of honey, neem leaf extract and Sodium Hypochlorite as intracanal irrigant: An ex-vivo study. *J Clin Diagn Res.* 2016;10:88-91.
18. Jose J, Shoba K, Aman S, Tomy N, Sheena, Christi. Comparative evaluation of antimicrobial activity of green tea extract, garlic extract, neem leaf extract and sodium hypochlorite as root canal irrigants against *E. faecalis* and *C. albicans* in vitro study. *Int J Curr Microbiol Appl Sci.* 2015;4:384-91.
19. Neelakantan P, Cheng CQ, Ravichandran V et al. Photo-activation of curcumin and sodium hypochlorite to enhance antibiofilm efficacy in root canal dentin. *Photodiagnosis Photodyn. Ther.* 2015;12:108-14.
20. Moghadamtousi SZ, Kadir HA, Hassan darvish P, Tajik H, Abubakar S, Zandi K. A review on antibacterial, antiviral, and antifungal activity of curcumin. *Biomed Res. Int.* 2014:186864. Epub 2014 Apr 29.
21. Gomes-Filho JE, Sivieri-Araujo G, Sipert CR et al. Evaluation of photodynamic therapy on fibroblast viability and cytokine production. *Photodiagnosis Photodyn. Ther.* 2016;13:97-100.
22. Mima EG, Pavarina AC, Dovigo LN, et al. Susceptibility of *Candida albicans* to photodynamic therapy in a murine model of oral candidosis. *Oral Surg Oral Med Oral Pathol Oral Radiol.* 2010;109:392-401.
23. Adel, M.M., Yoseif, N.S and Ibrahim, S.S.2018: Laboratory and Field Studies of Geranium Oil and GO-SLNs against the Cotton Leaf Worm *Spodoptera littoralis* (Boisd.) (Lep. Noctuidae). *International Review of Humanities and Scientific Research By International Scientific Indexing ISSN (Online): 2519-5336.*
24. El-boraey, A. N., Abo-Almaged, H. H., El-Ashmawy, A. A. E. R., Abdou, A. R., Moussa, A. R., Emara, L. H., & Ramzy, M. I. Biological and Mechanical Properties of Denture Base Material as a Vehicle for Novel Hydroxyapatite Nanoparticles Loaded with Drug. *Adv. Pharm. Bull.* 2021;11(1): 86.

25. Sultan, M., Nagieb, Z. A., El-Masry, H. M., & Taha, G. M. Physically-crosslinked hydroxyethyl cellulose-g-poly (acrylic acid-co-acrylamide)-Fe<sup>3+</sup>/silver nanoparticles for water disinfection and enhanced adsorption of basic methylene blue dye. *Int. J. Biol. Macromol.* 2022;196:180-93.
26. Konopka K, Goslinski T. Photodynamic therapy in dentistry. *J. Dent. Res.* 2007;86:694-707.
27. Shen Y, Stojicic S, Haapasalo M. Antimicrobial efficacy of chlorhexidine against bacteria in biofilms at different stages of development. *J Endod.* 2011;37(5):657-61.
28. R.C. Souza, J.C. Junqueira, R.D. Rossoni, C.A. Pereira, E. Munin, A.O. Jorge, Comparison of the photodynamic fungicidal efficacy of methylene blue, toluidine blue, malachite green and low-power laser irradiation alone against *Candida albicans*, *Lasers Med. Sci.* 2010;25(3):385-89.
29. Calzavara-Pinton P, Rossi MT, Sala R, Venturini M. Photodynamic antifungal chemotherapy. *Photochem Photobiol.* 2012;88:512-22.
30. Siddiqui SH, Awan KH, Javed F. Bactericidal efficacy of photodynamic therapy against *Enterococcus faecalis* in infected root canals: a systematic literature review. *Photodiagnosis Photodyn Ther.* 2013;10(4):632-43.
31. Silva EJ, Coutinho-filho WP, Andrade AO, Herrera DR, Coutinho-filho TS, Krebs RL. Evaluation of Photodynamic Therapy Using a Diode Laser and Different Photosensitizers against *Enterococcus faecalis*. *Acta Odontol Latinoam.* 2014;27(2):63-5.
32. Baltazar, L. M., Ray, A., Santos, D. A., Cisalpino, P. S., Friedman, A. J., and Nosanchuk, J. D. Antimicrobial photodynamic therapy: an effective alternative approach to control fungal infections. *Front. Microbiol.* 2015;6:202.
33. Pourhajibagher M, Chiniforush N, Raoofian R, Pourakbari B, Ghorbanzadeh R, Bazarjani F, et al. Evaluation of photo-activated disinfection effectiveness with methylene blue against *Porphyromonas gingivalis* involved in endodontic infection: An in vitro study. *Photodiagnosis Photodyn Ther.* 2016;16:132-5.
34. Bago I, Plečko V, Gabrić Pandurić D, Schauerperl Z, Baraba A, Anić I. Antimicrobial efficacy of a high-power diode laser, photo-activated disinfection, conventional and sonic activated irrigation during root canal treatment. *Int Endod J.* 2013;46(4):339-47.
35. Rios A, He J, Glickman GN, Spears R, Schneiderman ED, Honeyman AL. Evaluation of photodynamic therapy using a light-emitting diode lamp against *Enterococcus faecalis* in extracted human teeth. *J Endod.* 2011;37:856-9.
36. Bumb SS, Bhaskar DJ, Agali CR, Punia H, Gupta V, Singh V, Kadtane S, Chandra S, Garcia. Assessment of photodynamic therapy (PDT) in disinfection of deeper dentinal tubules in a root canal system: An in vitro study. *J Clin Diagn Res* 2014;8(11):67-71.
37. Pourhajibagher M, Bahador A. An in vivo evaluation of microbial diversity before and after the photo-activated disinfection in primary endodontic infections: traditional phenotypic and molecular approaches. *Photodiagnosis Photodyn Ther.* 2018;22:19-25.
38. Nagai Y, Suzuki A, Katsuragi H, Shinkai K. Effect of antimicrobial photodynamic therapy (aPDT) on the sterilization of infected dentin in vitro. *Odontology.* 2018;106:154-61.
39. Oda DF, Duarte MAH, Andrade FB, Moriyama LT, Bagnato VS, de Moraes IG. Antimicrobial action of photodynamic therapy in root canals using LED curing light, curcumin and carbopol gel. *Int Endod J.* 2019;52:1010-19.
40. Nardini EF, Almeida TS, Yoshimura TM, Ribeiro MS, Cardoso RJ, Garcez AS. The potential of commercially available phytotherapeutic compounds as new photosensitizers for dental antimicrobial PDT: A photochemical and photobiological in vitro study. *Photodiagnosis Photodyn Ther.* 2019;27:248-54.
41. Mozayeni MA, Vatandoost F, Asnaashari M, Shokri M, Azari-Marhabi S, Asnaashari N. Comparing the efficacy of toluidine blue, methylene blue and curcumin in photodynamic therapy against *Enterococcus faecalis*. *J Lasers Med Sci.* 2020;11:S49-S54.
42. Hendi SS, Shiri M, Poormoradi B, Alikhani MY, Afshar S, Farmani A. Antibacterial Effects of a 940 nm Diode Laser With/Without Silver Nanoparticles Against *Enterococcus faecalis*. *J Lasers Med Sci.* 2021;12:73.
43. Vaziri S, Kangarlou A, Shabbazi R, Nazari Nasab A, Naseri M. Comparison of the bactericidal efficacy of photodynamic therapy, 2.5% sodium hypochlorite, and 2% chlorhexidine against *Enterococcus faecalis* in root canals; an in vitro study. *Dent Res J (Isfahan).* 2012 Sep;9(5):613-8.
44. Ng R, Singh F, Papamanou DA, Song X, Patel C, Holewa C, Patel N, Klepac-Ceraj V, Fontana CR, Kent R, Pagonis TC, Stashenko PP, Soukos NS. Endodontic photodynamic therapy ex vivo. *J Endod.* 2011;37(2):217-22.
45. López-Jiménez L, Fusté E, Martínez-Garriga B, Arnabat-Domínguez J, Vinuesa T, Viñas M. Effects of photodynamic therapy on *Enterococcus faecalis* biofilms. *Lasers Med Sci.* 2015 Jul;30(5):1519-26.
46. Prazmo EJ, Godlewska RA, Mielczarek AB. Effectiveness of repeated photodynamic therapy in the elimination of intracanal *Enterococcus*



- faecalis biofilm: an in vitro study. *Lasers Med Sci.* 2017;32(3):655–61.
47. Attiguppe PR, Tewani KK, Naik SV, Yavagal CM, Nadig B. Comparative Evaluation of Different Modes of Laser Assisted Endodontics in Primary Teeth: An In vitro Study. *J Clin Diagn Res.* 2017;11(4):124-27..
48. Zand V, Milani AS, Amini M, Barhaghi MHS, Lotfi M, Rikhtegaran S, et al. Antimicrobial efficacy of photodynamic therapy and sodium hypochlorite on monoculture biofilms of *Enterococcus faecalis* at different stages of development. *Photomedicine and laser surgery.* 2014;32:245-51.
49. Soares JA, Soares SMCS, de Jesus Tavarez RR, de Castro Rizzi C, Vaz Rodrigues SCG, Maia Filho EM, et al. Exploring different photodynamic therapy parameters to optimize elimination of *Enterococcus faecalis* in planktonic form. *Photodiagnosis Photodynamic Ther.* 2018;22:127-31.
50. Vaid D, Shah N, Kothari D, Bilgi P. Additive effect of photoactivated disinfection on the antibacterial activity of QMix 2in1 against 6-week *Enterococcus faecalis* biofilms: An in vitro study. *J Conserv Dent.* 2017;20:41-5.
51. Asnaashari M, Kooshki N, Salehi MM, Azari-Marhabi S, Amin Moghadassi H. Comparison of antibacterial effects of photodynamic therapy and an irrigation activation system on root canals infected with *Enterococcus faecalis* : an in vitro study. *J Lasers Med Sci.* 2020;11(3):243-48.
52. Janani M, Jafari F, Samiei M, Lotfipour F, Nakhband A, Ghasemi N, Salari T. Evaluation of Antibacterial Efficacy of Photodynamic Therapy vs. 2.5% NaOCl against *E. faecalis*-infected Root Canals Using Real-time PCR Technique. *J Clin Exp Dent.* 2017;9(4):539-44.
53. De Frota MF, Guerreiro-Tanomaru JM, Tanomaru-Filho M, Bagnato VS, Espir CG, Berbert FLCV. Photodynamic therapy in root canals contaminated with *Enterococcus faecalis* using curcumin as photosensitizer. *Lasers Med Sci.* 2015;30:1867-72.
54. Marinic K, Manoil D, Filieri A, Wataha JC, Schrenzel J, Lange N, et al. Repeated exposures to blue light-activated eosin Y enhance inactivation of *E. faecalis* biofilms, in vitro. *Photodiagnosis Photodyn Ther.* 2015;12:393-400.
55. Sarda RA, Shetty RM, Tamrakar A, Shetty SY. Antimicrobial Efficacy of Photodynamic Therapy, Diode Laser, and Sodium Hypochlorite and Their Combinations on Endodontic Pathogens. *Photodiagnosis Photodyn Ther.* 2019;28:265-72.
56. Xhevdet A, Stubljarić D, Kriznar I, Jukic T, Skvarc M, Veranic P, Ihan A. The Disinfecting Efficacy of Root Canals with Laser Photodynamic Therapy. *J Lasers Med Sci.* 2014;5(1):19-26
57. Oliveira BP, Aguiar CM, Câmara AC, de Albuquerque MM, Correia ACR de B, Soares MF de LR. The efficacy of photodynamic therapy and sodium hypochlorite in root canal disinfection by a single-file instrumentation technique. *Photodiagnosis Photodyn Ther.* 2015;12(3):436–43.
58. Eldeniz AU, Guner MB, Akbulut MB. Comparative antifungal efficacy of light-activated disinfection and octenidine hydrochloride with contemporary endodontic irrigants. *Lasers Med Sci.* 2015;30(2):669–75.
59. Sabino CP, Garcez AS, Núñez SC, Ribeiro MS, Hamblin MR. Real-time evaluation of two light delivery systems for photodynamic disinfection of *Candida albicans* biofilm in curved root canals. *Lasers Med Sci.* 2015;30(6):1657-65.
60. Oliveira BP de, Lins CCSA, Diniz FA, Melo LL, Castro CMMB de. In Vitro antimicrobial photoinactivation with methylene blue in different microorganisms. *Braz J Oral Sci.* 2014;13:53-7.
61. Wiench R, Skaba D, Stefanik N, Kępa M, Gilowski Ł, Cieślak G, Kawczyk Krupka A. Assessment of sensitivity of selected *Candida* strains on antimicrobial photodynamic therapy using diode laser 635 nm and toluidine blue - In vitro research. *Photodiagnosis Photodyn Ther.* 2019;27:241-47.
62. A.S. Sousa, R.A. Prates, M.E. de Santi, R.G. Lopes, S.K. Bussadori, L.R. Ferreira, A.M. Deana, Photodynamic inactivation of *Candida albicans* biofilm: influence of the radiant energy and photosensitizer charge, *Photodiagnosis Photodyn Ther.* 2016;14:111–14.
63. L.R. Ferreira, A.S. Sousa, L.H. Alvarenga, A.M. Deana, M.E.O. Simões de Santi, I.T. Kato, C.R.L. Leal, M.S. Ribeiro, R.A. Prates, Antimicrobial photodynamic therapy on *Candida albicans* pre-treated by fluconazole delayed yeast inactivation, *Photodiagnosis Photodyn Ther.* 2016;15: 25–7.
64. I.T. Kato, R.A. Prates, C.P. Sabino, B.B. Fuchs, G.P. Tegos, E. Mylonakis, M.R. Hamblin, M. Simões Ribeiro. Antimicrobial photodynamic inactivation inhibits *Candida albicans* virulence factors and reduces in vivo pathogenicity. *Antimicrob Agents Chemother.* 2013; 57(1):445–51.
65. F. Freire, C. Ferraresi, A.O.C. Jorge, M.R. Hamblin, Photodynamic therapy of oral *Candida* infection in a mouse model. *J. Photochem. Photobiol.* 2016;159:161–68.
66. Nagata JY, Hioka N, Kimura E et al. Antibacterial photodynamic therapy for dental caries: evaluation of the photosensitizers used and light

source properties. *Photodiagnosis Photodyn. Ther.*  
2012;9:122–31.

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