

**Egyptian Journal of Chemistry** 



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# Computer modeling, docking, spectroscopic analysis, and antibacterial testing of metal chelates with dioxatetraaza ligand.

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## Abstract

The goal of our current research is to create novel compounds of Co(II), Ni(II), Cu(II), Cr(III), Fe(III) and Zn(II) metal ions from heterocyclic dioxatetraaza ligand by the reaction of N1,N1'-(ethane-1,2-diyl)bis(ethane-1,2-diamine)with 2,2'-(ethane-1,2-diyl)bis(oxy))dibenzaldehyde to find out combating agent against microorganism diseases. Various physicochemical and spectroscopic techniques, including FT-IR, 1H and magnetic susceptibility, mass spectrometry, TGA, and others, were used to precisely characterize the synthesized compounds. These techniques suggested that the complexes' octahedral geometry was formed by epoxy oxygen atoms, azomethine nitrogen atomsof the dioxatetraaza ligand, and oxygen from the water molecules. The compounds' in vitro antibacterial activity against two bacterial and fungal pathogens was evaluated using the serial dilution method. Theoretical molecular docking was used to confirm these findings.

Keywords: dioxatetraaza; physicochemical technique; Microorganism; Antimicrobial activity; Transition metal chelates

#### Introduction

In the past, microbial diseases have been responsible for a sizably high number of mortality worldwide. Since the discovery of penicillin as a strong antibacterial agent in the 1940s, the use of numerous natural and synthetic antibiotics has fortunately significantly aided human health[1]. The rapid proliferation of infectious diseases and the rise in the number of multidrug-resistant microorganisms make it difficult to treat bacterial infections. Due to shortcomings including antagonistic interactions, a lack of diversity, and a lack of disease-modifying agents in hospitals, currently existing treatments have lost their efficacy, contributing to an ongoing global epidemic that has a negative impact on people's health and the economy of their countries[2-5]. Therefore, one of the most important tasks in medicinal chemistry today is the hunt for new, highly effective therapeutic molecules with minimal side effects. As a result, we determined that transition metal complexes of dioxatetraaza ligands

offer a great possibility for the study and development of new active antimicrobial medicines that can efficiently create pathogenic deformities with the fewest adverse effects [5-7]. The structural diversity and activity of metal complexes are increased when dioxatetraaza ligands are bound to transition metal ions. This encourages researchers to create new metal complexes using dioxatetraaza ligands. They can alter the charge, substitution kinetics, lipophilicity, and mode of action of biological targets by using a variety of threedimensional geometries and potentially infinite design options for their coordination sphere. They are widely used as analytical reagents, fluorescent materials, and polymer coating pigments for coating polymers due to their prominence as chelation abilities, reactivity, preparative accessibilities, and flexibility as they contain -NH-C=O group in proximity to isocyanic acid, which makes them suitable for coordination with transition metal atoms [8-11], transition metal complexes with

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Receive Date: 15 October 2023, Revise Date: 23 November 2023, Accept Date: 27 November 2023 DOI: 10.21608/EJCHEM.2023.238209.8653

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dioxatetraaza occupy a main position in medicinal chemistry. Biological uses of transition metal complexes with dioxatetraaza ligands include antifungal [12], antibacterial, anticancer, antiplatelets, and antimalarial [13].antioxidants, antiradicals, antitubercular [14], anti-inflammatory, etc. Additionally, transition metal complexes with dioxatetraazaligand are gaining attention from researchers due to their variety of pharmacological and biological applications, but it is challenging for them to develop substantial pharmaceutical improved pharmacological compounds with applications, They are widely used as analytical reagents, fluorescent materials, and polymer coating pigments for coating polymers due to their prominence as chelation abilities, reactivity, preparative accessibilities, and flexibility as they contain -NH-C=O group in proximity to azomethine group, which makes them suitable for coordination with transition metal atoms[15-19].

## Experimental

The earlier work covered every detail in the experimental portion. [20]

## Materials and reagents

All chemicals were of the purest and highest analytical reagent grade (AR). CoCl<sub>2</sub>.6H<sub>2</sub>O (BDH), CuCl<sub>2</sub>.2H<sub>2</sub>O (BDH), NiCl<sub>2</sub>.6H<sub>2</sub>O (BDH), ZnCl<sub>2</sub>.2H<sub>2</sub>O (BDH), and CrCl<sub>3</sub>.6H<sub>2</sub>O (BDH)m was Prolabo. Without additional provided by purification, organic solvents like EtOH and DMF were employed as received.

## Solutions

In order to make prepared metal complex stock solutions, a carefully weighed quantity of the complex was dissolved in EtOH and DMF (1:3 v/v ratio) to obtain 1×10-3M. We created a stock solution of the ligand and its metal complexes (5  $\times$ 10-4 M) through dilution in order to examine their UV-Vis spectra.

# Synthesis of dioxatetraaza Ligand

Condensation 2-[2-(2of formylphenoxy)ethoxy]benzaldehyde with triethylene tetra amine gives the corresponding cyclic Schiff base as identified by IR, 1HNMR and mass spectra. Filtering was used to collect the solid that formed after cooling, and compound 1 was created as buff crystals by recrystallizing toluene. Yield: 81%. M.p.: 110 °C.FT-IR (KBr, cm-1): 3036m v(NH), 829s8 (NH), 1597sh v(HC=N) and 1049m υ(C-O-C). 1H-NMR (500 MHz. DMSO):2.654~2.975, (m, 6H, -NH-CH2); 4.429 (m, 2H, -0CH2); 3.545 (m, 1H, NH); 6.840~7.977 (m, 4H, ArH). MS (EI, m/z): f, 380 (M+), calc, 380 g/mol. Anal.Calcd.for C22H28N4O2: C,69.45; H, 7.42; N, 14.73. Found: C, 69.40; H, 7.38; N,14.69.

Synthesis of metal complexes

The Ni(II), Co(II), Cu(II), Cr(III), Zn(II) and Fe(III) Equivalent amounts of the ligand and metal chloride ratio were mixed (IM: 1L molar ratio) in EtOH, which was then heated for 3h. to create complexes. After filtering the resulting precipitates, the filtrates were repeatedly washed with hot ethanol until they were clear, yielding, respectively, 96, 80, 89, 82, 83, and 80 percent of Co(II), Ni(II), Cu(II), Zn(II),Cr(III), and Fe(III) complexes. The appropriate products were then dried in a desiccator over anhydrous CaCl<sub>2</sub>. [Cr(L)(H<sub>2</sub>O)<sub>2</sub>]Cl<sub>3</sub>; yield 83%; m.p.118 °C; Brown solid. Anal. Calc. for C<sub>22</sub>H<sub>32</sub>Cl<sub>3</sub>CrN<sub>4</sub>O<sub>4</sub>(%):C, 45.96; H, 5.61; N, 9.75;Cl, 18.50; M, 9.04. Found (%): C, 45.92; H, 5.57; N, 9.71; Cl, 18.45; M, 9.01.FT-IR (KBr, v, cm<sup>-1</sup>):2920m 815mδ(NH), υ(NH), 1604sh υ(HC=N), 1040mu(C-O-C),519wv(M-O)<sub>H2O</sub>, 540sv(M-O)<sub>ether</sub>,467sv(M-N)<sub>azo</sub>. $\mu_{eff}$  (BM) 3.48;  $\Lambda_m$  $(\Omega^{-1} \text{mol}^{-1} \text{cm}^2)$  161.8.  $[Fe(L)(H_2O)_2]Cl_3$ ; yield 83%; m.p. 110 °C: Yellowish Brown solid. Anal. Calc. for C<sub>22</sub>H<sub>32</sub>Cl<sub>3</sub>FeN<sub>4</sub>O<sub>4</sub>(%):C, 45.66; H, 5.57; N, 9.68;Cl, 18.38; M, 9.65. Found (%): C, 45.62; H, 5.52; N, 9.64; Cl, 18.34; M, 9.61. FT-IR (KBr, v, cm<sup>-1</sup>):2932sh υ(NH), 833mδ(NH), 1610shυ(HC=N), v(C-O-C),470wv(M-O)<sub>H2O</sub>,520sv(M-1039sh 0)<sub>ether</sub>,462sv(M-N)<sub>azo</sub>. $\mu_{eff}$ (BM) 5.34;  $\Lambda_{\rm m}$  $(\Omega^{-1} \text{mol}^{-1} \text{cm}^2)$  166.0.  $[Co(L)(H_2O)_2]Cl_2$ ; yield 96%; m.p.280 °C; Dark Greensolid. Anal. Calc. for C<sub>22</sub>H<sub>32</sub>Cl<sub>2</sub>CoN<sub>4</sub>O<sub>4</sub>(%):C,48.36; H, 5.90; N, 10.25;Cl, 12.98; M, 10.79. Found (%): C, 48.32; H, 5.85; N, 10.22; Cl, 12.95; M, 10.72. FT-IR (KBr, v, cm<sup>-1</sup>):2980s υ(NH), 800sδ(NH), 1643sh υ(HC=N), υ(C-O-C),443wv(M-O)<sub>H2O</sub>, 1056m 513sv(M-0)<sub>ether</sub>,  $417 sv(M-N)_{azo}$ .  $\mu_{eff}$ (BM) 5.02;  $\Lambda_{\rm m}$  $(\Omega^{-1} \text{mol}^{-1} \text{cm}^2)$  98.7. [Ni(L)(H<sub>2</sub>O)<sub>2</sub>] Cl<sub>2</sub>; yield 80%; m.p. 104 °C; Brown solid. Anal. Calc. for C<sub>22</sub>H<sub>32</sub>Cl<sub>2</sub>NiN<sub>4</sub>O<sub>4</sub>(%):C, 48.38; H, 5.91; N, 10.26;Cl, 12.98; M, 10.75. Found (%): C, 48.35; H, 5.87; N, 10.22; Cl, 12.92; M, 10.71. FT-IR (KBr, ν, cm<sup>-1</sup>):2928m υ(NH), 822s δ(NH), 1590shu(HC=N), 1055shu(C-O-C),444sv(M-O)<sub>H2O</sub>,  $522sv(M-0)_{ether}, 418wv(M-N)_{azo}, \mu_{eff}$  (BM) 3.44;  $\Lambda_m$  $(\Omega^{-1} \text{mol}^{-1} \text{cm}^2)$  104.3. [Cu(L)(H<sub>2</sub>O)<sub>2</sub>]Cl<sub>2</sub>; yield 89%; m.p.250 °C; Brown solid. Anal. Calc. for C<sub>22</sub>H<sub>32</sub>Cl<sub>2</sub>CuN<sub>4</sub>O<sub>4</sub>(%):C, 47.96; H, 5.85; N, 10.17;Cl, 12.87; M, 11.53. Found (%): C, 47.92; H, 5.81; N, 10.12; Cl, 12.82; M, 11.50. FT-IR (KBr,  $\nu$ , cm<sup>-1</sup>):3067m  $\nu$ (NH), 810sδ(NH), 1601shv(HC=N), 1045m v(C-O-C),

(BM) 1.77;  $\Lambda_{\rm m}$  ( $\Omega^{-1}$ mol<sup>-1</sup>cm<sup>2</sup>) 100.7. Zn(L)(H<sub>2</sub>O)<sub>2</sub>]Cl<sub>2</sub>; yield 82%; m.p. 170 °C; Honey Brown solid. Anal. Calc. for  $C_{22}H_{32}Cl_2ZnN_4O_4(\%):C, 47.80; H, 5.83; N,$ 10.14;Cl, 12.83; M, 11.83. Found (%): C, 47.75; H,

 $517 \text{sv}(\text{M-O})_{\text{H2O}}$ ,  $530 \text{sv}(\text{M-O})_{\text{ether}}$ ,  $470 \text{sv}(\text{M-N})_{\text{azo}}$ ,  $\mu_{\text{eff}}$ 

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5.80; N, 10.10; Cl, 12.79; M, 11.78. FT-IR (KBr, v, cm<sup>-1</sup>):2928shv(NH), 830s\delta(NH), 1605m v(HC=N) and 1053m v(C-O-C),471wv(M-O)\_{H2O}, 511sv(M-O)\_{ether},460wv(M-N)\_{azo}.\mu\_{eff} (BM) diamagnetic; $\Lambda_m$  ( $\Omega^{-1}mol^{-1}cm^2$ ) 100.3.

# Spectrophotometric studies

Over the wavelength range of 200 to 700 nm, the absorption spectra of  $1 \times 10-4$  M solutions of dioxatetraaza ligand and metal complexes were measured.

#### Molecular docking

Both Auto Dock 4.2 and docking calculations with the ligand (designed drug) atoms subjected to Gasteiger partial charges were used. Calculations of the ligand-protein pattern were done. Clarifying rotatable bonds and linking nonpolar hydrogen atoms. Kollmanunified atom type charges and solvation parameters were added using the Auto Dock tools after the introduction of fundamental hydrogen atoms [21]. The distance-dependent dielectric functions and the Auto Dock parameter set, respectively, were used to calculate the van der Waals and electrostatic terms. The Lamarckian genetic algorithm was used to mimic docking using the Solis and Wets local search approach. The beginning location, orientation, and torsion of the ligand molecule were all identified.

#### **Biological activity**

Testing for new antibacterial and antifungal compounds' susceptibility to microbes can be used to foretell how a treatment will work. The Clinical and Laboratory Standards Institute (CLSI) states that one of the manual AST techniques that is most frequently employed in clinical microbiology laboratories is the agar disc diffusion test [22], and it was used to test the evaluated substances for their in vitro antibacterial and antifungal activity. The main advantages include the test's simplicity, reproducibility, ease of customizing Antibacterial and Antifungal discs, and ability to be used as a screening test for a range of Antibacterial and Antifungal isolates. Mueller-Hinton agar plates are inoculated with tested antibacterial and antifungal inoculums that are standardized species; Aspergillus flavus(Fungus)(7BOP),

Candidaalbicans(Fungus)(5k04), Escherichia coli(G-)(3t88), and Staphylococcus aureus(G+)(3ty7) are examples of host organisms that can be used to attach ligand and complexes (guests). Each disc was placed on the inoculated agar surface using commercially prepared paper discs (about 6 mm in diameter) impregnated with 100  $\mu$ L of the required concentration of the tested drug. According to appropriate guidelines, agar plates are incubated for 16–24 hours at 35–37°C (CLSI2018a; EUCAST 2019b). The diameter of the clear inhibition zones

surrounding each compound-impregnated disc is then measured in millimetres and factored into the outcome. This is done by hand while holding a ruler against the back of the inverted agar plate [23-25]. Tetracycline and amphotericin Bwere utilized as reference drugs standard for Gram-positive, Gramnegative, and fungal activities, whereas DMSO was employed as negative control. The outcomes of all the inoculation plates were examined in a table after 35°C incubation. The MIC values have an inverse relationship with the inhibitory zone. The larger the zone of growth inhibition, the lower the antimicrobial medication concentration required to stop the growth. However, it is important to consider a compound's diffusibility[26].

## **Computational methodology**

Gaussian09 software was used to calculate the ligand's ideal structural geometry using the Ground state, DFT, B3LYP, and 3-21G. Gauss View, a molecular visualisation programme, was used to display Gaussian files [27]. In the view of compounds in the gas phase, the numerical pattern was consistent with the HOMO-LUMO energies used to construct the DFT/B3LYP quantum chemical parameters. Calculations were made for coordinating group charges, significant bond lengths, bond angles, dihedral angles, and excitation energy in optimised structures.

#### **Results and discussion**

The synthesis of dioxatetraaza cyclic ligand was reactingN1,N1'-(ethane-1,2carried out by diyl)bis(ethane-1,2-diamine) with 2,2'-(ethane-1,2diylbis(oxy))dibenzaldehydein hot methanol with the addition of 0.1mL of glacial acetic acid. Further, complexation of the ligand was carried out from the reaction of metal(II) and metal(III) chlorides with synthesized dioxatetraaza ligand in 1:1 molar ratio. The analytical techniques revealed that synthesized dioxatetraaza ligand was bonded to metal ions via the azomethine nitrogen atoms, etheric oxygen atoms and NH atoms resulting in octahedral geometry. The compounds were examined by several spectral and physical techniques to ascertain the geometry of the complexes and other characteristic data. With the exception of DMF and DMSO, the compounds were not soluble in the majority of solvents.

#### Mass spectral study

To support the above-proposed structure, the mass spectrum of the dioxatetraaza ligand was examined and recorded at 70 eV. The molecular ion of the dioxatetraaza ligand (molar mass = 380.22 g/mol) is referenced by a peak at m/z = 380 amu. This result confirmed the suggested empirical formula of the dioxatetraaza ligand as indicated from elemental analyses. The primary molecular ion and its putative

fragment ions are produced by the dioxatetraaza ligand's cleavage of various bonds in various places, as indicated in Scheme 1.The mass spectrum showed fragment ions at m/z = 324.42, 283.36,242.31, 136.19,106.16 and 78.11 amu which may be assigned to C20H24N2O2, C18H21NO2, C16H18O2,C9H12O, C8H10 andC6H6 fragments



Scheme 1.Mass fragmentation of ligand

#### **IR** spectral study

With the exception of a few modest shifts and intensity changes of a few vibration peaks brought on by various metal (III/II) ions, the IR spectra of the metal complexes are similar to one another, showing that the metal complexes have a similar structure. The most significant IR bands of Schiff base ligand and its metal complexes along with their likely intensities were given in the experimental part. The ligand has many sites for coordination, which resulted in a variety of coordination modes. When compared to the free ligand, the IR spectra of all complexes exhibit a shift in the band of the azomethine N, v(C=N), from 1590 to 1643(1597 cm-1 in the free ligand) confirming its involvement in chelation [28]. The band due to the etheric O, v(C-O-C), found at 1049 cm-1 in the free ligand IR spectrum was found at 1039-1056 cm-1 in the metal complexes IR spectra. This shift can be assigned to the participation of etheric oxygen in the coordination. The shift in the v(NH) or  $\Box$ (NH) towards lower or higher frequencies can be attributed to the change in the skeleton of the Schiff base ligand due to chelation to metal ions [29]. Additionally, the new bands between the areas of 417-470 cm-1, 511-540 cm-1 and 417-519 cm-1 can be attributed to v(M-N), v(M-O)etheric and v(M- of the parent Schiff base ligand. The suggested scheme lists potential fragment ion structural equations together with potential names using the IUPAC system. Thermal degradation of the dioxatetraaza ligand examined at two distinct heating rates has corroborated these fragmentations.

O)water, respectively[30-32]. This leads us to the conclusion that the complexes' coordination geometry, which includes two nitrogen and two oxygen donors of the macrocyclic Schiff base ligand and the two oxygen atoms from water molecules, is distorted octahedral, indicating that the Schiff base functions as a tetradentate neutral ligand.

## <sup>1</sup>H-NMR spectral study

Schiff base's 1H-NMR spectrum in d6-DMS0 exhibited signal at 8.53 ppm which assigned to azomethine CH=N proton. As a result of its participation in the chelation mode, its position in the Zn(II) complex has shifted to 7.95 ppm. The signal found at 3.55 ppm which assigned to NH proton for the Schiff base ligand, as a result of change in the carbon skeleton due to chelation, its position in the Zn(II) complex1H-NMR spectrum has shifted to 3.32 ppm. Additionally, at 6.84–7.98 ppm in the free ligand and 7.12–7.15 ppm in the Zn(II) complex, numerous signals attributed to aromatic ring protons were detected [33].

# Thermogravimetric analysis study

results Table 1 contains the of the thermogravimetric study of the Schiff base ligand and its metal complexes. The experimental findings showed that ligand degradation follows a complicated mechanism and occurs in a number of phases. For the Schiff basedioxatetraaza ligand, the first two estimated mass losses of 13.50% (calcd. 13.42%) in the range of 175~405 °C may be attributed to the liberation of 3NH<sub>3</sub> as gases, and in the following stages the remaining organic part ( $C_{21}H_{19}$  molecule),  $CO_2$  and  $\frac{1}{2}N_2$  gases are lost with an estimated mass loss of 86.50% (calcd. 86.58%) with a complete decomposition within the temperature range from 410 to ~935 °C. In contrast, the Fe(III), Co(II), and Cu(II) chelates displayed four steps of decomposition between 25 - 790 °C, 45 - 630 °C, and 35 - 830 °C. These decomposition can be attributed to the loss of water molecules, anions present in the outer coordination sphere and decomposition of the Schiff base ligand to gases (Table 1). The overall weight losses of 86.90% (calculated as 86.76%), 84.91% (calculated as 84.91%), and 84.91% (calculated as 86.76%), respectively, were reported for Fe(III), Co(II) and Cu(II) complexes.FeO (estimated mass loss = 13.10; calculated mass loss = 13.24), CoO (estimated mass loss = 15.09; calculated mass loss = 14.69), and CuO (estimated mass loss = 15.29; calculated mass

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loss = 15.46) were the metal oxides left over after the complexes broke down.

The Coats-Redfern relation (equation 1) is used to graphically analyse the activation energies  $(E^*)$ , enthalpies  $(H^*)$ , Gibbs free energy change of the decomposition  $(G^*)$  and entropies  $(S^*)$  of the thermodynamic processes that lead to the breakdown of dehydrated complexes:

$$\log \left[ \frac{\log \{W_{f}/(W_{f}-W)\}}{T^{2}} \right] = \log \left[ \frac{AR}{\Theta E^{*}} \left( 1 - \frac{2RT}{E^{*}} \right) \right] - \frac{E^{*}}{2.303 \text{ RT}}$$
(1)

Where Wf is the mass loss at reaction completion, R is the gas constant, and is the heating rate W is the

mass loss up to temperature T, E\* is the activation energy in kJ.mol-1, and  $(1-(2RT/E^*)) \cong 1$ .The slope from which the left side of equation (1) is plotted against 1/T can be used to compute E\*, and the intercept can be used to get A (Arrhenius factor).According to the Coats-Redfern equation [34], the complexes' high activation energies represent their thermal stability, and the fact that all of the complexes' activation entropies are negative means that the breakdown reactions happen more slowly than they would normally.

| Table 1 Thermoanal    | vtical results  | TG and DTG | ) for Schiff base | dioxatetraaza l | ligand and its | metal com- | plexes |
|-----------------------|-----------------|------------|-------------------|-----------------|----------------|------------|--------|
| . I dole I.I nermound | y ficul results |            | , for beinn buse  | ulonulouuuzu    | inguna una no  | metul com  | pienes |

| Compound                 | TGrange/°C | DTGmax/°C   | n* | Mass loss(Total mass     | Assignment  | Metallic     |
|--------------------------|------------|-------------|----|--------------------------|---|--------------|
|                          | U          |             |    | loss)                    | -   | Residue/%    |
| $H_2L$                   | 175-405    | 320,383     | 2  | 13.42(13.50)             | -Lossof3NH <sub>3</sub>   | -            |
|                          | 410-935    | 421,640,861 | 3  | 86.58(86.50),100(100)    | -Loss of CO2,1/2N2and C21H19                                    |              |
| $[Fe(H_2L)(H_2O)_2]Cl_3$ | 25-350     | 76,296      | 2  | 12.82(12.76)             | -LossofHCl,NOand <sup>1</sup> / <sub>2</sub> H <sub>2</sub>     | FeO          |
|                          | 350-465    | 394         | 1  | 16.04(15.99)             | -Lossof2HCland <sup>1</sup> / <sub>2</sub> N <sub>2</sub>       |              |
|                          | 465-790    | 671         | 1  | 57.90(58.15)86.76(86.90) | -LossofC <sub>22</sub> H <sub>22</sub> N <sub>2</sub>           | 13.24(13.10) |
| $[Co(H_2L)(H_2O)_2]Cl_2$ | 45-350     | 81,317      | 2  | 18.43(18.42)             | -Lossof2HCl,CH4and <sup>1</sup> / <sub>2</sub> H <sub>2</sub>   | CoO          |
|                          | 350-630    | 433,568     | 2  | 66.88(66.49),            | -Lossof4NH <sub>3</sub> ,COandC <sub>20</sub> H <sub>5</sub>    | 14.69(15.09) |
|                          |            |             |    | 85.31(84.91)             |   |              |
| $[Cu(H_2L)(H_2O)_2]Cl_2$ | 35-125     | 62          | 1  | 5.44(5.59)               | -LossofN <sub>2</sub>   | CuO          |
|                          | 125-320    | 212         | 1  | 10.20(10.01)             | -LossofHCl and <sup>1</sup> / <sub>2</sub> O <sub>2</sub>       | 15.45(15.29) |
|                          | 320-480    | 346         | 1  | 17.98(18.04)             | -LossofHCl,2NH <sub>3</sub> ,CH <sub>4</sub> and3H <sub>2</sub> |              |
|                          | 480-830    | 590         | 1  | 50.92(51.07),            | -LossofC <sub>21</sub> H <sub>10</sub>                          |              |
|                          |            |             |    | 84.54(84.71)             |   |              |

Table2.Calculated quantum chemical parameters of dioxatetraaza ligand.

| Parameter                |             |
|--------------------------|-------------|
| E <sub>HOMO</sub> (a.u.) | -0.23083    |
| E <sub>LUMO</sub> (a.u.) | 0.00838     |
| μ(D)                     | -39.4251    |
| T.E (a.u.)               | -1217.85    |
| $\Delta E$ (a.u.)        | 0.23921     |
| χ (a.u.)                 | 0.111225    |
| η (a.u.)                 | 0.119605    |
| $\sigma(a.u.)^{-1}$      | 8.360854    |
| Pi (a.u.)                | -0.11123    |
| S (a.u.) <sup>-1</sup>   | 4.180427    |
| ω(a.u.)                  | 0.05171607  |
| $\Delta N_{max}$         | 0.929936039 |

#### Molecular modeling

The geometrical geometry of the dioxatetraaza ligand as well as a theoretical tool for molecular modeling was both created using the Gaussian09 program. The electronic structure of the dioxatetraaza ligand can be determined using the separation in orbital energy between EHOMO and ELUMO [19] Examples of the molecular orbitals of the free dioxatetraaza ligand show that the donor atoms—nitrogen of the azomethine group and oxygen of the epoxy group—that are employed to donate to metal ion acceptors were mainly focused

LUMO, HOMO, the and charge on distribution.(Figure 1). Table2 contains chemical calculations and the data that were obtained. The dipole moment  $(\mu)$  and other parameters like the HOMO-LUMO energy gap (E), chemical potential (Pi), absolute electronegativity (E), absolute hardness (H), additional electronic charge ( $\Delta$ Nmax), absolute softness (S), and global softness (E) were calculated. The first derivative of the energy with respect to an applied electric field was utilized to evaluate and explain the proposed structure. One of the theoretical models used was the energy gap (E), a significant stability parameter that aids in explaining the structures and conformational barriers in many compounds. Values were derived using previously published equations. [35-37]. The free dioxatetraaza ligand was revealed to have the following properties following the determination of all the parameters (Table 2). The soft property of the free ligand was used to infer the flexible reactions to metal ions.

#### Atomic charges, bond lengths and bond angles

Atomic charge calculations provide the foundation of quantum mechanical computation [38].In the supplemental material (Supplementary Tables S1 and S2), the results of the charges population research with optimised geometry used to calculate the total atomic charge are presented. The two promising atoms in the dioxatetraaza ligand are nitrogen and oxygen. This demonstrates that a high electron density, particularly in the region surrounding dioxatetraaza, is what causes the more active site, which promises to produce chelation.



Figure1.LUMO and HOMO patterns of studied dioxatetraaza ligand.

Figure2 showed the numbering sequence of the dioxatetraaza ligand structure. It may be demonstrated that nitrogen and oxygen atoms melt at higher temperatures than atoms of other sizes do, suggesting that these atoms may be able to produce coordination compounds by chelating with metal atoms. The ideal coordination sites in the dioxatetraaza ligand were predicted using the Mulliken method and molecular electrostatic potential analysis. All of them indicate how brittle the bond is and imply that it will be the first to fail (Supplementary Tables S1 and S2). This theory fit in with the previous discussion and was consistent with how mass fragmentation and heat deterioration work.



Figure2.Optimized ligand structure with numbering system.

#### **Characterization of metal complexes**

Figure 2 illustrated how the dioxatetraaza ligand structure was numbered. We can show that the

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melting temperatures of nitrogen and oxygen atoms are greater than those of other atoms of a similar size, indicating that these atoms would be able to form coordination compounds by chelating with metal atoms. The ideal coordination sites for the dioxatetraaza ligand were determined using the Mulliken method and a study of the molecular electrostatic potential. The relationship is clearly tenuous, and all of them (Supplementary Tables S1 and S2) imply that it will be the first to fall apart. The implications of mass fragmentation and heat deterioration were supported by this theory, which also added to the earlier discussion. [**39**].

## **Molecular Docking**

With auto Dock, you can see the results of testing up close and discuss and demonstrate the biological benefits of ligands. Examples of hosts that can be utilized to attach ligands (guests) include Aspergillus *flavus*(Fungus)(7BOP), Candida albicans (Fungus)(5k04), Escherichia coli(G-)(3t88), and *Staphylococcus aureus*(G+)(3ty7). Figures 5-8 demonstrate that HB plots are capable of producing findings that are comparable and reveal a high level of interaction with all receptors. All proteins possessed inter hydrogen bonds that could be seen, according to calculations. The interactions between docking molecules can be depicted in three dimensions. Utilizing three-dimensional graphics, the mechanism of interaction within docking molecules may be demonstrated (Figures 3-6).

The interaction between the ligand and the amino acids of the protein in the following bacteria was shown to be largely mediated by hydrogen bonds. For Candida albicans fungus(5k04), An amino acid in the protein reacted with the ligand to H-bond reaction:5k04cause the h/5k04/B/ASN<sup>117/2HD2-</sup> with H bond length 2.3 A°, 5k04-h/5k04/B/TYR`124/OH-with H bond length 3.3 A°, 5k04-h/5k04/B/ASN`117/ND2-with Η bond length 2.5 Aº. 5k04h/5k04/B/GLU<sup>63</sup>/OE1-with H bond length 3.4 A° and 5k04-h/5k04/B/PHE`112/O-with hydrogen bond length 2.7 A°, with binding energy = -10.9kcal mol<sup>-1</sup>(Figures 3–6).

For *Aspergillus flavus* fungus(7BOP): amino acid of protein reacted with ligand by H-bond:1- 7bop-h//A/GLU`205/OE2–with H bond length 2 A°, 7bop-h//A/GLU`205/OE2–with hydrogen bond length 3.5 A°, 7bop-h//A/TRP`291/HE1–with hydrogen bond length 2.7 A° and 7bop-h//A/SER`389/O–with hydrogen bond length 3.1 A°,

with binding energy = -8.6 kcal mol<sup>-1</sup>(Figures 3–6).

For *Staphylococcus aureus*(G+)(3ty7), the amino acid of the protein combines with the ligand via a hydrogen bond:3ty7-correct-A-h//A/GLU`49/OE1– with hydrogen bond length 3.3 A°, 3ty7-correct-A-h//A/GLU`49/OE1–with H bond length 2.6 A°, 3ty7-correct-A-h//A/GLU`49/OE2–with hydrogen bond length 3.5 A°, 3ty7-correct-A-h//A/GLU`49/OE2– with H bond length 1.9A°, with binding energy =-5.8 kcal mol<sup>-1</sup>.

For Echerichia coli(G-)(3t88): ligand reacted with amino acids of protein by H-bond as follow:3t88-correct-A-h/A1/A/SER`84/HG–with hydrogen bond length 2.2 A°, 3t88-correct-A-h/A1/A/VAL`44/O– with H bond length 3.1 A° and3t88-correct-A-h/A1/A/GLY`78/O–with hydrogen bond length 2.7 A°, with binding energy = -7.6 kcal mol<sup>-1</sup> (Figures 3–6).



Figure 3.Three-dimensional plot of interaction of dioxatetraaza ligand with Candidaalbicans*fungus*(5k04) receptor.



Figure 4. Three-dimensional plot of interaction of dioxatetraaza ligand with *Aspergillus flavus* fungus-7BOPreceptor



Figure 5. Three-dimensional plot of interaction of dioxatetraaza ligand with *Escherichia coli*(G–)-3t88receptor.



Figure 6. Three-dimensional plot of interaction of dioxatetraaza ligand with *Staphylococcus aureus*(G+)-3ty7receptor.

3.9. Antimicrobial activity

We used the serial dilution approach to assess all the compounds' in vitro antimicrobial (antibacterial and antifungal) activities against two fungal pathogens and four bacterial pathogens because the entire world is currently dealing with various illnesses brought on by microorganisms. The results were compared to the benchmarks amphotericin B and tetracycline for their relative antibacterial and antifungal properties.

The information gathered led to the following conclusions. The antibacterial activity of produced dioxatetraaza ligand and metal complexes is significantly influenced by the presence of the azomethine group (-C=N-). The research findings showed that metal complexes are more powerful than their corresponding dioxatetraaza ligand. The higher potency of metal complexes was explained by chelation theory and overtone's concepts, which showed that lipophilicity is an important factor in the increased activity of metal complexes. Chelation

enhances the capacity of the complex to pass through lipid membranes and decreases the polarizability and positive charge on the metal, which prevents the active site of microorganisms.

- Among the bacterial strains all complexes have more potency against *Staphylococcus aureus* and *E. coli* with ordered Cu(II), Ni(II) and Zn(II) are most highest complexes activity[**40,41**].

- All the synthetic compounds showed strong antifungal activity against C. albicans and Aspergillus flavus, with the most effective activity being seen in the complex ordered Cu(II) and Zn(II), and Co(II) and Ni(II) complexes are moderated one while Cr(III) complexes and Fe(III) are the least one.



Figure 5.Biological activities of dioxatetraaza ligand and its metal complexes.

## Conclusion

Dioxatetraaza ligand and its Co(II), Cu(II), Ni(II), Zn(II), Cr(III), and Fe(III) complexes were studied using physicochemical methods. Thermal measurements (TGA and DTG) revealed that the complexes were thermally stable up to 800 0C. DFT calculations were used to confirm that the octahedral geometry of the Co(II), Cu(II), Ni(II), Zn(II), Cr(III), and Fe(III) complexes was disclosed by combining magnetic susceptibility data with electronic spectra. All of the synthesized compounds demonstrated a weak to strong antibacterial impact when used against pathogenic bacteria species. The antimicrobial study also found that complexes' growth inhibitory effects were stronger than those of their ligands. The higher antibacterial activity of the complexes is assumed to be caused by the hetero atoms present in them. To ascertain the antibacterial effectiveness, a docking research was conducted.

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