



## A Review on Traditional uses, Phytochemistry and Pharmacological Potential of Family Malpigiaceae

Haidy A. Abbas<sup>1</sup>, Soad H. Tadros<sup>2</sup>, Sayed A. El-Toumy<sup>3</sup>, Ahmed M. Salama<sup>1</sup>,

Rania A. El Gedaily<sup>2\*</sup>



CrossMark

<sup>1</sup>Department of Pharmacognosy<sup>c</sup>, Faculty of Pharmacy, Ahram Canadian University, 6<sup>th</sup> of October City 12451, Giza, Egypt

<sup>2</sup>Department of Pharmacognosy, Faculty of Pharmacy, Cairo University, Kasr El-Aini St., Cairo 11562, Egypt

<sup>3</sup>Chemistry of Tannins Department, National Research Centre, ElBhouth St. (Former ElTahrir St.) Dokki 12622, Giza, Egypt

### Abstract

Herbal medicine has become more popular in recent years. As a result, an effort is being made to document valuable phyto-constituents and pharmacological knowledge as part of the revitalization of herbal remedies. There are still several plants of certain families that haven't been researched. This is the situation with Malpigiaceae; a flowering plant family rich in secondary metabolites including alkaloids, flavonoids, carbohydrate like substance as vitamin C, proanthocyanidins and phenolic compounds, with promising therapeutic effects as anti-inflammatory, anti-ulcer, anti-cancer, anti-diabetic, antioxidant, anti-depressant, anti-HIV, and anti-microbial activities. The goal of this review is to offer an overview of the chemistry of plants belonging to Malpigiaceae, with a focus on their potential biological impact in recent years.

**Keywords:** Malpigiaceae; *Byrsonima*; *Malpighia*; Phytochemical constituents; Biological study.

### 1. Introduction

Nature, particularly plants, is an important source of compounds in healthcare. Only about 15% of the world's plant species have been studied for their pharmacological properties [1]. Even though a wide range of medicinal plants have developed novel and diversified chemical identities that might be utilized as drugs, some botanical families remain unstudied. This is the case of the Malpigiaceae family, a large plant family with about 65 genera and 1,250 species which can be located in tropical and subtropical regions in both hemispheres [2]. It is a family of flowering plants, including trees or shrubs usually lianas [3].

The infra-family classification is based on winged or unwinged fruit, even though the family is obviously monophyletic [4, 5]. Malpigiaceae is a family that belongs to Malpighiales order, Rosidae subclass, Magnoliopsida class, Magnoliophyta division, Spermatophyta superdivision, and Tracheobionta subkingdom [6, 7]. This family is

difficult to study due to the large number of species, nomenclatural issues, and difficulty in taxonomic identification. For example, glandular calyces are prevalent in the neotropical Malpigiaceae, but glandular calyces can be found in species belonging to the genera *Banisteriopsis*, *Byrsonima*, *Galphimia*, and *Pterandra* [8], making it difficult to discriminate between these genera using this morphological feature. Complications arise regularly because of morphological variety and species synonymies. [2, 9, 10]. Most botanists believe the family is related to the Geraniales; Hutchinson included it in his Malpighiales (along with the Erythroxylaceae) whereas Hallier placed it in the Polygalales [3].

Several secondary metabolites with medicinal effects have been discovered in the Malpigiaceae family. Although a wide range of medicinal plants have offered new and different chemical identities that could be effective as medications, only a few species in this family have been researched in terms of chemistry and biology [11]. The phytochemical

\*Corresponding author e-mail: [rania.elgedaily@pharma.cu.edu.eg](mailto:rania.elgedaily@pharma.cu.edu.eg); (Rania A. El Gedaily)

Receive Date: 01 February 2022; Revise Date: 08 March 2022; Accept Date: 10 March 2022.

DOI: [10.21608/EJCHEM.2022.119510.5372](https://doi.org/10.21608/EJCHEM.2022.119510.5372).

©2019 National Information and Documentation Center (NIDOC).

studies showed most plants belonging to that family contain  $\beta$ -carbolines alkaloids, vitamins, carotenoids, nor-secofriedelanes and nor-friedelane terpenoids, hydroxycinnamic acids, flavonoids, proantho-cyanidines, and phenolic compounds.

Malpighiaceae family contains several medicinally significant genera, including the following: *Acridocarpus*, *Aspidopterys*, *Banisteriopsis*, *Bunchosia*, *Byrsomina*, *Callaeum*, *Caucanthus*, *Camarea*, *Diplopterys*, *Echinopterys*, *Flabellaria*,

*Galphimia*, *Hiptage*, *Heteropterys*, *Hiraea*, *Malpighia*, *Stigmaphyllon*, *Tetrapteryis*, *Tristellateia*, and *Niedenzuella*. Pharmacological investigations have revealed that most genera in this family have significant biological activities such as antioxidant, anti-inflammatory, anti-diabetic, anti-microbial, anti-depressant, and cytotoxic properties

**Table 1. Constituents and biological activities of selected genera of the Malpighiaceae family**

Plant species	Part used	Phytoconstituents	Chemical Class	Biological activities	Ref.	
<i>Acridocarpus orientalis</i> A. Juss	Aerial parts	Morin and morin-3- <i>O</i> - $\beta$ -D-glucopyranoside.	Flavonoids	Antifungal, phytotoxic anticancer, anti-lipid peroxidation	[24]	
		$\beta$ -sitosterol, $\beta$ -sitosterol-3- <i>O</i> - $\beta$ -D-glucopyranoside, betulinic acid and botulin	Steroids and triterpenoids	NA	[49]	
		2,5-dimethoxy-1,4-benzoquinone, 2,6-dimethoxy-1,4-benzoquinone	Benzoquinone			
		Quercetin, choerospondin, morin, morin-3- <i>O</i> - $\alpha$ -L-rhamnopyranoside, and morin-3- <i>O</i> - $\beta$ -D-glucopyranoside.	Flavonoids			
			1-docosanol	Saturated fatty alcohol		
	Leaves and stem	NA	Flavonoids and phenolic compounds	Antioxidant, lipid peroxidation, anticancer, $\alpha$ -glucosidase, and urease inhibitory.	[27]	
	Leaves		Morin and morin-3- <i>O</i> - $\beta$ -D-glucopyranoside	Flavonoids	Anticancer (inhibit 4T1 cells and promotes mesenchymal stem cells (MSCs) proliferation).	[48]
			$\beta$ -sitosterol, $\beta$ -sitosterol-3- <i>O</i> - $\beta$ -D-glucopyranoside and betulinic acid.	Steroids and triterpenoids		
			Botulin	Miscellaneous		
	Aerial parts	NA	Flavonoids, tannins, carbohydrates	Hepatoprotective	[26]	
Leaves and stem	NA	NA	NA	Antidiabetic	[103]	
Leaves and stem		Methyl 8-pimaren-18-oate, octacosane, heptacosane, hexacosane, methyl dehydro-abietate, tetracosane, heptacosane, docosane, $\alpha$ -pinene, and heneicosane.	Volatile constituents	Urease, $\alpha$ -glucosidase, and carbonic anhydrase II (CA-II) enzyme inhibitory.	[41]	
<i>Acridocarpus Smeathmannii</i> (DC.) Guill. & Perr.	Roots	Octadecanoic acid ethyl ester, docosenoic acid ethyl ester, Octadecenoic acid ethyl ester, Octadecenoic acid	Fatty acids	A male reproductive enhancer	[21]	

<i>Aspidopterys obcordata</i> Hemsl	Vines	Obcordatas A-I	Polyoxypregnane glycosides	Antitumor (against the human cancer cell lines AGS, SW480, HuH-7 and MCF-7).	[17]	
		NA	NA	Kidney Stones (Inhibit NOX4 expression).	[18]	
		Aspidoptoids A–D, spruceanol and sonderianol.	Diterpenoids.	Cytotoxic and inhibit nitric oxide (NO) production.	[104]	
<i>Aspidopterys indica</i> Willd.	Aerial parts	NA	Tannins, phytosterols, flavonoids	Antioxidant	[57]	
<i>Banisteriopsis argyrophylla</i>	Leaves	Catechin, hyperoside, guaijaverin, and reinutrin	isoquercitrin, quercitrin,	Flavonoids	Antifungal and cytotoxic	[105]
		Catechin, quercetin-hexoside, quercetin-pentoside and quercetin-3-O- $\alpha$ -L-rhamnopyranoside, kaempferol-3-O- $\alpha$ -L-rhamnopyrano-side, quercetin-3-O-(2"-galloyl)- $\alpha$ -L-rhamnopyranoside, Procyanidin dimer, and procyanidin dimer monogallate.		Flavonoids	Antioxidant, and $\alpha$ -amylase, $\alpha$ -glucosidase, pancreatic lipase, and glycation inhibitors.	[58]
		Macarangioside A, and macarangioside B		Megastigmane glucosides		
	Stem	Harmine, harmaline and caffeine		Alkaloids	CNS stimulants	[106]
	Aerial parts	1-Carbamoyl-7-methoxy $\beta$ -carboline, 1-acetyl-7-methoxy $\beta$ -carboline and 1-methoxy-1,2,3,4-tetrahydro-1-oxo- $\beta$ -carboline.		Alkaloids	NA	[107]
		Banistenoside A, banistenoside B and their acetate		Alkaloidal glycosides	Treatment of Parkinsonism, and other neurodegenerative disorders.	
<i>Banisteriopsis Caapi</i> (Syn. <i>Banisteriopsis inebrians</i> )	Stem bark	Tetrahydroharmine, harmol, tetrahydro-norharmine harmaline, and harmine.		$\beta$ -carbolines alkaloids		
		Epicatechin, and procyanidin B2		Proanthocyanidines		
		$\beta$ -d-fructofuranosyl-(2-->5)-fructopyranose, saccharose, and $\beta$ -D-glucose		Monosaccharide, disaccharide		
	Aerial parts	Banistenoside A, banistenoside B, harmine, harmaline, and tetra-hydroharmine, and harmol. Epicatechin, and procyanidine B2		$\beta$ -carboline alkaloids	Treatment of neurodegenerative Disorders Relevant to Parkinson's Disease (MAO inhibition and antioxidant effects).	[29]
		Harmine, harmaline, and tetra-hydroharmine		$\beta$ -carboline alkaloids	Antidepressant	[28]
	Liana	5-hydroxytryptamine (serotonine), and N, N-dimethyltryptamine (DMT)		Miscellaneous		
		Harmine, harmaline, and tetra-hydroharmine		$\beta$ -carboline alkaloids	Monoamine oxidase (MAO) inhibitors.	[108]
		N, N-dimethyltryptamine (DMT)		Miscellaneous		

<i>Banisteriopsis campestris</i>	Flowers	Hexadecanoic acid (palmitic acid), nerolidol, myristic acid, linoleic acid, triacontane, heptacosane and linalool.	Volatile constituents and Fatty acids	Antibacterial, and antifungal	[42]
	leaves, stems, and roots	Palmitic acid, palmitoleic acid, phytol, triacontane, linoleic acid, and oleic acid.	Volatile constituents and Fatty acids	Antibacterial, antifungal, antioxidant, antiprotozoal, cytotoxicity on Vero cells, and glycation inhibitors.	[43]
<i>Banisteriopsis cornifolia</i>	Bark, leaf, and stem	NA	NA	Antidote, treat side effects caused by snakebite	[109]

<i>Bunchosia armeniaca</i>	Leaves	Rutin, afzelin, isoquercitrin, kaempferol and quercetin	Flavonoids	Antimicrobial, antioxidant, antiinflammatory	[59]
	Unripe, ripe fruit	Pyran-4-one, 2,3-dihydro-3,5-dihydroxy-6-methyl, 1H-Pyrrole-2,5-dione, 1-Nona-decene, 3-Eicosene, 2-Furanmethanol, 9,12,15-Octadecanoic acid, methyl ester and <i>n</i> -Hexadecanoic acid	Volatile constituents and Fatty acids	Antioxidant	[44]
	Fleshy Fruit	Vitamin C	Vitamins		[11, 110]
<i>Bunchosia glandulifera</i>	Leaves	NA	NA	Anti- against <i>Klebsiella pneumoniae</i> .	[74]
	Fruit pulp & seed	Lauric, linolenic, docosadienoic, myristic, cerotic, myristoleic, palmitic, palmitoleic, stearic, oleic, linoleic, arachidonic, and behenic acids.	Fatty acids	NA	[45]
	Ripe fruits	$\beta$ -Carotene, and lycopene Vitamin C	Carotenoids Vitamins		[111]
	Tree	NA	Vitamins, Carotenoids	Antioxidant	[60]
	Fruit pulp	Rutin, vitexin, and quercitrin $\beta$ -Carotene, and lycopene Vitamin C Caffeine	Flavonoids Carotenoids Vitamins Alkaloids		[112-114]
<i>Byrsomina Sericea</i> DC	Stem bark	NA	NA	Fevers, diarrheas, syphilis, and kidney diseases	[115]
	Leaves	NA	Flavonoids, proanthocyanidins and tannins	Antioxidant, gastro-protective properties	[116]
<i>Byrsomina bucidaefolia</i>	Leaves	Methyl gallate and Methyl <i>m</i> -trigallate	Phenolic acids	Antioxidant	[61]
<i>Byrsomina coccolobifolia</i>	Leaves & Stems	Isoquercitrin, catechin, epicatechin, quercitrin, quercetin and kaempferol	Flavonoids	Antileshimina	[98]
<i>Byrsomina crassa</i> Nied.	Leaves & Aerial parts	NA	NA	Antimicrobial (against <i>Mycobacterium fortuitum</i> )	[117]
	Leaves	Quercetin, and quercetin-3- <i>O</i> -(2''-galloyl)- $\alpha$ -L-arabinopyranoside.	Flavonoids	Antimicrobial	[75]
		Methyl gallate, and epigallocatechin gallate.	Phenolic acids		
		Quercetin-3- <i>O</i> - $\beta$ -d-galactopyranoside, quercetin-3- <i>O</i> - $\alpha$ -l-arabinopyranoside, amentoflavone, catechin and epicatechin.	Flavonoids	Antiulcerogenic	[118]
		Quercetin-3- <i>O</i> - $\beta$ -d-galactopyranoside, quercetin-3- <i>O</i> - $\alpha$ -l-arabino-pyranoside, amentoflavone and catechin. Methyl gallate	Flavonoids Phenolic acids	Mutagenic (using <i>Salmonella</i> mutagenicity and micronucleus tests).	[119]

<i>Byrsomina crassa</i> Nied.	leaves & bark	$\alpha$ -amyrin, $\beta$ -amyrin and their acetates, lupeol, oleanolic acid, ursolic acid, friedelin, and $\alpha$ -amyrinone.	Triterpenes	Antitubercular	[120]
	Aerial parts	NA	Polyphenolic compounds, flavonoids, tannins, and terpenoids.	Antimicrobial (Anti <i>H. pylori</i> ), immunostimulatory	[121]
<i>Byrsomina crassifolia</i> (L) Kunth (Syn. <i>Byrsonima fagifolia</i> Nied.)	Leaves	Betulin aldehyde, betulin, betulinic acid, lupeol, oleanolic acid and ursen-aldehyde $\beta$ -sitosterol and its glucoside Catechin, epicatechin, guajaverin, hyperin, quercetin and its 3-O-[6''-galloyl] galactoside Methyl gallate (an aromatic ester). Alanine, aspartic acid, proline, valine, pipercolic acid and 5-hydroxy-pipercolic acid	Triterpenes Sterols Flavonoids Phenolic acid Protein and non-protein amino acids.	Spasmogenic activity	[50]
	leaves & bark	NA	NA	Antidote and treat side effects caused by snakebite	[109]
	Leaves	Gallic acid, and methyl gallate Quercetin and its glycosides	Phenolic acids Flavonoids	Gastro protective, healing, and antidiarrheal	[94]
	Fruits & Seed	NA	NA	Antihyperglycemic, antihyperlipidemic and antiglycation	[70]
	Aerial parts	Quercetin 3-O-xyloside, quercetin, rutin and hesperidin	Flavonoids	Antidepressant	[88]
	Leaves	Bassic acid, lupeol, $\alpha$ - amyryn, $\beta$ -amyryn and their acetates.	Triterpene	Antitubercular	[51]
		NA	NA	Diabetic wound healing in rats (by tissue regeneration).	[122]
	Seeds	labda-17-acetoxi-13E-en-15-palmitate, and abda-8(17),13E-dien-19-carboxy-15-yl palmitate)	Diterpene labdane	Antimicrobial	[123]
		Byrsoninas A and B	Dimeric guaianolides sesquiterpene lactone	Antioxidant, hypoglycemic, and hypolipidemic.	[71]
	Bark	NA	Phenolic compounds, tannins, flavonoids, anthra-quinones, triterpenes, cardiotonic glycosides and reducing sugars.	Antifungal and antioxidant	[124]
Yellow & red nance fruits	Lutein and its isomers, zeaxanthin, $\beta$ -carotene and its isomers and lutein dimyristate	Carotenoids and xanthophyll esters	High lutein renders nance fruit as a nutritionally (micro-nutrient).	[125]	

<i>Byrsonima crassifolia</i> (L) Kunth (Syn. <i>Byrsonima fagifolia</i> Nied.)	Leaves	Catechin, epicatechin, and quercetin Ferulic acid	Flavonoids Phenolic acids	Antioxidant	[126]
	Fruits	NA	Polyphenols and carotenoids	Antioxidant effects and application to aging.	[62]
		NA	Phenolic compound, flavonoids, and vitamin C	Antioxidant	[63]
	Leaves	Gallic acid, ferric acid, ferrulic acid Myricetin and quercetin, catechin, and epicatechin	Phenolic acids Flavonoids	Antimicrobial	[127]
<i>Byrsonima duckeana</i> W. R. Anderson	Pulp	NA	Phenolic compounds, flavonoids, and fatty acids.	Antioxidant, Cytotoxic and Cytoprotective	[83]
		NA	NA	Hemolytic and cytotoxicity (against cell lines a U937 human monocyte, and HT29 tumor colon cell lines).	[84]
	Leaves	Ethyl gallate, quinic acid, and gallic acid, Catechin, epicatechin, quercetrin, and quercetin	Phenolic acids Flavonoids	Analgesic, antiinflammatory, and antioxidant	[64]
		Leaves	Catechin, epicatechin, gallic acid, methyl gallate, amentoflavone, quercetin and its glycosides.	Phenolic compound and flavonoids	Mutagenic (for the strains TA98 and TA100 in the Ames assay).
<i>Byrsonima intermedia</i> A. Juss.	Stem bark	NA	Flavonoids, saponins, tannins, triterpenes, and steroids.	Antiinflammatory and antinociceptive	[129]
	Leaves	Catechin	Flavonoids	Chronic and acute antiinflammatory	[130]
		Gallic acid, 3,4-di-O-galloylquinic acid, methyl gallate, catechin, epicatechin, 1,3,5-tri-O-galloylquinic acid, amento-flavone, quercetin, and its glycosides.	Phenolic acids, oligomeric proanthocyanidins, and flavonoids.	Gastric and duodenal antiulcer, antimicrobial and antidiarrheal	[95]
		Gallic acid, 3,4-di-O-galloylquinic acid, methyl gallate, catechin, epicatechin, 1,3,5-tri-O galloyl-quinic acid, 1,3,4,5-tetra-O-galloylquinic acid, quercetin and its glycosides.	Phenolic acids, oligomeric proanthocyanidins, and flavonoids.	Gastroprotective (against peptic ulcers, improve healing through antioxidant and antiinflammatory).	[96]
		Cinnamic acids, galloyl quinic and shikimic acid. Quercetin, epicatechin and their glycosides	Phenolic acids Proanthocyanidins, and glycosylated flavonoids	NA	[131]
		Lupane and oleanane, betulinic acid, oleanolic acid, $\beta$ -amyrin and 3-oxo-olean-12-en-28-al.	Triterpenes		

		NA	NA	Treatment of external ulcers and inflammations.	[115]
<i>Byrsonima gardneriana</i> (A. Juss.)	Leaves	Pyroglutamic, Hexadecanoic, heptanoic, and octanoic acids. Eucalyptol Retinal (Vitamin A)	Fatty acids Terpenoids Vitamins	Antifungal against <i>Candida</i> spp., antioxidant activity, and cytotoxicity.	[46]
	Stem bark	NA	NA	Treatment of external ulcers and inflammations	[115]
<i>Byrsonima verbascifolia</i> (L.) DC.	Leaves	Quinic acid, gallic acid, proto-catechuic acid, and their glycosides Epicatechin, catechin, rutin, apigenin, quercetin, kaempferol and their glycosides Procyanidins, and Prodelphinidin	Phenolic acids Flavonoids Proanthocyanidin	Antiinflammatory (inhibition of tumor necrosis factor alpha, prostaglandin E2 production and polymorphonuclear leucocyte migration).	[132]
		Quercetin, epicatechin, catechin, and amentoflavone Quinic acid, and gallic acid	Flavonoids Phenolic acids	Antiinflammatory	[133]
	Fruit	butanoic acid, ethyl ester; hexanal; 2-heptanone; methyl octanoate; butyl hexanoate; ethyl octanoate; decanoic acid, methyl ester; and hexanoic acid, ethyl ester	Volatile constituents	Antioxidant	[134]
<i>Byrsomina microphylla</i> A. Juss.	Leaves	24-hydroxy-urs-12-enyl 3b-eicosanate, estearate and palmitate, 24-hydroxy-olean-12-enyl 3b-eicosanate, oleanolic and 3b,24-dihydroxy-urs-12-en-28-oic acids. Quercetin Methyl galic ester	Triterpenes esterified with fatty acid Flavonoids Phenolic acid	NA	[135]
		NA	NA	Antiinflammatory, antihyperalgesic, antiplatelet and antiulcer	[34]
<i>Byrsomina japurensis</i> A. Juss.	Stem bark	NA	Phenolic compounds (anthocyanins/anthocyanidins, aurones, chalcones, flavanones and condensed tannins and steroid compounds (saponins, pentacyclic triterpenes, cardio-active steroids).	Antioxidant (by DPPH and ABTS radical scavenger's tests).	[136]
<i>Callaeum antifebrile</i> (Griseb.)	Stem & leaves	Harmine	Alkaloids	Antifever	[137]
<i>Caucanthus auriculatus</i> (Radlk.) Nied.	Leaves	NA	NA	Nutritive value for animal livestock	[138]
<i>Camarea. ericoides</i> <i>C. humifusa</i> <i>C. affinis</i> <i>C. hirsute</i> <i>C. elongates</i> <i>C. axillaris</i> <i>C. sericea</i>	Leaves	Apigenin, apigenin 7-O-glucoside, luteolin 7-O-galactoside, chrysoeriol, kaempferol, kaempferol 3-O-glucoside, kaempferol 3-O-galactoside, kaempferol 3-O-rutinoside, Quercetin, Quercetin 3-O-glucoside, Quercetin 3-O-galactoside, Quercetin 3-O-rutinoside (Rutin).	Flavonoids	NA	[139]

		NA	Flavonoids. Alkaloids, terpenes and prenylated xanthenes	Antioxidant	[65]
<i>Diplopterys pubipetala</i> (A.) Juss.	Leaves & stems	NA	Phenolic compounds, among them, mainly flavonoids	Antitumor (Melanoma Cell Line)	[54]
		N- <i>cis</i> -Feruloyl-tyramine, and Simulansamide	Alkaloids	NA	[140]
		Cucumerin A, Syringetin 3- glucuronide, and Macarangaflavanone A.	Flavonoids		
		3- $\beta$ - <i>O</i> -( <i>cis</i> -p-coumaroyl) corosolic acid, 25-anidro- alisol F, Phytuberina	Terpenoid		
		Ginsenoside S-cucujolide	Saponins Lactone		
<i>Echinopterys eglandulosa</i> (A.) Juss.	Flowers	NA	NA	Antibacterial	[141]
<i>Flabellaria paniculata</i> Cav.	Leaves	NA	Saponins, cardenolides, alkaloid and tannins.	Antibacterial (aganist <i>Staphylococcus</i> . <i>aureus</i> , <i>Pseudomonas</i> <i>aeruginosa</i> , <i>Eustarcia</i> <i>coli</i> and <i>Klappellia</i> <i>pneumoniae</i> ).	[76]
		NA	Tannins and amino- glycosides	Antiinfective and wound healing	[101]
		NA	NA	<i>In-vitro</i> antibacterial and <i>in-vivo</i> wound healing activities	[102]
	Whole plant	NA	Saponins, alkaloids, anthraquinones, flavonoids and tannins	Antimicrobial (against different <i>Candida</i> species)	[142]
	Leaves & roots	NA	Phenolics, flavonoids and proanthocyanidins	Antioxidant	[66]
	Leaves	NA	Terpenoids, tannins, saponin and flavonoids.	Gastric ulcers	[143]
		NA	Tannins, saponins, flavonoids, and of a steroidal nucleus (cardiac glycoside)	Sub-Chronic Oral Toxicity	[144]
		Friedelin and friedelinol Sitosterol and sitosterol- $\beta$ -d- glucoside) Kaempferol-3- <i>O</i> - $\alpha$ -l-rhamno- pyranosyl-(1 $\rightarrow$ 6)- $\beta$ -d- glucopyranoside	Triterpenoids Steroids Flavonoid glycoside	Gastroprotective	[52]

<i>Galphimia glauca</i> Cav.	Whole plant	Gallic acid, methyl gallate, and tetragalloylquinic acid  Quercetin	Phenolic acids  Flavonoids	Antiasthmatic	[145]
	Aerial parts	Galphimine B	Nor-secotriterpenoid	Sedative	[146]
	Whole plant	Galphin A, galphin B, galphin C, and galphimidin.  Quercetin  Stigmasterol and sitosterol 3-O- $\beta$ -D-glucoside.	Nor-secofriedelanes and nor-friedelane terpenoids  Flavonoids  Sterols	Antiprotozoal Activity	[99]
	Aerial parts	Galphimine B.	Nor-secotriterpene.	Anxiolytic and antidepressant	[89]
	Aerial parts	Galphimine-B, galphimine-A, galphimine-E, and galphimine-J  $\alpha$ -amyirin, $\beta$ -amyirin  $\beta$ -sitosterol, and $\beta$ -sitosteryl-3-O- $\beta$ -D-glucopyranoside  methyl-gallate, 4-methoxy methyl gallate, and gallic acid.  Kaempferol, quercetin, kaempferol 3-O- $\beta$ -D-glucopyranoside, quercetin 3-O- $\beta$ -D-glucopyranoside, kaempferol 3-O- $\beta$ -D-(2''-galloyl)-glucopyranoside, kaempferol 3-O- $\beta$ -D-(2''-galloyl) galactopyranoside, and quercetin 3-O- $\beta$ -D-(2''-galloyl)-galactopyranoside.	Nor-secotriterpene.  Triterpenoid  Sterols  Phenolic acids  Flavonoids	Antiinflammatory	[86]
	Leaves	NA	Amino acids, carbohydrates, proteins, flavonoids, tannins, and phenolic compounds.	CNS Depressant and muscle relaxant	[147]
	Bark & leaves	NA	Phenolic compounds, flavonoids, alkaloids, carbohydrates, steroids, protein & amino acids, anthraquinone, gums & mucilage, glycosides, tannins and saponins.	Antioxidant, antibacterial and anticancer Activities	[67]

<i>Hiptage benghalensis</i> (L.) kurz	Leaves	NA	NA	Anthelmintic	[148]
		NA	Carbohydrate, proteins, amino acids, saponins, tannins, glycosides, phenolic and flavonoids.	Antiinflammatory, and anthelmintic	[149]
		NA	Phenolic compounds	Antioxidant	[150]
	Leaves & stem bark	NA	Alkaloids, anthraquinones, coumarin, flavonoids, phenols, steroids, tannins, terpenoids & xanthoprotein	Antibacterial (against <i>Staphylococcus aureus</i> , <i>Klebsiella pneumoniae</i> , <i>Bacillus subtilis</i> , <i>Escherichia coli</i> , <i>Pseudomonas aeruginosa</i> and <i>Salmonella typhi</i> ).	[151]
	Leaves	NA	Steroids, terpenoids, carbohydrates, phenolics and glycosides.	Hepato-protective (against carbon tetrachloride-induced liver damage in rats).	[92]
	Root bark	NA	NA	Larvicidal, adulticidal, and repellent (against the larvae and adults of <i>Anopheles barbirostris</i> , <i>Culex quinquefasciatus</i> , and <i>Aedes albopictus</i> )	[152]
	Leaves, stem & flower	NA	Flavonoids (free + bound)	NA	[153]
	Stem	NA	Steroids, carbohydrate, flavonoid, alkaloid, tannins, phenol, mangiferin and terpenoids.	Antidiabetic	[154]
	Leaves	NA	Steroid, carbohydrate, flavonoid, alkaloid, tannin, phenol, mangiferin and terpenoids.	Analgesic and antiinflammatory	[87]
		NA	Steroids, terpenoids, carbohydrate, phenolics and glycosides.	Antidiabetic	[72]
	Root	NA	NA	Antiobesity	[100]
	Stem bark	Alnus-5(10)-en-3 $\beta$ -yl acetate, oleanan-3-one, 3 $\beta$ -acetoxy-9 $\beta$ -bauer-7-en-6-one, lupeol, 24-propylcholesterol, alnus-5(10)-en-3 $\beta$ -ol, 3 $\beta$ -acetoxy-20-hydroxy-lupane and betulonic acid.	Triterpenes and steroid compound	Antiinflammatory	[155]
		NA	NA	Antimicrobial (against <i>Klebsiella pneumonia</i> , <i>Escherichia coli</i> , <i>Micrococcus luteus</i> and <i>Pseudomonas aeruginosa</i> ).	[77]

<i>Hiptage benghalensis</i> (L.) kurz	Leaves	NA	NA	Anticancer (human cervical carcinoma (HeLa), human breast cancer (MCF-7) and human neuro-blastoma (IMR-32) cells).	[85]
	Leaves	NA	Phenols, tannins, flavonoids, steroids, and triterpenoids	Antidiabetic	[73]
<i>Heteropterys brachiate</i> L. DC.	Aerial parts	Chlorogenic acid and chlorogenic acid methyl ester	Hydroxycinnamic acids	Antidepressant, anxiolytic and anticonvulsive.	[156]
		NA	Mixture of terpene		
		NA	NA	AntiHIV and anti <i>Candida</i> effects	[80]
<i>Heteropterys byrsonimifolia</i> A. Juss.	Leaves	Guajaverin, quercetin 3- <i>O</i> - $\alpha$ -L-rhamnopyranoside, quercetin 3- <i>O</i> -robinobioside and rutin.	Flavonoids	Antifungal (against <i>Aspergillus ochraceus</i> ).	[157]
<i>Heteropterys chrysophylla</i> (Lam.) Kunth	Leaves & twigs	Palmitic acid	Fatty acids	Exhibited hormonal effects on prostate cancer cells	[158]
		Dihydroactinolide Kaempferol-3- <i>O</i> - $\alpha$ -L-rhamnoside, kaempferol-3- <i>O</i> - $\alpha$ -L-rhamnose-(2-1)- $\beta$ -D-xylopyranoside.	Volatile terpene Flavonoids		
<i>Heteropterys cotinifolia</i> A. Juss.	Aerial parts	Chlorogenic acid Rutin	Hydroxycinnamic acids Flavonoids	Antidepressant	[91]
<i>Heteropterys glabra</i> Hook. & Arn. (Syn. <i>Heteropterys angustifolia</i> Griseb)	Root	Hiptagin (1, 2, 4, 6-tetra-3-nitropropanoyl- $\beta$ -D-glucopyranoside)	Aliphatic nitro compound	NA	[159]
	Fruits	NA	NA	Anxiolytic and sedative in DBA/2J mice.	[90]
<i>Heteropterys tomentosa</i> A. Juss. (Syn. <i>Heteropterys aphrodisiaca</i> (O.) Mach.	Roots	NA	Flavonoid, cardiac glycosides with steroidal nucleus, or pentagonal lactonic ring, saponins, hydrolysable and condensed tannins and aliphatic nitro-compounds.	Improves learning and memory deficits in aged rats.	[97]
		2,3,4,6-tetra- <i>O</i> -(3-nitropropanoyl)- <i>O</i> - $\beta$ -D-glucopyranoside	Aliphatic nitro compound	Antimicrobial (against <i>staphylo-coccus aureus</i> , <i>Bacillus subtilis</i> , <i>Candida albicans</i> , <i>C. parapsilosis</i> , <i>C. krusei</i> , and <i>C. tropicalis</i> ).	[78]
		Neoastilbin, astilbin and isoastilbin	Flavonoids	NA	[160]
		2,3,4,6-tetra- <i>O</i> -(3-nitropropanoyl)- <i>O</i> - $\beta$ -D-glucopyranoside	Aliphatic nitro compound	Antiviral (against poliovirus type-1 (PV-1) and bovine herpes virus type-1 (BHV-1) by plaque reduction assay).	[79]
		NA	NA	Stimulant and aphrodisiac (by reduce Cyclosporine A (CsA) induced injuries in the testis.	[161]

<i>Heteropterys tomentosa</i> A. Juss. (Syn. <i>Heteropterys aphrodisiaca</i> (O.) Mach.)	Roots	NA	NA	Increasing the spermatogenic yield (by increase testosterone production and spermatogonia mitosis).	[162]
		NA	NA	Anabolic effects (Produce more organized collagen bundles and more resistant tendons to support higher loads from intense muscle contraction).	[163]
		Astilbin, isoastilbin and neoastilbin 2, 3, 4, 6-tetra- <i>O</i> -(3-nitropropanoyl)- <i>O</i> - $\beta$ -D-glucopyranoside.	Flavonoids Aliphatic nitro compound	Memory retrieval improvement in aging rats and antioxidant	[164]
	Roots, branches & Leaves	NA	NA	A protective action against the side effects of Cyclosporin A on the ventral prostate tissue.	[165]
		Catechin, taxifolin, and rutin. Chlorogenic acid	Flavonoids Hydroxy-cinnamic acids	Not show evidence of adaptogenic effect	[166]
		Caffeoylquinic acids Taxifolin derivatives	Phenolic acids Flavonoids	Decrease the viability of astrocytes.	[167]
		NA	NA	Improves the endurance capacity of skeletal muscles in trained rats	[168]
		NA	NA	Effect on anxiety and male exposure of female mice with advanced age.	[169]
		Taxifolin and taxifolin 3- <i>O</i> -rhamnoside (astilbin).	Flavonoids	Adaptogenic effect.	[170]
		<i>Hiraea reclinate</i> Jacq.	Leaves	Kaempferol 3- <i>O</i> -(6''-galloyl) b-D-galactopyranoside, hyperin 6''-gallate, vitexin 2'' rhamnoside, isovitexin 2'' rhamnoside, orientin 2'' rhamnoside, isoorientin 2'' rhamnoside. 1,3,4,5-tetragalloyl-quinic acid	Flavonoids Phenolic acids
Mature & immature fruit	Acethyl-methyl-carbinol, 2-methyl-propyl-acetate, limonene, E-Z-octenal, ethyl hexanoate, isoprenyl butirate and acetofenone, methyl hexanoate, 3-octen-1-ol and hexyl-butirate, methyl-propyl-ketone, E-Z-hexenyl-acetate and 1-octadecanol.			Volatile components	NA
<i>Malpighia glabra</i> L. (Syn. <i>Malpighia puniceifolia</i> L.)	Fruit	Pelargonidin, malvidin 3,5-di-glycoside and cyanidin 3-glycoside	Anthocyanins and anthocyanidin	NA	[40]
		Quercetin, and kaempferol <i>p</i> -Coumaric, ferulic, chlorogenic, caffeic acids.	Flavonoids Hydroxy-cinnamic acids		

<i>Malpighia glabra</i> L. (Syn. <i>Malpighia punicifolia</i> L.)	Fruit	Lutein, $\alpha$ -carotene, and $\beta$ -carotene $\beta$ -cryptoxanthin  Violaxanthin	Carotenes  Monohydroxy carotenoids Dihydroxy carotenoids	NA	[171]
	Unripe & ripe fruit	Ascorbic acid Rutin and quercetin	Vitamins Flavonoids	Antigenotoxic and antioxidant	[12, 172]
	Fresh Pulp	NA	Ascorbic acid, anthocyanins, carotenoids, phenols, and flavonoids.	Cytotoxic and mutagenic effects of iodine-131 and radioprotection	[173]
	Fruit	NA	Volatile components, vitamins, and phenolic compounds	Antimicrobial (against <i>Pseudo-monas putida</i> , <i>P. fluorescens</i> , <i>P. fragi</i> , and <i>Brochothrix, thermo-sphacta</i> by agar well diffusion and agar dilution tests).	[174]
	Fruit pulp	Tocopherol, and $\beta$ -carotene. Gallic acid, and ellagic acid. Catechin, epicatechin, rutin, quercitrin, quercetin and kaempferol. Caffeic acid	Carotenoids Phenolic acids  Flavonoids  Hydroxycinnamic acids	Antioxidant	[175]
	Fruit pulp	Kaempferol, myricetin, quercetin, and epicatechin Procyanidin B1 Trans-resveratrol	Flavonoids  Procyanidin Phenol	Antioxidant	[176]
	Leaves	Acacetin, hispertin, quercetrin, hesperidin, rutin, naringin and apigenin 6-glucose 8-rhamnose.  NA	Flavonoids  Cardiac glycosides, alkaloids, anthraquinone glycosides, carbohydrates, saponins, sterols and/or triterpenes, tannins, volatile constituents, and vitamins.	Anticancer and antimicrobial activities	[82]
		Caffeic acid, and chlorogenic acid Quercetin, and kaempferol	Hydroxycinnamic acids Flavonoids	Antioxidant	[68]
		Saponarin, vicanin, apigenin-C-hexoside-C-pentosyl, vitexin-O-pentoside, rutin, kaempferol, isorhamnetin-O-rutinoside, isoorientin, fisetin, luteolin, quercitrin, kaempferol-3-O-rutinoside, orientin, quercetin and myricetin.	Flavonoids	Hepatoprotective activity	[93]
		Caffeoyl quinic acid, caffeoyl feruloyl-quinic acid, <i>Trans</i> -Cinnamate and 4-methoxycinnamic acid.	Phenolic acids		

	Fruit	Vitamin C	Vitamins	Antimicrobial, and cytotoxic	[177]
	Ripe & unripe fruit	Tartaric acid, malic acid, and citric acid Ascorbic acid Catechin Caffeic, ferulic, and coumaric acids. Gallic acid, and syringic acid	Organic acids  Vitamins Flavonoids Hydroxycinnamic acids Phenolic acid	Antioxidant	[178]
		Cyanidin-3- $\alpha$ -O-rhamnoside and pelargonidin-3- $\alpha$ -O-rhamnoside.	Anthocyanins	Antihyperglycemic	[179]
	Fruit	Cyanidin 3,5-hexose pentose, Cyanidin 3-glucoside, Cyanidin 3-rutinoside, Pelargonidin 3-glucoside, Peonidin 3-glucoside, Cyanidin 3-rhamnoside and Cyanidin.	Anthocyanins	NA	[180]
		Epigallocatechin gallate, epicatechin and rutin Vitamin C Chlorogenic acid	Flavonoids  Vitamins Hydroxycinnamic acids Procyanidin	Antioxidant (by ABTS, DPPH and ORAC methods).	[181]
<i>Malpighia emarginata</i> DC.		NA	Vitamin C, phenolic compounds, and flavonoids	Antioxidant and antimicrobial	[182]
	Aerial parts	Acerolanins A, B, C  Vitamin C NA	Tetranor-diterpenes Vitamins Anthocyanins, flavonoids, and phenolics.	Cytotoxic	[39]
	Ripe & unripe fruit	Vitamin C Rutin	Vitamins Flavonoids	Effect on brain energy metabolism of mice fed a cafeteria diet.	[183]
	Leaves	Rhinocerotoic acid, and isotriptophenolide Quinic acid, and protocatechuic acid Galocatechin, apigenin-7-O-glucoside, and apigenin-8-O-glucoside Matricin	Diterpene  Phenolic acids  Flavonoids  Sesquiterpene	Antioxidant and anti-fungal	[184]
		Ascorbic acid, and vitamin B3 Rutin Ellagic acid Caffeic acid	Vitamins Flavonoids Phenolic acids Hydroxycinnamic acids	An immune-stimulatory and antiinflammatory.	[185]
	Fruits	Chlorogenic and isochlorogenic acids Cyanidin, delphinidin-3 $\beta$ -D-glucoside, phloretin, peonidin Genistein, luteolin, kaempferol, apigenin 7-glucoside, quercitrin, astragalin, kaempferol 3-O- $\beta$ -glucoside, vitexin, malonylapiin and isovitexin	Hydroxycinnamic acids Anthocyanins  Flavonoids	Antioxidant	[69]

<i>Niedenzuella multiglandulosa</i> (A. Juss.)	Leaves	Trigonelline	Alkaloids	NA	[186]
		Tryptophan	Amino acids		
<i>Stigmaphyllon ovatum</i> (Cav.)	Leaves	4-hydroxy-cinnamic acid, cinnamic acid and cis-p-coumaric acid	Phenolic acids		
		4-O- $\beta$ -D-glucopyranoside			
<i>Stigmaphyllon paralias</i> (A. Juss.)	Aerial parts	Luteoforol	Flavonoids		
		Integristerone A, icariside F2, epiecdysterone, ecdysterone, calonysterone, and odecydysone B.	Miscellaneous		
<i>Stigmaphyllon ovatum</i> (Cav.)	Leaves	NA	Alkaloids, steroids, flavonoids, terpenoids, phenolics, and eugenols	Antimalarial in <i>Plasmodium falciparum</i> infected mice	[187]
		NA	NA	Anticancer against human cervical carcinoma	[188]
<i>Stigmaphyllon paralias</i> (A. Juss.)	Aerial parts	Friedelin, lupenone, 3-oxo- $\alpha$ -amirin, 3-oxo- $\beta$ -amirin, mixture of $\alpha$ -amirinyol palmitate and stearate, lupeol, $\alpha$ -amirine, and 3,4-seco-friedelin-3-oic acid.	Triterpenes	NA	[189]
<i>Tetrapteryx mucronate</i> Cav.	Bark	Luteolin-7-rutinoside	Flavonoids		
		Mucronatin B, 5-hydroxy-N, N-dimethyltryptamine (bufotenine), 5-methoxy-N-methyl-tryptamine, 5-methoxy-N, N di-methyl-tryptamine, trans-N feruloyl-tyramine (Moupinamide), and 2-methyl-6-methoxy-1,2,3,4-tetrahydro- $\beta$ -carboline.	Alkaloids	AntiAlzheimer (acetylcholine-esterase inhibition in a TLC bioautography assay).	[55]
<i>Tetrapteryx mucronate</i> Cav.	Stem bark	Gentisic acid, gentisic acid 5-O- $\beta$ -xyloside, vanillic acid, and methoxy-4,5-(methylenedioxy) cinnamic acid.	Phenolic acids		
		Catechin	Flavonoids		
<i>Tristellateia australasiae</i> (A. Rich)	Leaves & stem	2,6-phenanthrenediol, 7-methyl-2,6-phenanthrenediol, and 6-dihydroxy-9,10 dihydro-phenanthrene.	Phenanthrene derivative		
		Lyonside, cannabisin F, and smilaside L	Carbohydrate		
<i>Tristellateia madagascariensis</i>	Leaves	Nudiposide-9'-dihydroxy-benzoic acid	Aromatic acids		
		Bufotenine, 5-methoxy-N-methyl-tryptamine, 5-methoxy-bufotenine, 2-methyl-6-methoxy-1,2,3,4-tetrahydro-carboline.	Alkaloids	NA	[56]
<i>Tristellateia australasiae</i> (A. Rich)	Leaves & stem	Acyclic hexitol, and dulcitol	Alcohol	NA	[190]
		Isorhamnetin	Flavonoids		
<i>Tristellateia madagascariensis</i>	Leaves	Friedelin, epifriedelinol, $\beta$ -amyryn, lupeol and $\beta$ -sitosterol.	Steroids and triterpenes		
		NA	NA	Antimalarial	[191]

NA: Not available



## 2. Experimental

Different databases are used to collect data conduct research, including SciFinder, PubMed, Science Direct, Scopus, Plos One, Web of Science, and Google Scholar in addition other sources including books, thesis, and official websites. "The Plant List" ([www.theplantlist.org](http://www.theplantlist.org)) was used to verify the accepted species number and names. All chemical structures were drawn using *ChemDraw* Ultra 7.0 software.

## 3. Distribution

Tropical and subtropical regions are the primary habitats for the plants of this family. The New World (the Caribbean and the southernmost United States to Argentina) has about 80% of the genera and 90% of the species, whereas the Old World (the remainder of the world including Africa, Madagascar and Indomalaya to New Caledonia and the Philippines) has the rest. Seven species of five genera are native to the warmer parts of the country: *Byrsonima* in southern Florida, *Malpighia* and *Thryallis* in Texas, and *Aspicarpa* and *Janusia* in western Texas and southern Arizona. *Banisteria* is one of the bigger genera in the family, with roughly 16 species native to Mexico [3]. There is no genus or species that is found in both hemispheres. A Caribbean and Atlantic coast species of a huge American genus, *Stigmaphyllon ovatum* (Cav.) Nied., has been collected numerous times in western Africa. *Heteropterys leona* (Cav.) Exell is a well-known species in western Africa, but it's hard to distinguish it from its closest relatives *H. platyptera* DC. and *H. multiflora* (DC.) Hochr., which grow in the Caribbean and along the Atlantic coast of Central and South America [8].

## 4. Traditional uses

A set of herbal preparations used in the Indian traditional health care system contains a variety of medicinal plants that have been used for thousands of years (Ayurveda). Although the species of Malpighiaceae family may not have a great economic potential, *Malpighia glabra* (Barbados cherry) and *Malpighia emarginata* (Acerola) have a high nutritional value due to their high vitamin C content [12, 13]. The juicy fruits of *Byrsonima* and *Malpighia* are also consumed fresh, prepared, or as canned juices, in jellies, ice cream, and preserves that are popular in Latin American cuisine [14].

*Malpighia* genus has not been widely utilized in traditional medicine, however *Malpighia glabra* is used as a tonic and diuretic [15].

*Acridocarpus*, *Banisteriopsis*, *Byrsonima*, *Galphimia*, *Heteropterys*, *Malpighia*, *Peixotoa*, and *Stigmaphyllon* species are also grown as ornamental plants [13, 16]. *Aspidopterys obcordate* has a long history in Dai traditional medicine, and it contains some important biochemical compounds with anti-phlogistic and diuretic effects that could be used to treat acute, chronic nephritis, cystitis, rheumatism, bone pain, and to expel stone as well as postpartum weight loss in the form of a health tea and children's digestive diseases [17-20]. *A. andamanica* and *A. cordata*, on the other hand, are utilized as postpartum remedy [15].

Several species of *Acridocarpus* are still used as a folk medicine all over the world, and more specific studies to support this is needed. Some species are reported to have many ecological advantages as well. Stomachaches are traditionally treated with *Acridocarpus alternifolius* roots, whereas diarrhoea and dysentery are treated with *A. excelsus* bark, which is an astringent. The root of *A. longifolius* is used to cure venereal diseases and stomach problems as a laxative. Its leaf sap is used to treat eye infections and as a febrifuge; however, its root and leaf sap are utilized to treat cutaneous and subcutaneous parasite infections. *A. plagiopterus* root is used as a febrifuge, vermifuge, reptile repellent, and for sleep sickness, superstitions, and magic, while *A. spectabilis* roots are used as diuretics, cutaneous and subcutaneous parasitic infection, kidney and nasopharyngeal affections, ceremonial, and superstitions [15].

According to folklore medicine, *A. smeathmannii* roots are used for aphrodisiac, anti-anemia, pain killers, and various cutaneous and subcutaneous parasite infections. Several Ayurvedic preparations containing *A. smeathmannii* root were used as aphrodisiacs and used to enhance fertility and barrenness. The effect of *A. Smeathmannii* (DC.) root on reproductive potentials and biochemical pathways in male Wistar rats was investigated in a pharmacological study to determine its folkloric medicine application [21-23]. In Yemen, *A. socotranus* is widely traditionally used to relieve headaches, paralysis, and muscle discomfort. While *A. orientalis* (AO) is a traditional medicinal plant

used for treatment of inflammatory diseases that may have potential in cancer treatment. *A. orientalis* has primarily been recorded from the border areas of UAE and Oman, where it is used for the treatment of muscle pain, headaches, paralysis, tendon, and joint pains as well as to treat the udder inflammation in cattle [24-27].

However, some Malpighiaceae species are used traditionally as CNS modulators. *Banisteriopsis caapi* distinguishes particularly among them because it is the main ingredient in Ayahuasca, an Amazonian psychoactive beverage used in religious ceremonies that contains 5-HT<sub>2A</sub> receptor agonist N, N-dimethyltryptamine (DMT), as well as monoamine oxidase inhibitor alkaloids (harmine, harmaline and tetrahydroharmine) [28-30]. The vine *Banisteriopsis caapi* is a primary source of  $\beta$ -Carbolines alkaloids in the ayahuasca drink, which is made by mixing its stems with the leaves of *Psychotria viridis*. The monoamine oxidase inhibitory (MAOI) action of the vine has been related to the traditional explanation of the vine's participation in this psychedelic beverage [31]. *Diplopterys cabrerana* (also known as *Banisteriopsis rusbyana*) is a hallucinogenic plant that has been used for religious, medical, and social purposes. Ayahuasca is a psychoactive substance used in religious ceremonies. It is an ingredient in the entheogenic tea, a South American hallucinogenic beverage produced by Amazon Indians from the bark of the Malpighiaceae liana *B. caapi* combined with the leaves of other admixture plants, such as *Psychotria viridis*, *Psychotria carthagenensis*, or *Diplopterys cabrerana* [15].

Some species of genus *Bunchosia* have been used traditionally as natural remedies, among of which *Bunchosia armeniaca* is used to treat endocrine, infectious, inflammatory, nutritional, and metabolic disorders, as well as some types of cancer [32]. Seeds of *Bunchosia nitida* (Jacq.) DC. were reported to be used as purgative and antiemetic. While *Bunchosia swartziana* Griseb. is used for externally for scabies while flower juice is often used in ear pain. The leaves and bark are used in treating fungal diseases. While the leaves, bark and flowers are aromatic in nature, and are commonly used as a refrigerant, expectorant, cardiogenic, anti-inflammatory and insecticidal. They are used in burning sensation, wounds, ulcers, leprosy,

epilepsy, convulsion, magic, ritual, and ceremonial to sweep away evil winds or spirits. Tea made from the roots of *Bunchosia glandulosa* (Cav.) DC. has been used for fertility, ritual, and ceremonial purposes, as well as to ward off evil spirits [15].

A decoction of the dry bark of *Byrsonima crassifolia* is used to treat asthma, bronchitis, colds, coughs, fevers, tonsillitis, and skin infections; an infusion is used to treat diarrhoea, gastrointestinal diseases, chronic colitis, chest colds, pulmonary complaints, wounds, skin diseases, stomachache, and as a snakebite antidote. Pounded bark was applied to wounds as a poultice, while pulverized bark was applied to ulcers. Leaves and bark is used to cure diarrhea, the bark ground in water and applied directly to the skin to treat measles. In the Amazonian savannas, *Byrsonima crassifolia* and *B. coccolobifolia*, sometimes known as mirixis, muricis, mantecos, or nances, are the most frequently used fruit species. Their fruits are used to make juices and other beverages, while the rest of the plant is used for a variety of reasons by some indigenous people. A decoction of bark from *Byrsonima spicata* is an antidote for rattlesnake bites, as well as a purgative and febrifuge [15, 33]. *B. japurensis* A. Juss. is used in folk medicine in rural areas of Amazonas State (Brazil), where it is known as "saratudo" and is used to treat gastrointestinal and genitourinary tract disorders, as well as being a potent anti-inflammatory [34].

*Flabellaria paniculata* leaves are used for skin infections and wound dressing. Leaves, seeds, and pods of *Heteropterys leona* have traditionally been used as an antiparasitic, febrifuge, analgesic, and as topical application for headaches and fevers [15]. *Hiptage benghalensis* is a plant that is used in traditional medicine. The leaf is considered one of the important plant organs for the treatment of many diseases. In Ayurveda, the leaves and bark are considered vulnerary, and the leaves are highly recommended for treating skin diseases. The leaf juice possesses insecticidal properties and is applied

cardiac debility, rheumatism, and hyperdipsia. The plant is also used in the treatment of chronic rheumatism, cough, and asthma [15, 35-37].

*Stigmaphyllon emarginatum* bark is used to reduce stress, while the stems and leaves of *Stigmaphyllon sinuatum* are crushed and used for hair cleaning. As a contraceptive, the seed is swallowed.

*Tetrapteryx mucronata* is a plant used in the preparation of ayahuasca in various parts of Brazil. A narcotic drink is made from its bark. *Tristellateia australasiae* is the last species on the list, and its pounded leaves are used to cure inflammation and edoema [15]

### 5. Phytochemical constituents

Due to the presence of several secondary metabolites, Malpighiaceae is well known for its therapeutic importance. Extensive and in-depth investigations of various genus have resulted in the isolation and characterization of various secondary metabolites belonging to alkaloids, triterpenoids, anthocyanins, steroids, flavonoids, isoflavonoids, volatile constituents, phenols, and phytosterols and tannins [38-40], which are listed in **Table 1** along with their chemical structures (**Fig. 1-6**).

#### *Volatile constituents and fatty acids (Fig. 1, 2):*

The main volatile constituents identified in the essential oil (EO) of *Acridocarpus orientalis* stem were methyl 8-pimaren-18-oate (43.8 %), octacosane (5.8 %), heptacosane (4.6 %) and hexacosane (4.1 %), methyl dehydroabietate (3.9 %) and methyl pimar-8(14)-en-18-oate (3.6 %), while tetracosane (16.6 %), heptacosane (16.4 %), docosane (13.9 %), hentriacontane (13.5 %), heneicosane (10.9 %) and  $\alpha$ -pinene (7.7 %) were all found in abundance in the EO of leaves [41]. Hexadecanoic acid (39.43%), (E)-nerolidol (10.51%), triacontane (9.08%), heptacosane (5.49%) and linalool (3.23%) were found in the essential oil of *Banisteriopsis campestris* flowers, along with other constituents such as myristic acid, palmitic acid, and linoleic acid, whereas the leaves had palmitic acid (22.98%), phytol (22.98%), and triacontane (14.88%) and the stem contained

palmitic (49.79%), linoleic (11.63%), oleic (4.83%), and palmitoleic (4.15%) fatty acids; in the root included palmitic acid (57.39%), linoleic (10.38%), and oleic acids (5.47%) [42, 43].

Phytochemicals such as 4H-Pyran-4-one, 2,3-dihydro-3,5-dihydroxy-6-methyl, 1H-pyrrole-2,5-dione, 2-furancarboxaldehyde,5-(hydroxymethyl), 1-nonadecene, 3-eicosene, 2-furanmethanol, 9,12,15-octadecanoic acid, methyl ester and *n*-hexadecanoic acid were found in the fruits of *Bunchosia armeniaca* [44]. Lauric acid, linolenic acid, docosadienoic acid, myristic acid, cerotic acid, myristoleic acid, palmitic acid, palmitoleic acid, stearic acid, oleic acid, arachidonic acid, and behenic acid were identified in the pulp and seed of the *Bunchosia glandulifera* fruit. The pulp has higher concentration of fatty acids than the seed. Palmitic acid was the most prevalent. Stearic acid was also present in high concentrations. This acid was found in higher concentration in the seed than in the pulp [45].

A phytochemical analysis of *Byrsonima gardneriana* leaf extract revealed that pyroglutamic acid, octanoic acid, and other acids such as hexadecanoic and heptanoic acids predominated. Pyroglutamic acid (90.77%) and octanoic acid (76.22%) were the most common compounds found in the extract [46]. The volatile fraction acerola (*Malpighia puniceifolia*) was studied to identify 31 compounds in the mature (red) fruits, such as acethyl-methyl-carbinol, 2-methyl-propyl-acetate, limonene, E-Z-octenal, ethyl hexanoate, isoprenyl butirate and acetofenone; 23 in the intermediate (yellow), such as, methyl hexanoate, 3-octen-1-ol and hexyl butirate; and 14 in the immature (green) fruit, such as methyl-propyl-ketone, E-Z-hexenyl-acetate and 1-octadecanol [47].

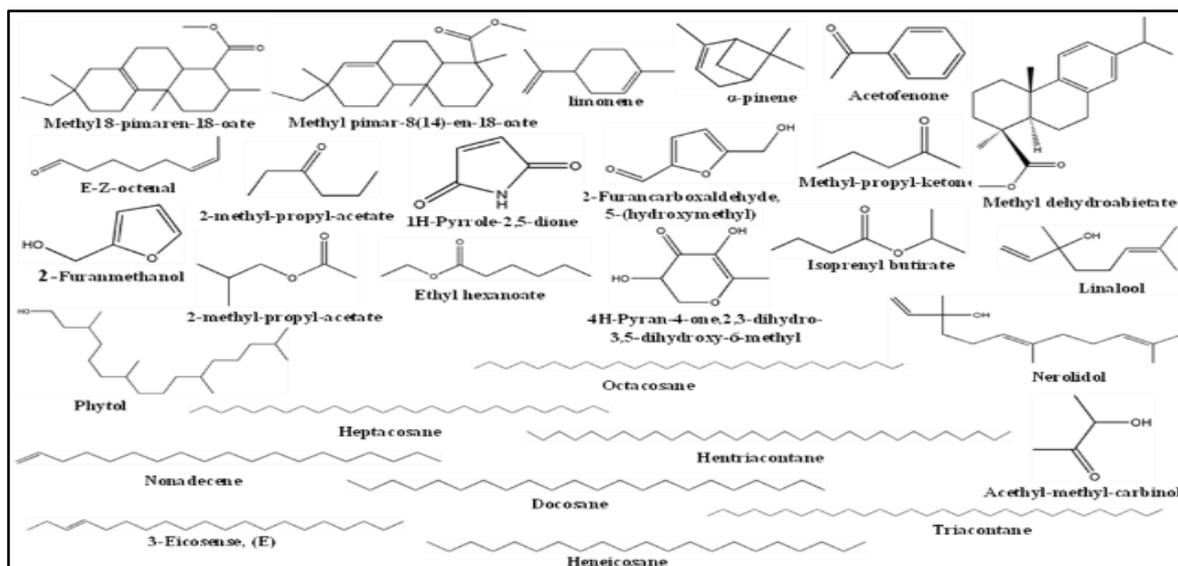


Fig. (1): Chemical structures of some volatile constituents of different genera of family Malpighiaceae

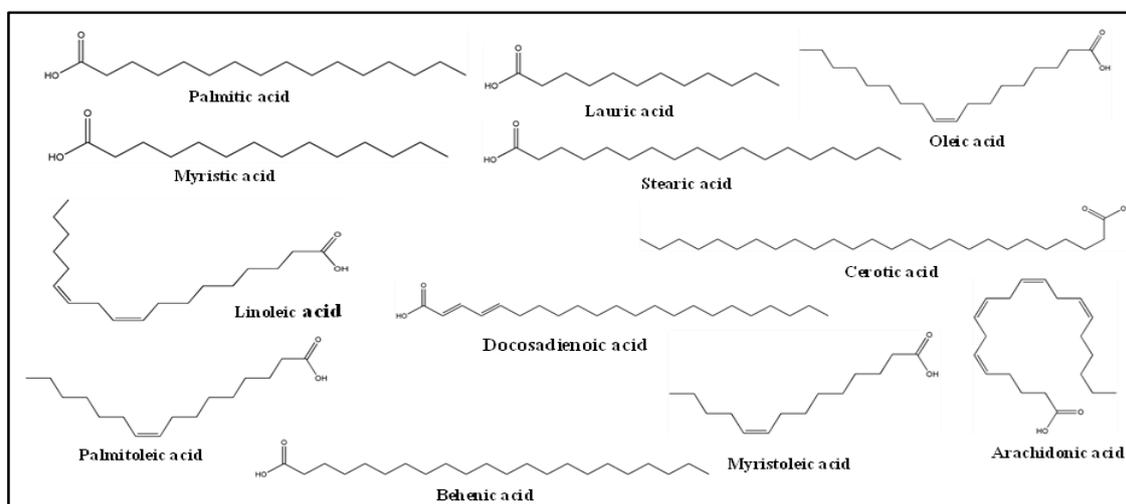


Fig. (2): Chemical structures of some fatty acids of different genera of family Malpighiaceae

#### Phenolic compounds (Fig. 3, 4):

Morin and morin-3-O-D-glucopyranoside were isolated through extraction and separation from leaves and aerial parts of *Acridocarpus orientalis* [24, 48]. The phenolic substances flavonoids and proanthocyanidins were abundant in the ethanolic extract, ethyl acetate fraction (EAF), and butanol fraction (BF) of *Banisteriopsis argyrophylla* leaves. Some of these compounds, such as catechin, quercetin-hexoside, quercetin-pentoside, and quercetin-3-O-L-rhamnose, have been identified by ESI-MS/MS in both EAF and BF due to solvent polarity similarities. However, only the EAF yielded kaempferol-3-O-L-rhamnose, quercetin-3-O-(2"-galloyl)-L-rhamnose, or quercetin-3-O-(3"-

galloyl)-L-rhamnose. Only the BF had quinic acid, the procyanidin dimer, the procyanidin dimer monogallate, and dihydroxy benzoic acid pentoside [49, 50]. Epicatechin and procyanidin B2, two known proanthocyanidins, were found in aqueous extracts of stem and stem bark *Banisteriopsis caapi* [29, 51]. By capillary electrophoresis and nuclear magnetic resonance of <sup>1</sup>H and <sup>13</sup>C, *Bunchosia armeniaca* afforded a mixture of flavonoid constituents, quercetin  $\alpha$ -L-rhamnopyranosyl-(1 $\rightarrow$ 6)- $\beta$ -D-glucopyranose (rutin), kaempferol 3-O- $\alpha$ -L-rhamnopyranoside (afzelin) and quercetin 3-O- $\beta$ -D-glucopyranoside (isoquercitrin) [52]. As shown in the Table (1), many phenolic acids and flavonoids have been extracted from several Malpighiaceae species.

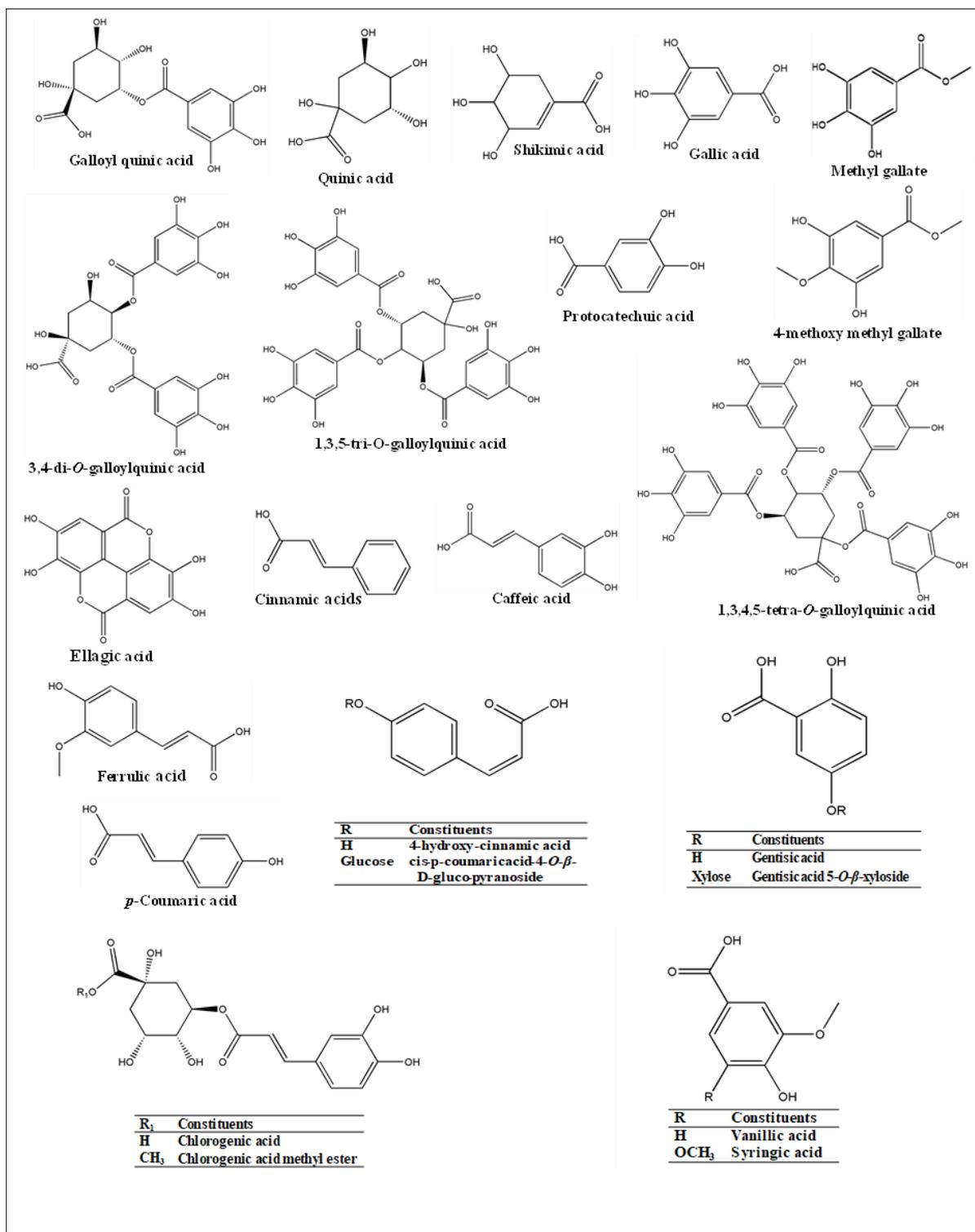


Fig. (3): Chemical structures of some phenolic acids of different genera of family Malpighiaceae

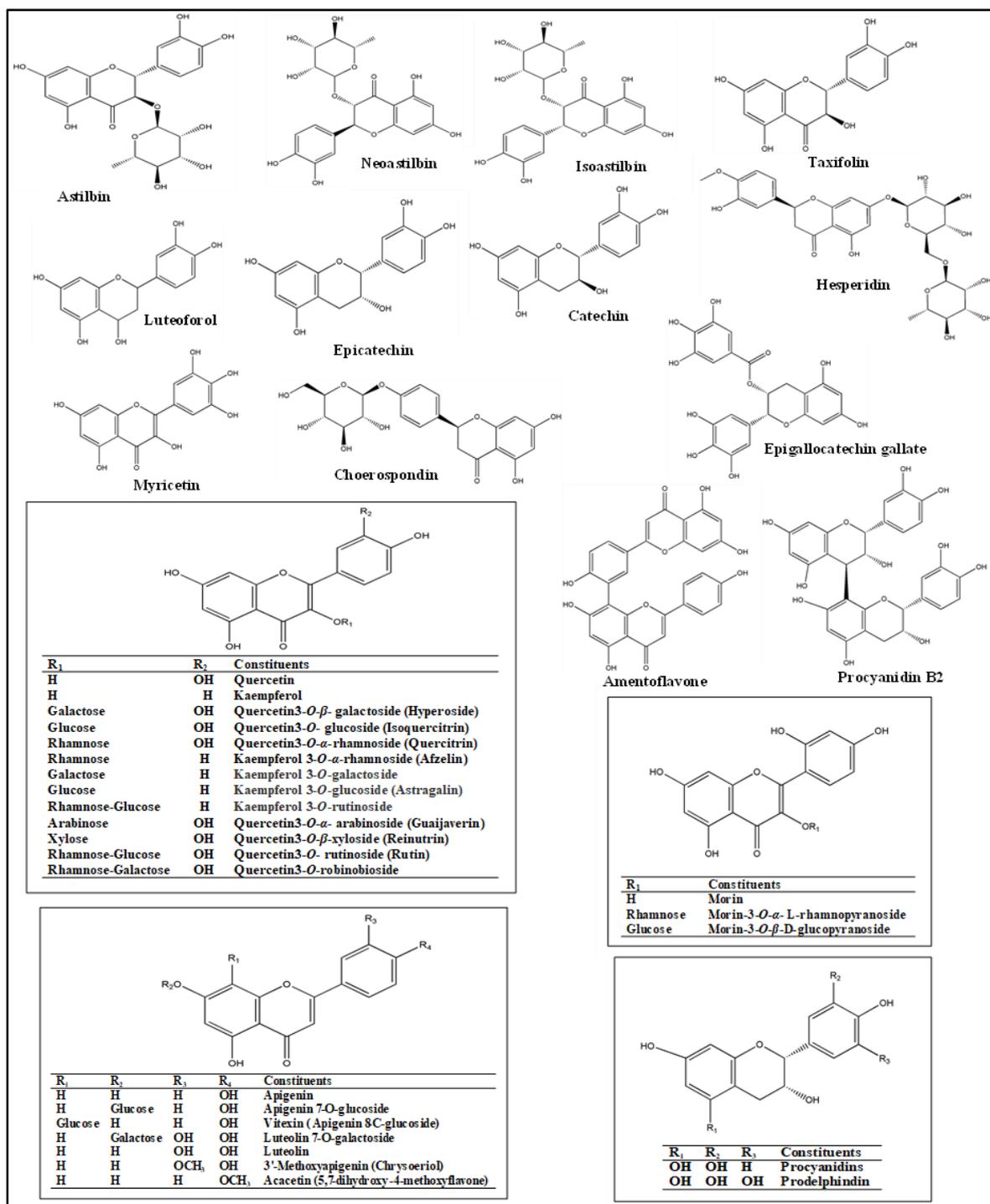


Fig. (4): Chemical structures of some flavonoids of different genera of family Malpighiaceae

#### Steroids and terpenoids (Fig. 5):

Steroids and lupane-type triterpenoids such as  $\beta$ -sitosterol,  $\beta$ -sitosterol-3-*O*- $\beta$ -D-glucopyranoside, betulin, betulinic acid isolated from methanolic extract and dichloromethane fraction of *Acridocarpus orientalis* [48, 49]. Spasmogenic bioassay-guided fractionation of methanol extract of *Byrsonima crassifolia* leaves yielded mixtures of

identified triterpenes: betulin, betulinic acid, and ursenaldehyde [50]. The potent antitubercular substances alkane dotriacontane, triterpenoids as bassic acid,  $\alpha$ -amyirin acetate, a mixture of lupeol,  $\alpha$ -amyirin,  $\beta$ -amyirin and a mixture of lupeol, and acetates of  $\alpha$ - and  $\beta$ -amyirin were isolated from the chloroform extract of *Byrsonima fagifolia* leaves using bioassay-guided fractionation [51]. Two

triterpenoids (friedelin and friedelanes), two steroids ( $\beta$ -sitosterol and sitosterol- $\beta$ -D-glucoside) were isolated from the ethyl acetate fraction of leaves of *Flabellaria paniculate* [52].

#### Alkaloids (Fig. 6):

Harmic amide, acetyl norharmine, and keto-tetrahydronorharmine, banistenoside A, banistenoside B, and their acetate, tetrahydroharmine, harmaline, and harmine were isolated from

*Banisteriopsis caapi* [28, 29, 53, 54]. The extraction and purification of bark of *Tetrapteryx mucronate* revealed the presence of alkaloid compounds as mucronatine B, 5-hydroxy-N, N-dimethyltryptamine (bufotenine), 5-methoxy-N-methyl-tryptamine, 5-methoxy-N, N di-methyl-tryptamine, trans-N feruloyl-tyramine, and 2-methyl-6-methoxy-1,2,3,4-tetrahydro- $\beta$ -carboline [55, 56].

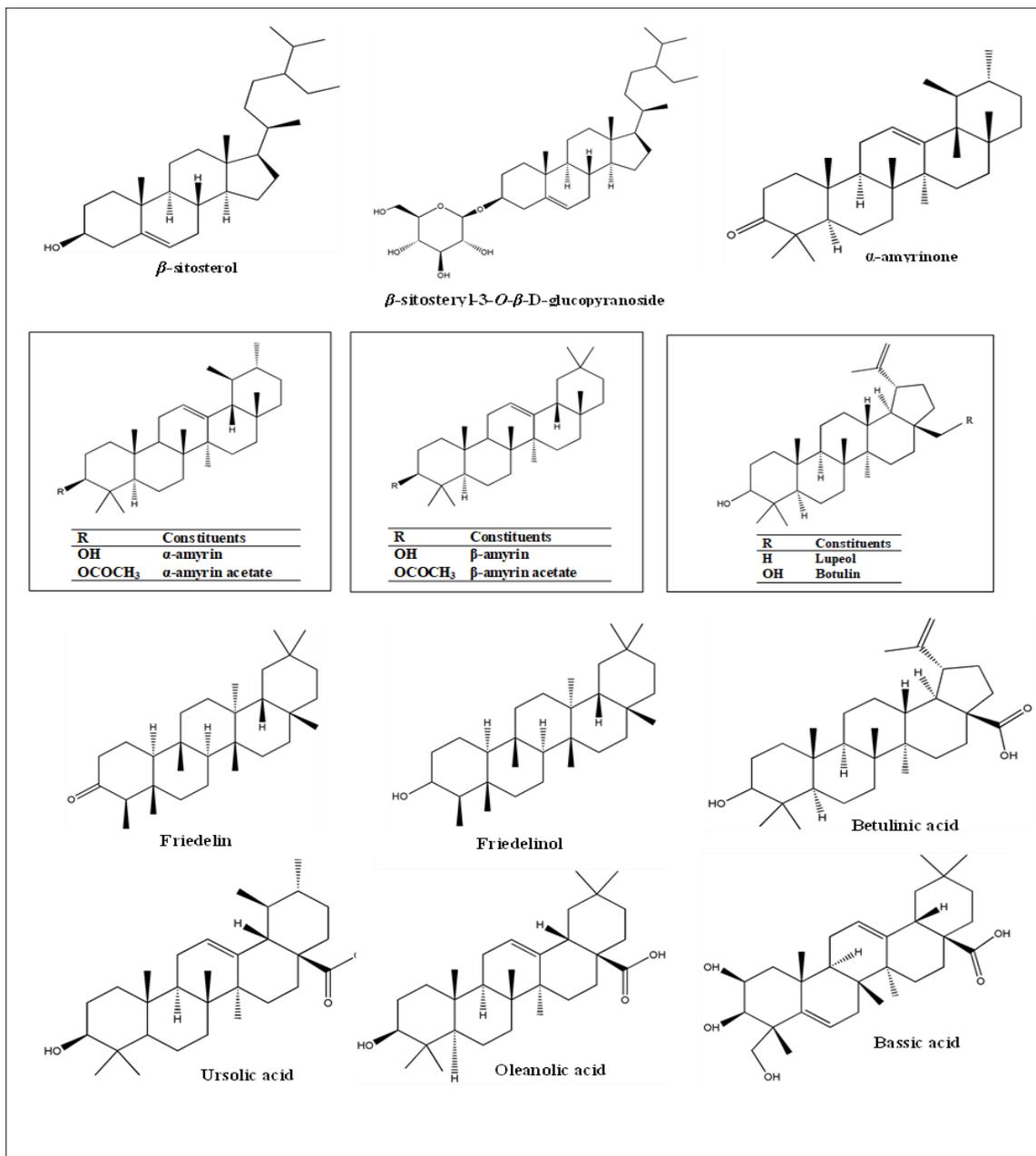


Fig. (5): Chemical structures of some steroids and triterpenoids of different genera of family Malpighiaceae

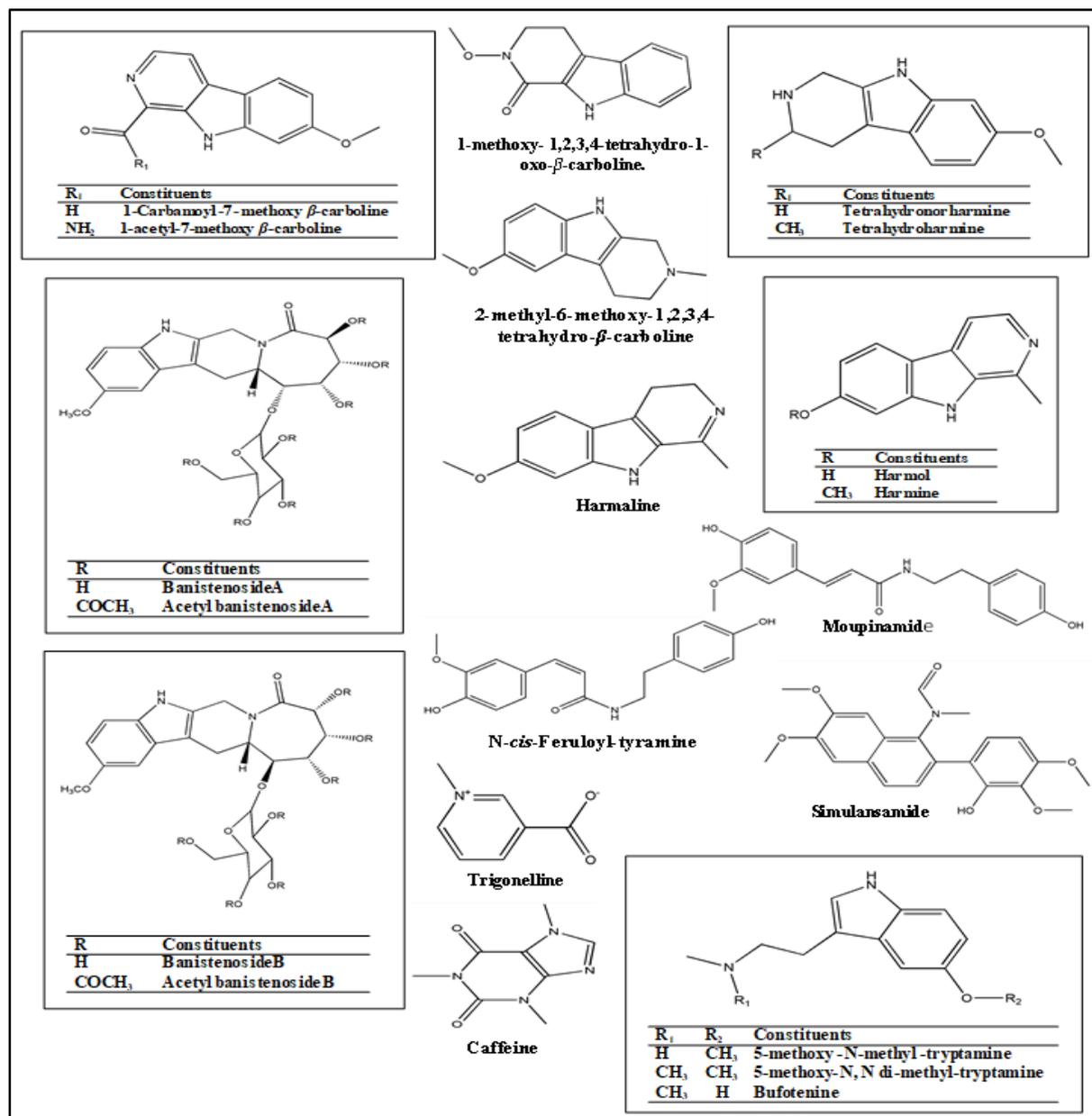


Fig. (6): Chemical structures of some alkaloids of different genera of family Malpighiaceae

### 5. Pharmacological activities

In various Malpighiaceae species, a diverse range of biological activities has been reported. Malpighiaceae have been discovered to have a wide range of biological actions, including antioxidant, antiinflammatory, antidiabetic, anticancer, antilipid peroxidation, antibacterial, hepatoprotective, neuroprotective, anxiolytic, and antidepressant properties, according to the literature. A summary of the curative activity assessments performed on this family has been represented in Table 1. These findings endorse the traditional uses of plants with respect to the pharmacological actions.

### Antioxidant activity

The antioxidant potential of the plants of family Malpighiaceae most probably is attributed to the presence of phenols and flavonoids [57]. The ethyl acetate fraction of *Acridocarpus orientalis* stem had a greater antioxidant effect than the leaves using DPPH (1,1-Diphenyl-2-picrylhydrazyl) radical scavenging activities. This is probably due to the high flavonoid and phenolic content [27]. The methanolic extract of *Aspidopterys indica* (wild.) aerial parts exhibited high DPPH scavenging antioxidant activity, but while aqueous and chloroform extracts showed a moderate effect. The ethanolic extract of *Banisteriopsis argyrophylla*

leaves exhibited the lowest IC<sub>50</sub> value in the DPPH free-radical sequestration assay, comparable to the positive BHT and ascorbic acid controls. In the ORAC (oxygen radical absorbance capacity test), the EE had a strong antioxidant activity. According to both antioxidant activity techniques (DPPH, and ORAC), the ethyl acetate fraction and *n*-butanol fraction displayed high antioxidants, with similar values to the ethanolic extract and controls [58].

*Bunchosia armeniaca* leaf ethyl acetate and *n*-butanol fractions displayed strong antioxidant activity. This implies that the scavenging action of free radicals with DPPH is mainly attributed to the presence of flavonoids content [59]. *Bunchosia armeniaca* methanolic fruit extract demonstrated a significant free radical scavenging efficacy comparable to butylated hydroxytoluene (BHT). The antioxidant activity of the fruit extract was higher than BHT on average. As a result of *in-vitro* investigation, *Bunchosia armeniaca* fruit can be considered a substantial source of antioxidant constituents [44]. Ferric ion Reducing Antioxidant Potential (FRAP), 2,2'-Azinobis-(3-Ethylbenzothiazoline-6-Sulfonic Acid (ABTS), DPPH, and electrochemical techniques were used to assess the total antioxidant capacity of *Bunchosia glandulifera* extracts. The antioxidant capacity of root and leaf ethanolic extracts was very high, followed by bark, fruit pulp, and seed extracts, which had lower values. The presence of phenolic compounds is mainly responsible for antioxidant activity [60].

The DPPH reduction assay revealed that methyl gallate and methyl *m*-trigallate fractions of leaf extract of *Byrsonima bucidaefolia* had higher antioxidant activity than vitamin C [61]. The fruit extract of Murici (*muByrsonima crassifolia*) can interfere with brain electrophysiological parameters, demonstrating a novel strategy for combating the effects of ageing in the brain. Murici contains significant chemicals that have improved antioxidant actions while also increasing protection from free radicals. It was reported that Murici extract may be useful in preventing aging-related damage, such as reactive species production [62]. The combination of *Byrsonima crassifolia* (L.) Kunth and *Spondias mombin* L. can act synergistically in the improvement of the antioxidant capacity in the beverage's development. In the DPPH, ABTS, and FRAP techniques, *Spondias mombin* L had higher antioxidant capacity than *Byrsonima crassifolia*, however there was no significant difference in ORAC [63]. In the phosphomolybdenum, DPPH, and Thiobarbituric Acid Reactive Substances (TBARS) assays, leaves of *Byrsonima duckeana* revealed high polyphenol

content and antioxidant capacity. The ethyl acetate fraction has high antioxidant capability, including free radical scavenging and lipid peroxidation inhibition, which might benefit for pain relief. In addition, chemical analysis of the ethyl acetate fraction revealed that ethyl gallate is a prominent ingredient, which could explain the high antioxidant activity detected [64].

Hydroethanolic extract of *Diplopterys pubipetala* leaves and stems demonstrated antioxidant activity. The stem extract had moderate antioxidant activity, whereas leaves had higher activity. The phytochemical investigation of *D. pubipetala* revealed the presence of flavonoids, alkaloids, and terpenes, as well as prenylated xanthenes and glycoside flavonoids, all of which contribute to the medicinal potential of the plant, which is mostly antioxidant [65]. The phosphomolybdenum assay was used to evaluate ethanol, aqueous, and chloroform extracts of *Flabellaria paniculata* leaf and root for free radical scavenging against DPPH and hydroxyl radicals, ex vivo lipid peroxidation, ferrous ion chelating activity, reducing power, and total antioxidant capacity. They had high hydroxyl radical scavenging capacity and reduced lipid peroxidation. The leaf and root extracts inhibited lipid peroxidation and scavenged hydroxyl radicals significantly. The extracts also exhibited moderate chelating properties, which could explain their affinity for iron (Fe) and thus their antioxidant properties [66].

The antioxidant activity of methanolic extracts of *Galphimia glauca* bark and leaf was investigated using DPPH Free Radical Scavenging (DFRS), Ferric ion Reducing Antioxidant Potential (FRAP), and Ferric Reducing Power (FRP) assays. Using three assays, methanolic extract of bark had somewhat better antioxidant activity than methanolic leaf extract [67]. The methanolic extract of *Malpighia glabra* L leaves showed higher antioxidant activity towards DPPH radical with IC<sub>50</sub> = 49.8(mg/ml) [68]. Immature *Malpighia emarginata* fruit alcoholic extract had stronger DPPH and ABTS scavenging activity than mature fruit one. There were strong relationships between antioxidant potential and its ascorbic acid concentration [69].

#### **Antidiabetic activity**

In  $\alpha$ -glucosidase enzyme inhibition assay, *n*-hexane, chloroform, *n*-butanol, and aqueous fractions obtained from stem of *Acridocarpus orientalis* exhibited significant inhibition. In comparison to the standard inhibitor Acarbose, the leaf aqueous fraction demonstrated modest inhibition. These findings revealed crucial details about the fractions that contain active components,

which are responsible for enzyme inhibition [27]. Along with ethanolic extract, ethyl acetate fraction and *n*-butanol fraction from *Banisteriopsis argyrophylla* leaves had higher inhibitory actions of  $\alpha$ -amylase,  $\alpha$ -glucosidase, and pancreatic lipase. The presence of phenolic substances, catechin, procyanidins, and glycosylated flavonoids produced from quercetin, kaempferol, and megastigmane glycosides can explain remarkable actions found [58].

Hexane and chloroform extracts from *Byrsonima crassifolia* fruits and seeds raised superoxide dismutase (SOD), glutathione (GSH), oxidized glutathione (GSSG), and catalase (CAT) levels, as well as hepatic glycogen content, glucose-6-phosphatase (G6Pase), and plasma insulin levels. They also reduced the levels of glucokinase (GK) and TBAR (thio- barbituric acid assay). After four hours of a single oral dose, *Byrsonima crassifolia* has considerable antihyperglycemic effects and can also improve streptozotocin-induced diabetic rats with hyperlipidemia and hyperinsulinemia. Both extracts inhibited the production of AGEs (advanced glycation end products) with IC<sub>50</sub> values ranging from 94.3 to 138.7  $\mu$ g/ml. [70]. Sesquiterpene lactone dimeric guaianolides Byrsonina A and Byrsonina B, which were isolated from hexane extracts of *Byrsonima crassifolia* seeds, have antioxidant, hypoglycemic, and hypolipidemic properties, and play a key role in blood glucose control in STZ-induced hyperglycemia by improving pancreatic islet function, increasing glycolysis, and decreasing gluconeogenesis. The mechanism of antidiabetic activity may involve an antioxidant effect, improvement in insulin resistance, and an effect on pancreatic  $\beta$ -cells to secrete insulin [71].

In alloxan-induced diabetic rats, the methanolic leaves extract of *Hiptage bengalensis* reduced blood glucose levels significantly at doses of 100 and 200 mg kg<sup>-1</sup> and had a positive effect on the lipid profile. These findings revealed that a methanolic extract of *Hiptage bengalensis* provided antihyperglycemic action in rats that was dose dependent [72]. The lipid and lipoprotein levels were significantly improved after administration of the ethanolic extract of *Hiptage bengalensis* and its fractions orally for 21 days. The extracts and fractions restored lipid and lipoprotein levels to normal levels, possibly due to its potent antidiabetic activity. After Streptozotocin (STZ) diabetic rats were treated with ethanolic extract, their urea and creatinine levels were significantly reduced, and serum total protein and albumin levels were significantly higher than normal [73].

#### **Antimicrobial activity**

*Chaetomium globosum* was mildly inhibited by *n*-hexane and aqueous fractions of *Acridocarpus orientalis*, while *Fusarium oxysporum* mold was stimulated by *n*-hexane, chloroform, *n*-butanol, and aqueous fractions. In the case of *Aspergillus niger*, none of the plant fractions impeded the fungal growth [27]. *Bunchosia glandulifera* leaves ethanolic extract had antibacterial efficacy against *Klebsiella pneumonia*. The extract was tested for antimicrobial activity using the agar well diffusion method in Muller Hinton Agar (MHA) plates [74]. *B. armeniaca* crude hydroalcoholic leaves extract showed high antibacterial activity against *S. aureus* and moderate activity against *E. coli* and *P. aeruginosa*. The activity of a flavonoid compound mixture containing rutin, afzelin, and isoquercitrin was also investigated, and it exhibited remarkable activity against all the microorganisms tested [59]. Four components of the methanolic extract of leaves from *Byrsonima crassa*, a Brazilian medicinal plant, quercetin, methyl gallate, epigallocatechin gallate, and quercetin-3-*O*-(2"-galloyl)- $\alpha$ -L-arabinopyranoside, evoked considerable antibacterial activity against tested pathogenic strains of oxacillin-resistant *S. aureus*, coagulase-negative *S. saprophyticus*, *E. coli* (two different strains), *Proteus mirabilis* and *P. aeruginosa* (two different strains) [75]. The pure triterpene basic acid with potent antitubercular activity was obtained by bioassay-directed separation of the chloroform extract of *Byrsonima fagifolia* leaves. The Microplate Alamar Blue Assay (MABA) was used to test antimycobacterial activity, and spectroscopy was used to determine the structures of interesting compounds [51].

The crude extract of *Flabellaria paniculata* possesses antimicrobial properties. *S. aureus* > *P. aeruginosa* > *K. pneumoniae* > *E. coli* were the most susceptible to the extract in that order. The chloroform fraction had the highest antibacterial activity, while petroleum ether was absolutely inactive [76]. The antibacterial activity of a methanolic extract of *Hiptage benghalensis* (L) Kurz. was investigated using the disc diffusion method, which measured the zone of inhibition and compared it to a standard antibiotic of 10  $\mu$ g tetracycline. The extract is efficacious against *K. pneumonia*, *E. coli*, *Micrococcus luteus*, and *P. aeruginosa*. On the four test organisms, the varied concentrations of the extract exhibited a zone of inhibition. The extract gave MIC value 0.625mg/ml on *K. pneumonia*, *M. luteus* and *P. aeruginosa* and 0.3125 mg/ml on *E. coli* [77].

Microdilution techniques were used to determine the antimicrobial activity of 3,4,6-tetra-*O*-(3-nitropropanoyl)-*O*-D-glucopyranoside isolated from

roots of *Heteropteris aphrodisiaca* against Gram-positive and Gram-negative bacteria, as well as Sabouraud dextrose broth for *Candida albicans*, *C. parapsilosis*, *C. krusei*, and *C. tropicalis*. The minimal inhibitory concentration (MIC) and minimal bactericidal concentration (MBC) against *Bacillus subtilis* and *Staphylococcus aureus* were 125 and 250 µg/ml and 250 and 500 µg/ml, respectively. The antifungal activity was significantly greater than the antibacterial activity, as seen by the MIC. The latter was 125 µg/ml, while the minimum fungicidal concentration (MFC) against all *Candida* species was 250 µg/ml [78]. Plaque reduction assays in cell culture were used to assess the antiviral activity of the aliphatic nitro compound (NC) isolated from *Heteropteris aphrodisiaca* against poliovirus type 1 (PV-1) and bovine herpes virus type 1 (BHV-1). In HEP-2 (human larynx carcinoma) cells, the NC had moderate antiviral activity against PV-1 and BHV-1, with 50 % inhibitory concentrations (IC<sub>50</sub>) of 22.01 µg/ml and 21.10 µg/ml, respectively [79]. The antihuman immunodeficiency virus (HIV) activity of a methanolic extract from *Heteropteris brachiata* was investigated using a non-radioactive colorimetric method that targets HIV-1 reverse transcriptase as an enzymatic target. The anticandidal effect of the extract was assessed using a standardized yeast microdilution test methodology employing the *Candida albicans* strain. The methanolic extract of *Heteropteris brachiata* has a high anticandida and moderate antiHIV impact, suggesting that the plant extract could be evaluated as a viable HIV/AIDS therapeutic candidate [80]. 1,3,4,5-tetragalloyl quinic acid was extracted from methanolic extract of *Hiraea reclinata* leaves and demonstrated antiHIV action [81]. Using an agar well diffusion assay, the methanolic extract of *Galphimia glauca* bark (GGB) and leaves (GGL) demonstrated considerable antibacterial activity against clinical and standard methicillin-resistant *Staphylococcus aureus* (ATCC 33591) (MRSA) strains. The antibacterial spectrum of both extracts (GGB and GGL) was assessed using an agar well-diffusion technique to estimate the Zone of Inhibition (ZI) against clinical and standard MRSA. ZI value for the extracts - GGB and GGL against the clinical MRSA strain was 16 ± 2 mm and 15 ± 1.5 mm respectively, while against the standard MRSA strain was 15.3 ± 0.57 mm and 14 ± 1 mm respectively, at 10 mg/ml [67]. *Malpighia glabra* methanolic leaf extract is effective against *Bacillus subtilis*, but have no effect on *Staphylococcus aureus*, *Escherichia coli*, *Pseudomonas aeruginosa*, *Aspergillus flavus*, and *Candida albicans* [82].

### Cytotoxic activity

Morin and morin-3-O-D-glucopyranoside were identified in the ethyl acetate fraction of *Acridocarpus orientalis*. When a 100-ppm concentration of morin was administered to the human hepatoma cancer cells (HepG2), colorectal adenocarcinoma (HT29, and HCT116), the viability of the cancer cells was reduced to 63.8 %, 64.5 %, and 45.3 %, respectively. Morin-3-O-D-glucopyranoside, on the other hand, reduced the viability of HepG2, HCT116, and HT29 cell lines when compared to control [24]. The treatment of colorectal adenocarcinoma (HT29 and HCT116) cell lines with chloroform and *n*-hexane fractions of *Acridocarpus orientalis* reduced cancer cell viability compared to other extracts at concentrations of 500 and 1000 g/mL. Only the chloroform fraction was found to be effective against proliferating cancer cells in human hepatoma (HepG2) cancer cells when compared to the other fractions [27]. Obcordatas A-I polyoxypregnane glycosides isolated from *Aspidopterys obcordata* Hemsl vines showed significant cytotoxicity against HuH-7 cells, and exhibited moderate cytotoxicity against the AGS and SW480 cell lines [17].

*Byrsomina crassifolia* oil had a cytoprotective effect in HepG2 cells after 72 hours of treatment, where the longer exposure time encouraged cell proliferation and prevented cell death, effectively reducing the oxidative stress induced by H<sub>2</sub>O<sub>2</sub> [83]. At 100 and 1000 µg/mL and 10 to 1000 µg/mL, chloroform, and ethyl acetate fractions of *Byrsomina duckeana* revealed decreased cell viability on the HT29 line respectively. The same fractions displayed hemolytic activity, implying that the ethanol extract's more polar elements are more closely associated to its toxicity [84]. The 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-2H-tetrazolium bromide (MTT) test was used to evaluate and estimate the IC<sub>50</sub> value for antiproliferative and cytotoxic effects of methanolic extract of *Galphimia glauca* bark (GGB) and leaf (GGL). At 24 hours, IC<sub>50</sub> value for the GGB against cancer cell proliferation (A549, SW480), and normal cell line (HEK293) was 157.8 ± 2.44 µg/ml, 136.6 ± 2.73 µg/ml, and 388.67 ± 6 µg/ml, respectively, while IC<sub>50</sub> value for the GGL against A549, SW480, and HEK293 was 194 ± 4.64 µg/ml 178 ± 3.1 µg/ml, and 317.2 ± 9.4 µg/ml, respectively [67].

The anticancer activity of *Hiptage benghalensis* methanolic leaves extract was assessed against three cancer cell cultures: human cervical carcinoma (HeLa), human breast cancer (MCF-7) and human neuroblastoma (IMR-32) cells using the MTT assay, which is based on the reduction of MTT at

different concentrations (10, 30, 100, 300, and 500 µg/ml). The extract improved the percentage inhibition of MCF7, HeLa, and IMR32 cells. Furthermore, the extract's apoptotic activity increased the production of ROS and caspase-3 activity in all cancer cell lines in a dose-dependent manner [85]. *Malpighia glabra* methanolic leaves extract significantly reduced cell viability in breast (MCF-7) and colon cell lines (HCT-116). *M. glabra* revealed no significant difference from standard doxorubicin (0.1 µg/mL), indicating that it has potent anticancer activity against colon cell line [82].

#### **Analgesic and antiinflammatory activities**

The hydroalcoholic extract of *Bunchosia armeniaca* leaves at a dose of 200 mg/kg displayed strong antiinflammatory effect, resulting in a considerable reduction in inflammation as measured by leukocyte count and myeloperoxidase enzyme activity, comparable to dexamethasone standard at 0.5 mg/kg [59]. The presence of ethyl gallate, quinic acid, gallic acid, catechin, epicatechin, quercetin, and quercetin in the leaves of *Byrsonima duckeana* reduced leukocyte migration in a carrageenan-induced peritonitis by 43% and licking time in a formalin test by 57%. The chloroform (FCL) and ethyl acetate (FEA) fractions were the most active samples in the acetic acid-induced writhing test. The hot plate test was conducted with FEA, and all the dosages tested (5, 50, and 200 mg/kg) exhibited considerable analgesic efficacy [64]. It has been observed that the stem bark of *Byrsonima japurensis* has significant and safe antiinflammatory effect, which is strongly related to its potent antioxidant activity, confirming its widespread use as an antiinflammatory drug in Amazonas State (Brazil). Only the dose of 400 mg/kg of aqueous extract showed antihyperalgesic effect in both phases of the carrageenan-induced inflammatory response, with more activity than the positive control [34].

Galphimines are nor-seco-triterpenoids with anxiolytic activities that are found in the aerial parts of *Galphimia glauca*. Anxiolytic activity of galphimines has been demonstrated in preclinical and clinical studies, and this combined with antiinflammatory activity, suggests that standardized extracts or fractions obtained from this plant could be effective for the treatment of degenerative disorders that have an inflammatory component, such as anxiety and Alzheimer's disease [86]. In Lipopolysaccharide LPS-stimulated RAW 264.7 macrophages, the antiinflammatory activity of triterpenes and steroids derivatives isolated from the chloroform stem bark fraction of *Hiptage benghalensis* was investigated, which resulted in a significant reduction in NO and PGE2 production,

as well as protein expressions of iNOS and COX-2. In LPS-stimulated RAW264.7 macrophages, they similarly found that IB protein expression increased while p-p65 protein expression and NF- $\kappa$ B transcriptional activity decreased [87].

#### **Anxiolytic and antidepressant activities**

Using recombinant human brain MAO-A and B enzymes, aqueous extracts of fresh and dried large branches of *Banisteriopsis caapi* and isolated compounds demonstrated significant inhibitory effects against MAO-A and slight effect against MAO-B. Inhibition of MAO-B activity by  $\beta$ -carbolines harmine and harmaline, in addition to potent MAO-A inhibition responsible for antidepressant action, protects against neurodegeneration and could be used to treat Parkinson's disease [53]. When compared to controls, the ayahuasca (decoction of *Psychotria viridis* and *Banisteriopsis caapi* plants) and fluoxetine groups demonstrated a significant decrease in locomotion in the open field and elevated plus-maze tests. The ayahuasca-treated animals swam more than the controls in the forced swimming test, a behaviour that was not observed in the fluoxetine group. All brain regions involved in serotonergic neurotransmission revealed increased neuronal activity in treated mice. Although there was some brain damage because of this, no permanent impairment was discovered. These findings show that high doses of ayahuasca have antidepressant properties in Wistar females, an effect that should be explored further [28].

In the forced swimming test (FST) in mice, the methanolic extract standardized on flavonoids content of *Byrsonima crassifolia* possesses potential antidepressant-like effects and might be deemed rather safe toxicologically when orally supplied. The flavonoids rutin, hesperidin, and quercetin may be implicated in the antidepressant effects [88]. The elevated plus-maze, light-dark test, and forced swimming paradigm were used to assess the anxiolytic and antidepressant-like effects of *Galphimia glauca* methanolic extract (standardised on galphimine B content) on ICR albino mice. It's impossible to rule out the possibility that the anxiolytic-like effects of *G. glauca*'s methanolic extract are due in part to GB's activity via a mechanism involving ionic channels or the regulation of the GABAA receptor in a different area than that of benzodiazepines [89]. To examine the sleep wakefulness cycle, electroencephalogram (EEG), and visual evoked potentials (VEP) in DBA/2J mice, an ethanolic extract of *Heteropterys glabra* fruits was used. The ethanolic extract reduced motor activity and altered EEG and VEP characteristics, indicating that it may perform as an anxiolytic/sedative agent [90]. In the forced

swimming test in mice, the methanolic extract of *Heteropterys cotinifolia* shows a dose-dependent antidepressant effect at doses range from 31 to 310 mg/kg, with no reduction in mice locomotion [91].

#### **Hepatoprotective activity**

The liver marker enzymes serum alanine transaminase (ALT) and aspartate transaminase (AST) were significantly reduced in almost all concentrations after pretreatment with *Acridocarpus orientalis* ethanolic extract. Furthermore, serum reduced glutathione (GSH) levels in *A. orientalis* medicated mice groups were considerably higher. At a dose of 250 mg/kg BW, a reduction in liver weights in pretreated mice with *A. orientalis* indicated substantial weight loss and the histological liver study revealed a near-normal repair of liver architecture [26]. *Hiptage benghalensis* methanolic leaves extract (MEHB) showed hepatoprotective efficacy in rats against carbon tetrachloride-induced liver damage that was comparable to the standard medication silymarin (50 mg/kg). The markers of high serum liver damage enzymes such as aspartate transaminase, alanine transaminase, total bilirubin, and alkaline phosphatase were considerably reduced ( $p < 0.01$ ) after methanolic extract administration (200 mg & 400 mg / kg). MEHB also displayed strong antioxidant effects by increasing glutathione levels, as well as free radical scavenging activities, according to the studies [92]. The dose of 800 mg/kg of *Malpighia glabra* methanolic leaves extract had the greatest hepatoprotective effect, lowering elevated serum levels of ALT, AST, NO, and TNF- $\alpha$  liver content by 26, 24, 23, and 42 %, respectively, while also significantly increasing serum catalase levels by 102 %. When compared to silymarin, all doses tested (200, 400, and 800 mg/kg) demonstrated a greater reduction in serum TNF- $\alpha$ , indicating their significant antiinflammatory activity. The leaves of *M. glabra* were shown to be a rich source of secondary metabolites and to have considerable hepatoprotective properties [93].

#### **Gastroprotective activity**

*Byrsonima fagifolia* methanolic leaf extract effectively reduced stomach lesions generated by ethanol and HCl/ethanol, and endogenous mucosal sulphhydryl groups contributed efficaciously to BF gastro-protection. *B. fagifolia* inhibited the progression of the inflammatory process and possesses antidiarrheal properties. With negligible toxicity, this extract expedited the healing of gastric ulcerated mucosa by activating proliferative factors and enhancing gastric mucus production [94]. The methanolic extract of *Byrsonima intermedia* (MBI) leaves completely prevented gastric and duodenal

lesions (69%) and completely repaired gastric (49%) and duodenal lesions (45%) on 7 and 14 days. Endogenous sulphhydryl compounds, vanilloid receptors, and an elevation in GSH level are all involved in *B. intermedia*'s gastroprotective action, resulting in effective gastric and duodenal protection. MBI had antidiarrheal effects that were both curative (42%) and preventative (49%) when opiate receptors were involved [95]. *Byrsonima intermedia* ethyl acetate (EtOAc) and water (AcoAq) both reduced gastric lesions, but AcoAq was more effective than EtOAc in terms of anti-*Helicobacter pylori* activity, as well as protecting the gastric mucosa from ethanol, non-steroidal antiinflammatory drugs (NSAIDs), and cysteamine-induced duodenal mucosal damage. After acetic acid damage, both partitions were linked to a considerable increase in gastric and duodenal repair, as well as increased stomach mucosal GSH content. However, after 6 days of treatment, EtOAc was more efficient than AcoAq in reducing stomach damage following the start of the gastric I/R, which was accompanied by a significant decrease in gastric mucosal MPO, IL-1, and TNF- $\alpha$  activity, as well as an increase in IL-10 and GSH content [96]. In indomethacin and pylorus ligation-induced ulcer models, the methanolic extract of *Flabellaria paniculata* and the ethyl acetate fraction from this extract showed significant gastroprotective effects [52].

#### **Miscellaneous activities**

##### **Anti-Alzheimer activity**

In aged rats, treatment with standardized root extract of *Heteropterys aphrodisiaca* for 7 days or longer improves learning and memory deficits. The memory deficits in the passive avoidance test were restored after treatment with standard extract for 7 days (50 mg/kg) or 26 days (100 mg/kg). However, after acute administration of standard extract (100 mg/kg) to aged rats, there was no improvement in memory [97]. The ethanolic extract of the bark of *Tetrapterys mucronate* exhibited *in-vitro* acetylcholinesterase (AChE) inhibition in TLC bioautography assay [55].

##### **Antilipid peroxidation**

When assayed with the Thiobarbituric Acid Reactive Substance (TBARS) Test, the crude extract, and fractions of *Acridocarpus orientalis* leaves revealed a higher proportion of oxidative degradation of lipids than the stem extract and other fractions. Aqueous and chloroform fractions of leaves showed higher inhibition of 60.6% and 49.9%, respectively. In case of TBARS bioassay of stem, ethyl acetate (34.5%) and chloroform (34.2%)

fractions revealed higher percentages of lipid peroxidation than the other fractions [27].

#### **Antiprotozoal activity**

Using arginase (ARG) from *Leishmania amazonensis* as a molecular target, leishmanicidal compounds from *Byrsonima coccolobifolia* leaf and stem ethanolic extracts were identified as flavonoids. They inhibited the enzyme with IC<sub>50</sub> values ranging from 0.9 to 4.8 µM and were discovered to be non-competitive ARG inhibitors with dissociation constants (K<sub>i</sub>) ranging from 0.24 to 3.8 µM, indicating high affinity. Studies of the structural characteristics of flavonoids linked to ARG action revealed significant commonalities [98]. Quercetin was only substance isolated from methanolic extract of *Galphimia glauca* aerial parts that had antiprotozoal activity, and it was weak. The IC<sub>50</sub> values were 14 µM against *Plasmodium falciparum* K1, 13.2 µM against *Trypanosoma brucei*, and 63.8 µM against *Leishmania donovani* [99].

#### **Antiobesity activity**

For 40 days, rats were given ethanolic extract of *Hiptage madablota* root orally at doses of 100, 200, and 400 mg/kg, which resulted in a significant decrease in food intake, body weight, lee index, serum lipids, atherogenic index, and coronary risk index, as well as an inverse increase in brain serotonin. As a result of its hypophagic and hypolipidemic actions, *Hiptage madablota* root extract was found to have strong antiobesity efficacy and to increase brain serotonin levels in rats fed a high-fat diet [100].

#### **Wound healing activity**

*Flabellaria paniculata* methanol leaf extract resulted in sic wound contraction and a shorter epithelisation duration. On day 14, which extract, and chloroform fraction achieved 100 % wound contraction in non-infected and *Staphylococcus aureus* groups, whereas on day 18, *Pseudomonas aeruginosa* group obtained 100% wound contraction. The extract had antiinfective and wound-healing properties, justifying the plant's usage in the treatment of skin illnesses and sores on a local level. When compared to the aqueous fraction, the chloroform fraction revealed extremely significant wound healing characteristics in the non-infection group. The proportion of wound contraction of the chloroform fraction in the *Staphylococcus aureus* infected group is like that of the reference drug, as evidenced by epithelization times of 16 days and 17 days for the reference drug and chloroform fraction, respectively. The *Pseudomonas aeruginosa* infected group's data revealed that the chloroform fraction was also significantly more potent than controls [101, 102].

Plants of Malpighiaceae have long been used as ayurvedic treatment of many illnesses, this article gives collective information concerning the pharmacological activities of these plants, giving scientific evidence for their use in management of various diseases, as well as their phytochemical constituents. Hopefully, these plants will be more profoundly used in medicinal treatments in the future.

#### **Conclusion**

As previously stated, the Malpighiaceae family appears to contain a wide range of active constituents, including alkaloids (harmine, harmaline, caffeine, and tetrahydroharmine), Flavonoids ( rutin, vitexin, quercitrin, Isoquercitrin, catechin, epicatechin, quercetin and kaempferol), vitamine C, and terpenoids ( $\alpha$ -amyrin,  $\beta$ -amyrin and their acetates, lupeol, oleanolic acid, ursolic acid and  $\alpha$ -amyrinone). Pharmacologically, Malpighiaceae has the most potent effect on neurodegenerative disorders including Parkinson's Disease through MAO inhibition and antioxidant actions, as well as cytotoxic and inhibitory effects on NO generation. Also, they are medicinally used as antileishimina, antimicrobial, antiulcerogenic, antitubercular, antioxidant, antidepressant, wound healing, spasmogenic, antidiabetic, CNS stimulants and antiinflammatory properties. As a result, for the first time, this article presents a study of Malpighiaceae plants that contain a variety of active compounds that are effective against a variety of diseases. Hopefully, we will be able to use these plants in medicinal therapies in the future.

#### **Conflict of interest**

The author declare no conflict of interest

#### **References**

1. Rates S.M.K., Plants as source of drugs. *Toxicon*, **39**(5), 603-613 (2001). Doi: 10.1016/S0041-0101(00)00154-9.
2. Anderson C., The identity of two water-dispersed species of Heteropterys (Malpighiaceae): *H. leona* and *H. platyptera*. *Contr. Univ. Michigan Herb.*, **23**, 35-47 (2001).
3. Lawrence G.H.M., Taxonomy of vascular plants, 1951, Macmillan, New York, p. 823 (2017).
4. Anderson W.R., Byrsonimoideae, a new sub-family of Malpighiaceae. *Biotropica*, **7**, 5-18 (1977).
5. Davis C.C., Anderson W.R., and Donoghue M.J., Phylogeny of Malpighiaceae: evidence from chloroplast *ndhF* and *trnL-F* nucleotide sequences. *American journal of botany*, **88**(10), 1830-1846 (2001). Doi: 10.2307/3558360.
6. Group A.P., An update of the Angiosperm Phylogeny Group classification for the orders and families of flowering plants: APG III. *Botanical*

- Journal of the Linnean Society*, **161**(2), 105-121 (2009). Doi: 10.1111/j.1095-8339.2009.00996.x.
7. USDA N. *USDA plants database*. URL: <http://plants.usda.gov/java/>. 2014.
  8. Awoerson W.R., The origin of the Malpighiaceae-The evidence from morphology. *Mem. N. Y. Bot. Gard.*, **64**, 210-224 (1990).
  9. Gates B., Banisteriopsis, Diplopterys (Malpighiaceae). *Flora Neotropica*, **30**, 1-237 (1982).
  10. Makino-Watanabe H., Melhem T., and Barth O., Morfologia dos grãos de pólen de espécies brasileiras de Janusia. *Adr. Juss. e Schwannia Endl.. Hoehnea*, **20**, 79-86 (1993).
  11. Karunasena G., Chandrajith V., and Nawaratne S., Physicochemical Characteristics of Pea Nut Butter Fruit (*Bunchosia armeniaca*). *International Journal of Food Science and Nutrition*, **3**(3), 46-51 (2018b).
  12. Da Silva Nunes R., Silva Kahl V.F., Da Silva Sarmiento M., Richter M.F., Abin- Carriquiry J.A., Martinez M.M., De Barros Falcão Ferraz A., and Da Silva J., Genotoxic and antigenotoxic activity of acerola (*Malpighia glabra* L.) extract in relation to the geographic origin. *Phytotherapy Research*, **27**(10), 1495-1501 (2013). Doi: 10.1002/ptr.4896.
  13. Lombello R.A. and Forni-Martins E.R., Malpighiaceae: correlations between habit, fruit type and basic chromosome number. *Acta Botanica Brasilica*, **17**(2), 171-178 (2003). Doi: 10.1590/S0102-33062003000200001.
  14. Heywood V.H., Brummitt R., Culham A., and Seberg O., Flowering plant families of the world, Vol. 88, Firefly Books, Ontario, p. 424 (2007).
  15. Quattrocchi U., CRC world dictionary of medicinal and poisonous plants: common names, scientific names, eponyms, synonyms, and etymology, 5 Volume Set. CRC press, Taylor & Francis group, London, NewYork, p. 3960 (2012).
  16. Leon M.E., Catalog of useful species of Malpighiaceae family in the state of Mexico and surrounding areas, *Undergraduate Thesis*, Faculty of Higher Iztacala, Mexico (2005).
  17. Hu M., Li Y., Sun Z., Huo X., Zhu N., Sun Z., Liu Y., Wu H., Xu X., and Ma G., New polyoxyypregnane glycosides from *Aspidopterys obcordata* vines with antitumor activity. *Fitoterapia*, **129**, 203-209 (2018). Doi: 10.1016/j.fitote.2018.07.003.
  18. Li Y., Ma G., Lv Y., Su J., Li G., and Chen X., Efficacy of obcordata A from *Aspidopterys obcordata* on kidney stones by inhibiting NOX4 expression. *Molecules*, **24**(10), 1957 (2019). Doi: 10.3390/molecules24101957.
  19. Rui W., Qi Y., Nengyu C., and Guolin Z., Study on the chemical constituents of *Aspidopterys obcordata* Hemsl. *Natural Product Research Development*, **13**(1), 14-16 (2001).
  20. Yihang L., Guang L., Meifang S., Xuelan L., Xia Z., Juan L., and Xi C., Acute toxicity study of *Aspidopterys obcordata* aqueous extract in Sprague-Dawley rats. *Journal of Traditional Chinese Medicine*, **36**(3), 377-381 (2016). Doi: 10.1016/S0254-6272(16)30052-8.
  21. Kale O., Awodele O., and Akindele A., *Acridocarpus smeathmannii* (DC.) Guill. & Perr. Root enhanced reproductive behavior and sexual function in male wistar rats: Biochemical and pharmacological mechanisms. *Journal of ethnopharmacology*, **230**, 95-108 (2019). Doi: 10.1016/j.jep.2018.10.024.
  22. Van Andel T., Croft S., Van Loon E., Quiroz D., Towns A., and Raes N., Prioritizing West African medicinal plants for conservation and sustainable extraction studies based on market surveys and species distribution models. *Biological Conservation*, **181**, 173-181 (2015). Doi: 10.1016/j.biocon.2014.11.015.
  23. Catarino L., Havik P.J., and Romeiras M.M., Medicinal plants of Guinea-Bissau: Therapeutic applications, ethnic diversity and knowledge transfer. *Journal of ethnopharmacology*, **183**, 71-94 (2016). Doi: 10.1016/j.jep.2016.02.032.
  24. Hussain J., Ali L., Khan A.L., Rehman N.U., Jabeen F., Kim J.-S., and Al-Harrasi A., Isolation and bioactivities of the flavonoids morin and morin-3-O- $\beta$ -D-glucopyranoside from *Acridocarpus orientalis*—a wild Arabian medicinal plant. *Molecules*, **19**(11), 17763-17772 (2014). Doi: 10.3390/molecules191117763.
  25. Ksiksi T. and Hamza A.A., Antioxidant, lipoxigenase and histone Deacetylase inhibitory activities of *Acridocarpus orientalis* from Al Ain and Oman. *Molecules*, **17**(11), 12521-12532 (2012). Doi: 10.3390/molecules171112521.
  26. Lotfy M., Al-Hammadi R., Palakkott A.R., Yasin J., Al-Hammadi S., and Ksiksi T., Hepatoprotective potentials of *Acridocarpus orientalis* in mice. *Clinical Phytoscience*, **6**(38), 1-9 (2020a). Doi: 10.1186/s40816-020-00184-x.
  27. Rehman N.U., Mabood F., Khan A.L., Ali L., Gillani S.A., Abbas G., Khan A., Al-Harrasi A., and Hussain J., Evaluation of biological potential and physicochemical properties of *Acridocarpus orientalis* (Malpighiaceae). *Pak. J. Bot.*, **51**(3), 1099-106 (2019b). Doi: 10.30848/PJB2019-3(8).
  28. Pic-Taylor A., Da Motta L.G., De Moraes J.A., Junior W.M., Santos A.D.F.A., Campos L.A., Mortari M.R., Von Zuben M.V., and Caldas E.D., Behavioural and neurotoxic effects of ayahuasca infusion (*Banisteriopsis caapi* and *Psychotria viridis*) in female Wistar rat. *Behavioural processes*, **118**, 102-110 (2015). Doi: 10.1016/j.beproc.2015.05.004.
  29. Wang Y.-H., Samoylenko V., Tekwani B.L., Khan I.A., Miller L.S., Chaurasiya N.D., Rahman M.M., Tripathi L.M., Khan S.I., and Joshi V.C., Composition, standardization and chemical profiling of *Banisteriopsis caapi*, a plant for the treatment of neurodegenerative disorders relevant to Parkinson's disease. *Journal of ethnopharmacology*, **128**(3), 662-671 (2010). Doi: 10.1016/j.jep.2010.02.013.
  30. Bouso J.C., González D., Fondevila S., Cutchet M., Fernández X., Ribeiro Barbosa P.C., Alcázar-Córcoles M.Á., Araújo W.S., Barbanoj M.J., and Fábregas J.M., Personality, psychopathology, life attitudes and neuropsychological performance

- among ritual users of ayahuasca: a longitudinal study. *journals.plos.org.*, **7**(8), (2012). Doi: 10.1371/journal.pone.0042421.
31. Politi M., Friso F., Saucedo G., and Torres J., Traditional use of *Banisteriopsis caapi* alone and its application in a context of drug addiction therapy. *Journal of Psychoactive Drugs.*, **53**(1), 76-84 (2021). Doi: 10.1080/02791072.2020.1820641.
  32. Giraldo M. and Hanazaki N., Uso e conhecimento tradicional de plantas medicinais no Sertão do Ribeirão, Florianópolis, SC, Brasil. *Acta botanica brasílica*, **24**(2), 395-406 (2010).
  33. Oliveira R.L.C.D., Scudeller V.V., and Barbosa R.I., Use and traditional knowledge of *Byrsonima crassifolia* and *B. coccolobifolia* (Malpighiaceae) in a Makuxi community of the Roraima savanna, northern Brazil. *Acta Amazonica*, **47**(2), 133-140 (2017). Doi: 10.1590/1809-4392201600796
  34. Guilhon-Simplicio F., De Souza Pinheiro C.C., Conrado G.G., Dos Santos Barbosa G., Dos Santos P.A., De Meneses Pereira M., and Lima E.S., Anti-inflammatory, anti-hyperalgesic, antiplatelet and antiulcer activities of *Byrsonima japurensis* A. Juss.(Malpighiaceae). *Journal of ethnopharmacology*, **140**(2), 282-286 (2012). Doi: 10.1016/j.jep.2012.01.018.
  35. Bobbarala V., Katikala P.K., Naidu K.C., and Penumajji S., Antifungal activity of selected plant extracts against phytopathogenic fungi *Aspergillus niger* F2723. *Indian Journal of Science and Technology*, **2**(4), 87-90 (2009). Doi:10.17485/ijst%2F2009%2Fv2i4%2F2943.
  36. Parrotta J.A., Healing plants of peninsular India, CABI publishing, Rio Piedras, Puerto Rico, p. 917 (2001).
  37. Chatterjee A. and Pakrashi S.C., Treatise on Indian medicinal plants, Publications & Information Directorate, Vol. 3, New Delhi (1991).
  38. De Frias U.A., Costa M.C.M., Takahashi J.A., and Oki Y., *Banisteriopsis* species: a source of bioactive of potential medical application. *International Journal of Biotechnology for Wellness Industries*, **1**, 163-171 (2012).
  39. Liu J.-Q., Deng Y.-Y., Li T.-Z., Han Q., Li Y., and Qiu M.-H., Three new tetranorditerpenes from aerial parts of acerola cherry (*Malpighia emarginata*). *Molecules*, **19**(2), 2629-2636 (2014). Doi: 10.3390/molecules19022629.
  40. Vendramini A.L.A. and Trugo L.C., Phenolic compounds in acerola fruit (*Malpighia puniceifolia*, L.). *Journal of the Brazilian Chemical Society*, **15**(5), 664-668 (2004). Doi: 10.1590/S0103-50532004000500009
  41. Rehman N.U., Alsabahi J.N., Alam T., Khan A., Rafiq K., Khan M., and Al-Harrasi A., Chemical Constituents and Carbonic Anhydrase II Activity of Essential Oil of *Acridocarpus orientalis* A. Juss. in Comparison With Stem and Leaves. *Journal of Essential Oil Bearing Plants*, **24**(1), 68-74 (2021). Doi: 10.1080/0972060X.2021.1873195.
  42. Rocha E., Cunha L., Silva M., Freitas T., Nascimento E., Silva L., Aquino F., Martins C., Chang R., and Morais S., Composição química e atividade antimicrobiana do óleo essencial das flores de *Banisteriopsis campestris* (A. Juss.) Little. *Revista Virtual de Química*, **10**(5), 1562-1577 (2018). Doi: 10.2174/1573407216666200129101433.
  43. Rocha E.D.O., Chang R., Do Nascimento E.A., Martins M.M., De Morais S.A., De Aquino F.J.T., Cunha L., Silva L.D.O., Martins C.H., and Teixeira T.L., Chemical Composition and Bioactive Potential of Essential Oils from *Banisteriopsis campestris*. *Current Bioactive Compounds*, **16**(8), 1205-1214 (2020). Doi: 10.2174/1573407216666200129101433.
  44. Premathilaka R. and Silva M., Bioactive Compounds and Antioxidant Activity of *Bunchosia armenica*. *World Journal of Pharmacy and Pharmaceutical Sciences*, **5**(10), 1237-1247 (2016). Doi: 10.20959/wjpps201610-7783.
  45. Blank D.E., Fraga S., Bellaver M., Santos C.E.I.D., Costa L., and Moura N., Proximate Composition, Nutrient Mineral and Fatty Acid of the *Bunchosia glandulifera* Fruit. *Journal of Food and Nutrition Research*, **5**(8), 575-578 (2017). Doi: 10.12691/jfnr-5-8-7.
  46. De Souza-Melo W.O., Figueiredo-Júnior E.C., Freire J.C.P., Costa B.P., Lira A.B., Freires I.A., Cavalcanti Y.W., Lopes W.S., Tavares J.F., and Pessôa H.D.L.F., Phytochemistry, antifungal and antioxidant activity, and cytotoxicity of *byrsonima gardneriana* (A. Juss) extract. *Archives of Oral Biology*, **123**, 104994 (2021). Doi: 10.1016/j.archoralbio.2020.104994.
  47. Vendramini A.L. and Trugo L.C., Chemical composition of acerola fruit (*Malpighia puniceifolia* L.) at three stages of maturity. *Food chemistry*, **71**(2), 195-198 (2000). Doi: 10.1016/S0308-8146(00)00152-7.
  48. Jamshidi-Adegani F., Vakilian S., Rehman N.U., Al-Broumi M., Al-Kindi J., Alam K., Mozafarinahavandi P., Hasan A., Al-Riyami H., and Hussain J., Secondary metabolites from *acridocarpus orientalis* inhibits 4T1 cells and promotes mesenchymal stem cells (MSCs) proliferation. *Molecular Biology Reports*, **47**(7), 5421-5430 (2020). Doi: 0.1007/s11033-020-05632-y.
  49. Rehman N.U., Hussain H., Ali L., Khan A., Mabood F., Shinwari Z.K., Hussain J., and Al-Harrasi A., Chemical constituents of *acridocarpus orientalis* and their chemotaxonomic significance. *Chemistry of Natural Compounds*, **55**(3), 586-588 (2019a). Doi: 10.1007/s10600-019-02752-1.
  50. Bejar E., Amarquaye A., Che C.-T., Malone M.H., and Fong H.H., Constituents of *Byrsonima crassifolia* and their spasmogenic activity. *International journal of pharmacognosy*, **33**(1), 25-32 (1995). Doi: 10.3109/13880209509088143.
  51. Higuchi C.T., Sannomiya M., Pavan F.R., Leite S., Sato D., Franzblau S., Sacramento L.V.S.D., Vilegas W., and Leite C.Q.F., *Byrsonima fagifolia* Niedenzu apolar compounds with antitubercular activity. *Evidence-Based Complementary and Alternative Medicine*, **2011**, (2011). Doi: 10.1093/ecam/nen077.
  52. Sofidiya M.O., Taiwo E., Awolola V., Habila J., and Koorbanally N.A., Gastroprotective Effect and

- Chemical Constituents of *Flabellaria paniculata* (Malpighiaceae). *Natural Product Communications*, **14**(7), 1934578-19860342 (2019). Doi: 10.1177/1934578X19860342.
53. Samoylenko V., Rahman M.M., Tekwani B.L., Tripathi L.M., Wang Y.-H., Khan S.I., Khan I.A., Miller L.S., Joshi V.C., and Muhammad I., *Banisteriopsis caapi*, a unique combination of MAO inhibitory and antioxidative constituents for the activities relevant to neurodegenerative disorders and Parkinson's disease. *Journal of ethnopharmacology*, **127**(2), 357-367 (2010). Doi: 10.1016/j.jep.2009.10.030.
  54. Santos K.T., Almeida C.A., Souza L.R., De Paula A.M.B., De Oliveira D.A., and De Andrade Royo V., *In vitro* Antitumor Effect on Melanoma Cell Line and Chemical Composition of *Diplopterys pubipetala* (A. Juss) WR Anderson and C. Davis. *Pharmacognosy Reviews*, **14**(28), 146-154 (2020). Doi: 10.5530/phrev.2020.14.18.
  55. Queiroz M.M.a.F., Queiroz E.F., Zeraik M.L., Ebrahimi S.N., Marcourt L., Cuendet M., Castro-Gamboa I., Hamburger M., Da Silva Bolzani V., and Wolfender J.-L., Chemical composition of the bark of *Tetrapteryx mucronata* and identification of acetylcholinesterase inhibitory constituents. *Journal of natural products*, **77**(3), 650-656 (2014). Doi: 10.1021/np401003p.
  56. Queiroz M.M.F., Marti G., Queiroz E., Marcourt L., Castro-Gamboa I., Bolzani V.D.S., and Wolfender J.L., LC-MS/MS quantitative determination of *Tetrapteryx mucronata* alkaloids, a plant occasionally used in Ayahuasca preparation. *Phytochemical Analysis*, **26**(3), 183-188 (2015). Doi: 10.1002/pca.2548.
  57. Chandrika P.U. and Sunitha K., Pharmacognostic Evaluation, Estimation of Phenolic, Flavonoid Composition and Antioxidant Activity of *Aspidopteryx indica* (Willd.) W. Theob: An Endemic Plant to Peninsular India. *Annals of the Romanian Society for Cell Biology*, **25**(4), 13884-13891 (2021).
  58. Quaresma D.M., Justino A.B., Sousa R.M., Munoz R.A., De Aquino F.J., Martins M.M., Goulart L.R., Pivatto M., Espindola F.S., and De Oliveira A., Antioxidant compounds from *Banisteriopsis argyrophylla* leaves as  $\alpha$ -amylase,  $\alpha$ -glucosidase, lipase, and glycation inhibitors. *Bioorganic Chemistry*, **105**, 104335 (2020). Doi: 10.1016/j.bioorg.2020.104335.
  59. Queiroz G.S., Flavonoides de *Bunchosia armeniaca* e derivados de 2-arilideno-1-a-tetralona: obtenção e atividades biológicas. (2012).
  60. De Menezes Peixoto C.R., Fraga S., Da Rosa Justim J., Gomes M.S., Carvalho D.G., Jarenkow J.A., and De Moura N.F., Voltammetric determination of total antioxidant capacity of *Bunchosia glandulifera* tree extracts. *Journal of electroanalytical chemistry*, **799**, 519-524 (2017). Doi: 10.1016/j.jelechem.2017.07.003.
  61. Castillo-Avila G.M., García-Sosa K., and Peña-Rodríguez L.M., Antioxidants from the leaf extract of *Byrsonima bucidaefolia*. *Natural product communications*, **4**(1), 83-86 (2009). Doi: 10.1177/1934578X0900400118.
  62. Sousa M.S.B. and De Souza Buarque D., Murici (*Byrsonima crassifolia* (L.) Kunth): Antioxidant effects and application to aging, Aging, Elsevier, Chap. 25, p. 259-265 (2020).
  63. Aniceto A., Montenegro J., Cadena R.D.S., and Teodoro A.J., Physicochemical Characterization, Antioxidant Capacity, and Sensory Properties of Murici (*Byrsonima crassifolia* (L.) Kunth) and Taperebá (*Spondias mombin* L.) Beverages. *Molecules*, **26**(2), 332 (2021). Doi: 10.3390/molecules26020332.
  64. Verdam M.C.D.S., Guilhon-Simplicio F., Andrade K.C.D., Fernandes K.L.M., Machado T.M., Da Silva F.M.A., Souza M.P.D., Koolen H.H.F., Paula C.D.S., and Hirota B.C.K., Analgesic, anti-inflammatory, and antioxidant activities of *Byrsonima duckeana* WR Anderson (Malpighiaceae). *The Scientific World Journal*, **2017**, (2017). Doi: 10.1155/2017/8367042.
  65. Sacramento V.D.M., Santos K.T., Rocha D.F.D.O., Cabral E.C., Eberlin M.N., Mercadante-Simões M.O., Fonseca F.S.a.D., Melo Junior A.F., Menezes E.V., and Oliveira D.a.D., Chemical profile and antioxidant activity in *Diplopteryx pubipetala* (Malpighiaceae). *Natural Product Research*, **1-5** (2020). Doi: 10.1080/14786419.2020.1855644.
  66. Sofidiya M.O. and Familoni O., Antioxidant activities of different solvent extracts of leaves and root of *Flabellaria paniculata* Cav. (Malpighiaceae). *UNILAG Research Repository*, **6**(31), 4682-4690 (2012). Doi: 10.5897/JMPR12.395.
  67. Gupta R. and Jeevaratnam K., A Comparative Evaluation of *In Vitro* Phytochemical Analysis, Antioxidant, Antibacterial and Anticancer Activity of Methanolic (MeOH) Crude Extract of Bark (GGB) and Leaf (GGL) of *Galphimia glauca*, **6**(6), (2019).
  68. Fekry A., Elsabbagh W., Abu Bakr M., El-Ghazaly M., and Mohamed A.E.-S., Antioxidant activity of *Malpighia glabra* L., leaves extract. *Azhar International Journal of Pharmaceutical Medical Sciences*, **1**(2), 88-93 (2021). Doi: 10.21608/aijpm.2021.59935.1042
  69. Xu M., Shen C., Zheng H., Xu Y., Xue C., Zhu B., and Hu J., Metabolomic analysis of acerola cherry (*Malpighia emarginata*) fruit during ripening development via UPLC-Q-TOF and contribution to the antioxidant activity. *Food Research International*, **130**, 108915 (2020). Doi: 10.1016/j.foodres.2019.108915.
  70. Perez-Gutierrez R.M., Muñoz-Ramirez A., Gomez Y.G., and Ramirez E.B., Antihyperglycemic, antihyperlipidemic and antiglycation effects of *Byrsonima crassifolia* fruit and seed in normal and streptozotocin-induced diabetic rats. *Plant foods for human nutrition*, **65**(4), 350-357 (2010). Doi: 10.1007/s11130-010-0181-5.
  71. Gutiérrez R.M.P. and Ramirez A.M., Hypoglycemic Effects of sesquiterpene lactones from *Byrsonima crassifolia*. *Food science and biotechnology*, **25**(4),

- 1135-1145 (2016). Doi: 10.1007/s10068-016-0182-8.
72. Maheshwari P., Baburao B., and Ch P.K., Antidiabetic activity of methanolic extract of *Hiptage benghalensis* leaves in alloxan induced diabetic models. *Pakistan journal of biological sciences: PJBS*, **16**(17), 844-851 (2013). Doi: 10.3923/pjbs.2013.844.851.
  73. Nadh A.R.S., Rao P.R., and Rani A.P., Antidiabetic activity of *Hiptage Benghalensis* in chemical-induced diabetic rats. *International Journal of Advanced Research Ideas and Innovations in Technology (IJARIIT)*, **4**(2), 1092-1098 (2018).
  74. Lozano C.M., Vasquez-Tineo M.A., Ramirez M., and Jimenez F., *In vitro* antimicrobial activity screening of tropical medicinal plants used in Santo Domingo, Dominican Republic. Part I. *Pharmacognosy Communications*, **3**(2), 64-69 (2013). Doi: 10.5530/pc.2013.2.13.
  75. Sannomiya M., Michelin D., Rodrigues C., Santos L.C.D., Salgado H., Hiruma-Lima C., Brito A., and Vilegas W., *Byrsonima crassa* Niedenzu (IK): antimicrobial activity and chemical study. *Revista de Ciências Farmacêuticas Básica e Aplicada*, **26**(1), 71-75 (2005b).
  76. Abo K. and Olugbuyiro J., Phytochemical and antibacterial studies of extracts of *Flabellaria paniculata*. *African Journal of Biomedical Research*, **7**(1), 35-36 (2004). Doi: 10.4314/ajbr.v7i1.54064.
  77. Lalnundanga L.N. and Thanzami K., Antimicrobial Activity of Methanol Extract of Root Bark of *Hiptage benghalensis* (L) Kurz. *Journal of Pharmacognosy and Phytochemistry*, **3**(6), 119-121 (2015).
  78. Júnior W.a.R., Cardoso M.L.C., Vilegas W., Nakamura C.V., Dias Filho B.P., and De Mello J.C.P., A new antimicrobial from the roots of *Heteropteris aphrodisiaca*. *Acta Farm Bonaerense*, **24**(4), 543-5 (2005).
  79. Melo F.L., Benati F.J., Junior W.a.R., De Mello J.C.P., Nozawa C., and Linhares R.E.C., The *in vitro* antiviral activity of an aliphatic nitro compound from *Heteropteris aphrodisiaca*. *Microbiological Research*, **163**(2), 136-139 (2008). Doi: 10.1016/j.micres.2006.03.011.
  80. Huerta-Reyes M., Sánchez-Vargas L.O., Villanueva-Amador G.S., Gaitán-Cepeda L.A., and Health P., Anti-HIV and Anti-Candidal Effects of Methanolic Extract from *Heteropteris brachiata*. *International Journal of Environmental Research*, **18**(14), 7270 (2021). Doi: 10.3390/ijerph18147270
  81. Hussein A.A., Gomez B., Ramos M., Heller M., Coley P.D., Solis P.N., and Gupta M.P., Constituents of *Hiraea reclinata* and their anti-HIV activity *Revista Latinoamericana de Química*, **1**, 5-8 (2003).
  82. El-Hawary S.S., Mousa O.M., El-Fitany R.A., and El Gedaily R.A., Cytotoxic, antimicrobial activities, and phytochemical investigation of three peach cultivars and acerola leaves. *Journal of Reports in Pharmaceutical Sciences*, **9**(2), 221 (2020). Doi: 10.4103/jrptps.JRPTPS\_88\_19.
  83. Pires F.C.S., Oliveira J.C.D., Menezes E.G.O., Ferreira M.C.R., Siqueira L.M.M., Almada-Vilhena A.O., Pieczarka J.C., Nagamachi C.Y., and Carvalho Junior R.N.D., Bioactive Compounds and Evaluation of Antioxidant, Cytotoxic and Cytoprotective Effects of Murici Pulp Extracts (*Byrsonima crassifolia*) Obtained by Supercritical Extraction in HepG2 Cells Treated with H<sub>2</sub>O<sub>2</sub>. *Foods*, **10**(4), 737 (2021). Doi: 10.3390/foods10040737
  84. Verdam M.C.D.S., Guilhon-Simplicio F., Paula C.D.S., De Oliveira V.B., Miguel M.D., Campelo P.M.S., and Miguel O.G., Cytotoxicity of *Byrsonima duckeana* WR Anderson (malpighiaceae) on colon cancer cells. *International Journal of Pharmacy and Pharmaceutical Sciences*, **6**(11), 5719-23 (2013).
  85. Bhukya B.R. and Yellu N.R., Evaluation of anticancer activity of methanolic extract of *Hiptage benghalensis* (L.) Kurz on Cancer Cell Lines. *Pharmacognosy Research*, **10**(3), (2018). Doi: 10.4103/pr.pr\_102\_17.
  86. González-Cortazar M., Herrera-Ruiz M., Zamilpa A., Jiménez-Ferrer E., Marquina S., Álvarez L., and Tortoriello J., Anti-inflammatory activity and chemical profile of *Galphimia glauca*. *Planta medica*, **80**(01), 90-96 (2014). Doi: 10.1055/s-0033-1360150.
  87. Hridi S.U., Ferdous N., Majumder F.U., and Hannan J.A., Phytochemical Screening and Investigation of the Central and Peripheral Analgesic and Anti-Inflammatory Activity of Ethanol Extract of *Hiptage benghalensis* (L) Kurz. *Journal of Pharmaceutical Research International*, **3**(4), 1045-1057 (2013b). Doi: 10.9734/BJPR/2013/4454
  88. Herrera-Ruiz M., Zamilpa A., González-Cortazar M., Reyes-Chilpa R., León E., García M., Tortoriello J., and Huerta-Reyes M., Antidepressant effect and pharmacological evaluation of standardized extract of flavonoids from *Byrsonima crassifolia*. *Phytomedicine*, **18**(14), 1255-1261 (2011). Doi: 10.1016/j.phymed.2011.06.018.
  89. Herrera-Ruiz M., Jiménez-Ferrer J., De Lima T., Avilés-Montes D., Pérez-García D., González-Cortazar M., and Tortoriello J., Anxiolytic and antidepressant-like activity of a standardized extract from *Galphimia glauca*, *Phytomedicine*. **13**(1-2), 23-28 (2006). Doi: 10.1016/j.phymed.2005.03.003.
  90. Galiotta G., Giuliani G., Loizzo A., Amat A.G., Fumagalli E., De Feo V., Quaranta E., Paladino L., and Capasso A., Neurophysiological studies of *Heteropteris glabra* Hok. & Arn.(Malpighiaceae) in DBA/2J mice. *Journal of Ethnopharmacology*, **97**(3), 415-419 (2005). Doi: 10.1016/j.jep.2004.12.003.
  91. Huerta-Reyes M., Zamilpa A., Álvarez-Chimal R., Luna-Manzanares J.Á., León-Velasco M.E., Aguilar-Rojas A., Jiménez-Estrada M., and Campos-Lara M.G., *Heteropteris cotinifolia*: A neuropharmacological and phytochemical approach with possible taxonomic implications. *The Scientific World Journal*, **2013**, (2013b). Doi: 0.1155/2013/870468.
  92. Maheshwari P., Baburao B., Reddy A.R.N., and Methods, Hepatoprotective activity of methanolic extract of *Hiptage benghalensis* leaves against CCl<sub>4</sub>-induced hepatotoxicity in rats. *Toxicology*

- mechanisms*, **22**(6), 483-487 (2012). Doi: 10.3109/15376516.2012.674068.
93. El- Hawary S.S., El- Fitianny R.A., Mousa O.M., Salama A.A., and El Gedaily R.A., Metabolic profiling and *in vivo* hepatoprotective activity of *Malpighia glabra* L. leaves. *Journal of Food Biochemistry*, **45**(2), e13588 (2021). Doi: 10.1111/jfbc.13588.
  94. Lima Z.P., Dos Santos R.D.C., Torres T.U., Sannomiya M., Rodrigues C.M., Dos Santos L.C., Pellizzon C.H., Rocha L.R.M., Vilegas W., and Brito A.R.M.S., *Byrsonima fagifolia*: an integrative study to validate the gastroprotective, healing, antidiarrheal, antimicrobial and mutagenic action. *Journal of ethnopharmacology*, **120**(2), 149-160 (2008). Doi: 10.1016/j.jep.2008.07.047.
  95. Santos R.C., Kushima H., Rodrigues C.M., Sannomiya M., Rocha L.R.M., Bauab T.M., Tamashiro J., Vilegas W., and Hiruma-Lima C.A., *Byrsonima intermedia* A. Juss.: gastric and duodenal anti-ulcer, antimicrobial and antidiarrheal effects in experimental rodent models. *Journal of Ethnopharmacology*, **140**(2), 203-212 (2012). Doi: 10.1016/j.jep.2011.12.008.
  96. Dos Santos R.D.C., Bonamin F., Périco L.L., Rodrigues V.P., Zanatta A.C., Rodrigues C.M., Sannomiya M., Dos Santos Ramos M.A., Bonifácio B.V., and Bauab T.M., *Byrsonima intermedia* A. Juss partitions promote gastroprotection against peptic ulcers and improve healing through antioxidant and anti-inflammatory activities. *Biomedicine & Pharmacotherapy*, **111**, 1112-1123 (2019). Doi: 10.1016/j.biopha.2018.12.132.
  97. Galvão S., Marques L., Oliveira M., and Carlini E., *Heteropterys aphrodisiaca* (extract BST0298): a Brazilian plant that improves memory in aged rats. *Journal of Ethnopharmacology*, **79**(3), 305-311 (2002). Doi: 10.1016/S0378-8741(01)00402-0.
  98. De Sousa L.R.F., Ramalho S.D., Burger M.C.D.M., Nebo L., Fernandes J.B., Da Silva M.F.T.D.G.a.F., Iemma M.N.R.D.C., Correa C.J., Souza D.H.F.D., and Lima M.I.S.S., Isolation of arginase inhibitors from the bioactivity-guided fractionation of *Byrsonima coccolobifolia* leaves and stems. *Journal of natural products*, **77**(2), 392-396 (2014). Doi: 10.1021/np400717m.
  99. Del Rayo Camacho M., Phillipson J.D., Croft S.L., Marley D., Kirby G.C., and Warhurst D.C., Assessment of the Antiprotozoal Activity of *Galphimia glauca* and the Isolation of New Norsecofriedelanes and Nor-friedelanes. *Journal of natural products*, **65**(10), 1457-1461 (2002). Doi: 10.1021/np010419i.
  100. Retnasamy G. and Adikay S., Effect of *Hiptage madablota* Gaertn. on High Fat Diet--Induced Obese Rats. *Jordan Journal of Biological Sciences*, **7**(2), 113-118 (2014).
  101. Abo A., Olugbuyiro J., and Famakinde S., Anti-infective and wound healing properties of *Flabellaria paniculata*. *African Journal of Biomedical Research*, **7**(2), (2004). Doi: 10.4314/ajbr.v7i2.54075
  102. Olugbuyiro J.A., Abo K., and Leigh O., Wound healing effect of *Flabellaria paniculata* leaf extracts. *Journal of ethnopharmacology*, **127**(3), 786-788 (2010). Doi: 10.1016/j.jep.2009.10.008.
  103. Lotfy M., Ksiksi T.S., Palakkot A.R., D'souza C.M., Mohsin S., and Adegate E.A., Anti-diabetic Effect of *Acridocarpus Orientalis*. *The Open Medicinal Chemistry Journal*, **14**(1), 132-144 (2020b). Doi: 10.2174/1874104502014010132.
  104. Sun P., Cao D.-H., Xiao Y.-D., Zhang Z.-Y., Wang J.-N., Shi X.-C., Xiao C.-F., Hu H.-B., and Xu Y.-K., Aspidopteroids A–D: Four New Diterpenoids from *Aspidopterys obcordata* Vine. *Chemistry*, **25**(3), 529 (2020). Doi: 10.3390/molecules25030529.
  105. Oliveira D.M., Silva T.F., Martins M.M., De Moraes S.A., Chang R., De Aquino F.J., Da Silva C.V., Teixeira T.L., Martins C.H., and Moraes T.S., Antifungal and cytotoxicity activities of *Banisteriopsis argyrophylla* leaves. *Journal of Pharmacy Pharmacology*, **70**(11), 1541-1552 (2018). Doi: 10.1111/jphp.12996.
  106. O'connell F., Isolation of caffeine from *Banisteriopsis inebrians* (Malpighiaceae). *Naturwissenschaften*, **56**(3), 139-140 (1969).
  107. Hashimoto Y. and Kawanishi K., New alkaloids from *Banisteriopsis caapi*. *Phytochemistry*, **15**(10), 1559-1560 (1976). Doi: 10.1016/S0031-9422(00)88936-0.
  108. Santos B.W.L., Oliveira R.C.D., Sonsin-Oliveira J., Fagg C.W., Barbosa J.B.F., and Caldas E.D., Biodiversity of  $\beta$ -Carboline Profile of *Banisteriopsis caapi* and Ayahuasca, a Plant and a Brew with Neuropharmacological Potential. *Plants*, **9**(7), 870 (2020). Doi: 10.3390/plants9070870.
  109. Coe F.G. and Anderson G.J., Snakebite ethnopharmacopoeia of eastern Nicaragua. *Journal of ethnopharmacology*, **96**(1-2), 303-323 (2005). Doi: 10.1016/j.jep.2004.09.026.
  110. Karunasena G., Chandrajith V., and Navaratne S., Antioxidant capacity and total phenol content of peanut butter fruit (*Bunchosia armenica*). *Journal of Pharmacognosy and Phytochemistry*, **7**(4), 343-346 (2018a).
  111. Croda M.F., Carvalho D., Fraga S., Espindola J.D.S., and Fernandes De Moura N., Compostos bioativos em suco misto de *Euterpes edulis* e *Bunchosia glandulifera*. *Brazilian Journal of Food Technology*, **20**, e2016147 (2017). Doi: 10.1590/1981-6723.14716.
  112. Blank D.E., Justen D., Fraga S., Peixoto C.R., and De Moura N.F., Chemical Composition and Antioxidant Activity of *Bunchosia glandulifera* Fruit at Different Ripening Stages. *Food and Nutrition Sciences*, **9**(10), 1147-1159 (2018b). Doi: 10.4236/fns.2018.910083
  113. Blank D.E., Bellaver M., Fraga S., Lopes T.J., and De Moura N.F., Drying kinetics and bioactive compounds of *Bunchosia glandulifera*. *Journal of Food Process Engineering*, **41**(4), e12676 (2018a). Doi: 10.1111/jfpe.12676.
  114. Silva S.D.F., Blank D.E., Peixoto C.R., De Jesus Da Silveira Moreira J., and Fernandes De Moura N., Bioactive compounds and antioxidant activity of

- Bunchosia glandulifera*. *International Journal of Food Properties*, **19**(2), 467-473 (2016). Doi: 10.1080/10942912.2015.1033547.
115. Agra M.D.F., Silva K.N., Basílio I.J.L.D., Freitas P.F.D., and Barbosa-Filho J.M., Survey of medicinal plants used in the region Northeast of Brazil. *Revista brasileira de farmacognosia*, **18**(3), 472-508 (2008).
  116. De Araújo Rodrigues P., De Moraes S.M., Aguiar L.A., Vila-Nova N.S., and Benjamin S.R., Effect of *Byrsonima sericea* DC. leaf extracts on mice gastrointestinal tract. *Toxicology reports*, **6**, 1182-1187 (2019). Doi: 10.1016/j.toxrep.2019.10.018.
  117. Arantes V., Sato D., Vilegas W., Santos L., and Leite C.Q.F., Plantas do cerrado brasileiro com atividade contra *Mycobacterium fortuitum*. *Revista de Ciências Farmacêuticas Básica e Aplicada*, **26**(3), 195-198 (2005).
  118. Sannomiya M., Fonseca V.B., Da Silva M., Rocha L., Dos Santos L., Hiruma-Lima C., Brito A.S., and Vilegas W., Flavonoids and antiulcerogenic activity from *Byrsonima crassa* leaves extracts. *Journal of Ethnopharmacology*, **97**(1), 1-6 (2005a). Doi: 10.1016/j.jep.2004.09.053.
  119. Cardoso C.R.P., De Syllos Cólus I.M., Bernardi C.C., Sannomiya M., Vilegas W., and Varanda E.A., Mutagenic activity promoted by amentoflavone and methanolic extract of *Byrsonima crassa* Niedenzu. *Toxicology*, **225**(1), 55-63 (2006). Doi: 10.1016/j.tox.2006.05.003.
  120. Higuchi C.T., Pavan F.R., Leite C.Q.F., Sannomiya M., Vilegas W., Leite S.R.D.A., Sacramento L.V.S., and Sato D.N., Triterpenes and antitubercular activity of *Byrsonima crassa*. *Química nova*, **31**(7), 1719-1721 (2008). Doi: 10.1590/S0100-40422008000700023
  121. Bonacorsi C., Raddi M.S.G., Carlos I.Z., Sannomiya M., and Vilegas W., Anti-*Helicobacter pylori* activity and immunostimulatory effect of extracts from *Byrsonima crassa* Nied.(Malpighiaceae). *BMC Complementary and Alternative Medicine*, **9**(1), 1-7 (2009). Doi: 10.1186/1472-6882-9-2.
  122. Gutiérrez R.M.P. and Ramirez A.M., Hexane extract of the seeds of *Byrsonima crassifolia* accelerates wound healing in streptozotocin-induced diabetic rats. *Chinese journal of integrative medicine*, 1-7 (2013). Doi: 10.1007/s11655-013-1556-x.
  123. Muniz-Ramirez A., Perez-Gutierrez R.M., Garcia-Baez E., and Mota-Flores J.M., Antimicrobial activities of diterpene labdane from seeds of *Byrsonima crassifolia*. *Boletín Latinoamericano y del Caribe de Plantas Medicinales y Aromáticas*, **13**(1), 31-37 (2014).
  124. Andrade B., Matias R., Corrêa B., Oliveira A., Guidolin D., and Roel A., Phytochemistry, antioxidant potential and antifungal of *Byrsonima crassifolia* on soil phytopathogen control. *Brazilian Journal of Biology*, **78**(1), 140-146 (2017). Doi: 10.1590/1519-6984.166532.
  125. Irias-Mata A., Jiménez V.M., Steingass C.B., Schweiggert R.M., Carle R., and Esquivel P., Carotenoids and xanthophyll esters of yellow and red nance fruits (*Byrsonima crassifolia* (L.) Kunth) from Costa Rica. *Food Research International*, **111**, 708-714 (2018). Doi: 10.1016/j.foodres.2018.05.063.
  126. Sobrinho A.C.G., Rogez H.L.G., Do Nascimento V.H.A., Teixeira B.J.B., De Sousa Dias A.L., and De Souza J.N.S., Determinação de compostos bioativos e capacidade sequestradora de radicais livres em extratos de folhas de *Byrsonima crassifolia* e *Inga edulis*. *Brazilian Journal of Development*, **6**(6), 34954-34969 (2020). Doi: 10.34117/bjdv6n6-147.
  127. Do Nascimento V.H.A., Sobrinho A.C.G., De Oliveira Souza C., De Souza J.N.S., and Sousa C.L., Determination of Phenolic Compounds with Antimicrobial Activity of *Byrsonima Crassifolia* and *Inga Edulis* Leaves Extracts. *Ensaio e Ciência C Biológicas Agrárias e da Saúde*, **25**(1), 21-28 (2021). Doi: 10.17921/1415-6938.2021v25n1p21-28.
  128. Sannomiya M., Cardoso C.R., Figueiredo M.E., Rodrigues C.M., Dos Santos L.C., Dos Santos F.V., Serpeloni J.M., Cólus I.M., Vilegas W., and Varanda E.A., Mutagenic evaluation and chemical investigation of *Byrsonima intermedia* A. Juss. leaf extracts. *Journal of ethnopharmacology*, **112**(2), 319-326 (2007). Doi: 10.1016/j.jep.2007.03.014.
  129. Orlandi L., Vilela F.C., Santa-Cecília F.V., Dias D.F., Alves-Da-Silva G., and Giusti-Paiva A., Anti-inflammatory and antinociceptive effects of the stem bark of *Byrsonima intermedia* A. Juss. *Journal of Ethnopharmacology*, **137**(3), 1469-1476 (2011). Doi: 10.1016/j.jep.2011.08.032.
  130. Moreira L.Q., Vilela F.C., Orlandi L., Dias D.F., Santos A.L.A., Da Silva M.A., Paiva R., Alves-Da-Silva G., and Giusti-Paiva A., Anti-inflammatory effect of extract and fractions from the leaves of *Byrsonima intermedia* A. Juss. in rats. *Journal of ethnopharmacology*, **138**(2), 610-615 (2011). Doi: 10.1016/j.jep.2011.10.006.
  131. Zanatta A.C., Vilegas W., and Edrada-Ebel R., UHPLC-(ESI)-HRMS and NMR-based metabolomics approach to access the seasonality of *Byrsonima intermedia* and *Serjania marginata* from Brazilian Cerrado flora diversity. *Frontiers in chemistry*, **9**, 534 (2021). Doi: 10.3389/fchem.2021.710025.
  132. Saldanha A.A., De Siqueira J.M., Castro A.H.F., De Azambuja Ribeiro R.I.M., De Oliveira F.M., De Oliveira Lopes D., Pinto F.C.H., Silva D.B., and Soares A.C., Anti-inflammatory effects of the butanolic fraction of *Byrsonima verbascifolia* leaves: Mechanisms involving inhibition of tumor necrosis factor alpha, prostaglandin E2 production and migration of polymorphonuclear leucocyte *in vivo* experimentation. *International immunopharmacology*, **31**, 123-131 (2016a). Doi: 10.1016/j.intimp.2015.12.031.
  133. Saldanha A.A., Do Carmo L.F., Do Nascimento S.B., De Matos N.A., De Carvalho Veloso C., Castro A.H.F., De Vos R.C., Klein A., De Siqueira J.M., and Carollo C.A., Chemical composition and anti-inflammatory activity of the leaves of *Byrsonima verbascifolia*. *Journal of natural medicines*, **70**(4), 760-768 (2016b). Doi: 10.1007/s11418-016-1011-3.

134. De Barros G.L.R.R., Sanches M.a.R., Barcia M.T., Rodrigues D., and Pertuzatti P.B., Murici (*Byrsonima verbascifolia*): A high bioactive potential fruit for application in cereal bars. *LWT - Food Science and Technology*, **160**, 113279 (2022). Doi: 10.1016/j.lwt.2022.113279.
135. Mendes C.C., Cruz F.G., David J.M., Nascimento I.P., and David J.P., Triterpenes esterified with fatty acid and triterpene acids isolated from *Byrsonima microphylla*. *Química Nova*, **22**(2), 185-188 (1999). Doi: 10.1590/S0100-40421999000200007
136. De Souza T.P., De Almeida P.D.O., Alex P., Ohana D.T., Lima E.S., and De Meneses Pereira M., Antioxidant activity of a standardized extract of *Byrsonima japurensis* A. Juss.(Malpighiaceae) stem bark. *Journal of Medicinal Plants Research*, **7**(26), 1926-1930 (2013). Doi: 10.5897/JMPR12.1133.
137. De Siqueira-Jaccoud R., *Contribuição para o estudo farmacognóstico do Cabi Paraensis Ducke: I.*(1959).
138. Shenkute B., Hassen A., Assafa T., Amen N., and Ebro A., Identification and nutritive vale of potential fodder trees and shrubs in the mid Rift Valley of Ethiopa. *Journal of Animal & Plant Sciences*, **22**(4), 1126-1132 (2012).
139. Motta L.B., Furlan C.M., Salatino A., and Salatino M.L., Flavonoids and the taxonomy of *Camarea* (Malpighiaceae). *Biochemical Systematics and Ecology*, **37**(3), 201-205 (2009). Doi: 10.1016/j.bse.2009.03.005.
140. De on Costa M., Santos K.T., Almeida C.A., Da Fonseca F.S., Angolini C.F., De Oliveira D.A., De Melo Júnior A.F., Menezes E.V., and Royo V.D.A., Chemical Composition of *Diplopterys pubipetala* (Malpighiaceae). *European Journal of Medicinal Plants*, **31**(15), 43-48 (2020). Doi: 10.9734/EJMP/2020/v31i1530323.
141. Orozco-Martínez J., Lira-Saade R., Jiménez-Estrada M., Ávila-Acevedo J.G., Serrano-Parrales R., and Hernández-Delgado T., Medicinal plants of Oaxaca, Mexico: ethnobotany and antibacterial activity. *Bol. latinoam. Caribe plantas med. aromát.*, **19**(2), 221-235 (2020). Doi: 10.37360/blacpma.20.19.2.14.
142. Fayemi Scott O. and Osho A., Comparison of antimicrobial effects of *Mezoneuron benthamianum*, *Heliotropium indicum* and *Flabellaria paniculata* on *Candida* species. *Journal of Microbiology Research*, **2**(1), 18-23 (2012). Doi: 10.5923/j.microbiology.20120201.04.
143. Sofidiya M.O., Agufobi L., Akindele A.J., Olowe J.A., and Familoni O.B., Effect of *Flabellaria paniculata* Cav. extracts on gastric ulcer in rats. *BMC complementary and alternative medicine*, **12**(1), 1-6 (2012).
144. Akindele A.J., Adeneye A.A., Salau O.S., Sofidiya M.O., and Benebo A.S., Dose and time-dependent sub-chronic toxicity study of hydroethanolic leaf extract of *Flabellaria paniculata* Cav.(Malpighiaceae) in rodents. *Frontiers in pharmacology*, **5**, 78 (2014). Doi: 10.3389/fphar.2014.00078.
145. Dorsch W., Bittinger M., Kaas A., Müller A., Kreher B., and Wagner H., Antiasthmatic effects of *Galphimia glauca*, gallic acid, and related compounds prevent allergen-and platelet-activating factor-induced bronchial obstruction as well as bronchial hyperreactivity in guinea pigs. *International archives of allergy and immunology*, **97**(1), 1-7 (1992). Doi: 10.1159/000236088.
146. Tortoriello J. and Ortega A., Sedative effect of galphimine B, a nor-seco-triterpenoid from *Galphimia glauca*. *Planta medica*, **59**(5), 398-400 (1993). Doi: 10.1055/s-2006-959717.
147. Garige B.S.R., Keshetti S., and Vattikuti U.M.R., CNS Depressant effects and muscle relaxant activity of *Galphimia glauca* leaf methanol extract. *International Journal of PharmTech Research*, **9**(6), 230-240 (2016).
148. Chordiya S., Pimprikar R., Yeshwante S., Tanvir S., Patil P., Kale M., and Firke B., Anthelmintic Activity of *Hiptage benghalensis* (L) Kurz Leaves. *Research Journal of Pharmacognosy and Phytochemistry*, **1**(3), 234-235 (2009).
149. Kumudhavalli M., Jayakar B., Chandira R.M., Kumar M., and Saravanan C., Phytochemical and Pharmacological studies on leaves of *Hiptage bengalensis* (L) Kurz. *International Journal of Pharm Tech Research*, **2**(1), 1017-1020 (2010).
150. Amudha P. and Shanthi P., Antioxidant activity of some rare medicinal plants. *Journal of Pharmacy Research*, **4**(3), 698-699 (2011).
151. Murugan M. and Mohan V., Evaluation of phytochemical analysis and antibacterial activity of *Bauhinia purpurea* L. and *Hiptage benghalensis* L. Kurz. *Journal of Applied Pharmaceutical Science*, **1**(9), 157 (2011).
152. Ngente L., Nachimuthu S.K., and Guruswami G., Insecticidal and repellent activity of *Hiptage benghalensis* L. Kruz (Malpighiaceae) against mosquito vectors. *Parasitology research*, **111**(3), 1007-1017 (2012). Doi: 10.1007/s00436-012-2925-7.
153. Yadav S. and Kumar P., Production, isolation and identification of flavonoids from aerial parts of *Hiptage benghalensis*. *International Journal of Life Science and Pharma Research*, **2**(3), 1-5 (2012).
154. Hridi S.U., Ferdous N., Majumder F.U., and Hannan J., Phytochemical screening and anti-diabetic efficacy of stem of *Hiptage benghalensis* (L) Kurz. *Journal of Scientific and Innovative Research*, **2**(4), 736-744 (2013a).
155. Hsu C.-L., Fang S.-C., Huang H.-W., and Yen G.-C., Anti-inflammatory effects of triterpenes and steroid compounds isolated from the stem bark of *Hiptage benghalensis*. *Journal of functional foods*, **12**, 420-427 (2015). Doi: 10.1016/j.jff.2014.12.009.
156. Huerta-Reyes M., Herrera-Ruiz M., Gonzalez-Cortazar M., Zamilpa A., Leon E., Reyes-Chilpa R., Aguilar-Rojas A., and Tortoriello J., Neuropharmacological *in vivo* effects and phytochemical profile of the extract from the aerial parts of *Heteropterys brachiata* (L.) DC.(Malpighiaceae). *Journal of Ethnopharmacology*, **146**(1), 311-317 (2013a). Doi: 10.1016/j.jep.2012.12.049.

157. Júnior H.M.S., Campos V.A., Alves D.S., Cavalheiro A.J., Souza L.P., Botelho D.M., Chalfoun S.M., and Oliveira D.F., Antifungal activity of flavonoids from *Heteropterys byrsonimifolia* and a commercial source against *Aspergillus ochraceus*: *In silico* interactions of these compounds with a protein kinase. *Crop Protection*, **62**, 107-114 (2014). Doi: 10.1016/j.cropro.2014.04.012.
158. Bobach C., Schurwanz J., Franke K., Denkert A., Van Sung T., Kuster R., Mutiso P.C., Seliger B., and Wessjohann L.A., Multiple readout assay for hormonal (androgenic and antiandrogenic) and cytotoxic activity of plant and fungal extracts based on differential prostate cancer cell line behavior. *Journal of ethnopharmacology*, **155**(1), 721-730 (2014). Doi: 10.1016/j.jep.2014.06.008.
159. Stermitz F.R., Hnatsyzyn O., Bandoni A.L., Rondina R.V., and Coussio J.D., Screening of Argentine plants for aliphatic nitro compounds: hiptagin from *Heteropterys angustifolia*. *Phytochemistry*, **14**(5-6), 1341-1345 (1975). Doi: 10.1016/S0031-9422(00)98622-9.
160. Marques L.C., Pieri C.D., Roman-Júnior W.A., Cardoso M.L., Milaneze-Gutierrez M.A., and Mello J.C., Pharmacognostic analysis of the roots of *Heteropterys aphrodisiaca* O. Mach.(Malpighiaceae). *Revista Brasileira de Farmacognosia*, **17**(4), 604-615 (2007). Doi: 10.1590/S0102-695X2007000400021
161. Monteiro J.C., Predes F.S., Matta S.L., and Dolder H., *Heteropterys aphrodisiaca* infusion reduces the collateral effects of cyclosporine A on the testis. *The Anatomical Record: Advances in Integrative Anatomy and Evolutionary Biology*, **291**(7), 809-817 (2008). Doi: 10.1002/ar.20709.
162. Gomes M.L., Monteiro J.C., Freitas K.M., Sbervelheri M.M., and Dolder H., Association of the infusion of *Heteropterys aphrodisiaca* and endurance training brings spermatogenic advantages. *Biological Research*, **44**(3), 235-241 (2011). Doi: 10.4067/S0716-97602011000300004.
163. Monteiro J.C., Gomes M.L., Tomiosso T.C., Nakagaki W.R., Sbervelheri M.M., Ferrucci D.L., Pimentel E.R., and Dolder H., More resistant tendons obtained from the association of *Heteropterys aphrodisiaca* and endurance training. *BMC complementary and alternative medicine*, **11**(1), 1-11 (2011).
164. Galvão S.M.P., Mendes F.R., Oliveira M.G.M.D., Mattei R., Mello J.C.P.D., Roman Júnior W.A., and Carlini E.D.A., Memory retrieval improvement by *Heteropterys aphrodisiaca* in aging rats. *Brazilian Journal of Pharmaceutical Sciences*, **47**(4), 825-832 (2011). Doi: 10.1590/S1984-82502011000400020
165. Freitas K.M., Monteiro J.C., Gomes M.L., Taboga S.R., and Dolder H., *Heteropterys tomentosa* (A. Juss.) infusion counteracts Cyclosporin a side effects on the ventral prostate. *BMC complementary and alternative medicine*, **13**(1), 1-9 (2013).
166. Paula-Freire L.I., Mendes F.R., Molska G.R., Duarte-Almeida J.M., and Carlini E.A., Comparison of the chemical composition and biological effects of the roots, branches and leaves of *Heteropterys tomentosa* A. Juss. *Journal of ethnopharmacology*, **145**(2), 647-652 (2013). Doi: 10.1016/j.jep.2012.12.004.
167. Bezerra A.G., Negri G., Duarte-Almeida J.M., Smaili S.S., and Carlini E.A., Phytochemical analysis of hydroethanolic extracts from powdered roots of *Panax ginseng* C. A. Meyer and *Heteropterys tomentosa* A. Juss and evaluation of their effects on astrocyte cell death. *Química Nova*, **39**(5), 581-587 (2016). Doi: 10.5935/0100-4042.20160069.
168. Pirovani J.C.M., Gomes M.L., and Fernando O., *Heteropterys tomentosa* Improves the Endurance Capacity of Skeletal Muscles in Trained Rats. *IOSR Journal of Pharmacy and Biological Sciences*, **11**(4), 39-45 (2016). Doi: 10.9790/3008-1104033945.
169. Farias D.M., Ostetto M.S., Colleti R., Barbosa T., Barros J.A., Machado S., Murillo-Rodríguez E., Moreno S.E., and Veras A.B., Effect of *Heteropterys aphrodisiaca* on Anxiety and Male Exposure of Female Mice with Advanced Age. *Current clinical pharmacology*, **12**(2), 106-112 (2017). Doi: 10.2174/1574884712666170622085129.
170. Fraga G.A., Balogun S.O., Pascqua E.D., De Oliveira R.G., Botelho G., Pavan E., Da Rosa Lima T., Avila E.T.P., De Medeiros Amorim Krueger C., and Filho V.C., *Heteropterys tomentosa* A. Juss: toxicological and adaptogenic effects in experimental models. *Nutrition health*, **23**(4), 289-298 (2017). Doi: 10.1177/0260106017729908.
171. De Rosso V. and Mercadante A., Carotenoid composition of two Brazilian genotypes of acerola (*Malpighia puniceifolia* L.) from two harvests. *Food Research International*, **38**(8-9), 1073-1077 (2005). Doi: 10.1016/j.foodres.2005.02.023.
172. Da Silva Nunes R., Kahl V.F.S., Da Silva Sarmiento M., Richter M.F., Costa-Lotufo L.V., Rodrigues F.a.R., Abin-Carriquiry J.A., Martinez M.M., Ferronato S., and Ferraz A.D.B.F., Antigenotoxicity and antioxidant activity of Acerola fruit (*Malpighia glabra* L.) at two stages of ripeness. *Plant foods for human nutrition*, **66**(2), 129-135 (2011). Doi: 10.1007/s11130-011-0223-7.
173. Almeida I., Düsman E., Heck M., Pamphile J., Lopes N., Tonin L., and Vicentini V., Cytotoxic and mutagenic effects of iodine-131 and radioprotection of acerola (*Malpighia glabra* L.) and beta-carotene *in vitro*. *Genetics and Molecular Research*, **12**(4), 6402-6413 (2013). Doi: 10.4238/2013.December.10.1.
174. Tremonte P., Sorrentino E., Succi M., Tipaldi L., Pannella G., Ibanez E., Mendiola J.A., Di Renzo T., Reale A., and Coppola R., Antimicrobial effect of *Malpighia puniceifolia* and extension of water buffalo steak shelf- life. *Journal of food science*, **81**(1), M97-M105 (2016). Doi: 10.1111/1750-3841.13141.
175. Nascimento E.M., Rodrigues F.F., Costa W.D., Teixeira R.N., Boligon A.A., Sousa E.O., Rodrigues F.F., Coutinho H.D., Da Costa J.G.M.J.F., and Toxicology C., HPLC and *in vitro* evaluation of antioxidant properties of fruit from *Malpighia glabra* (Malpighiaceae) at different stages of

- maturation. *Food and Chemical Toxicology*, **119**, 457-463 (2018). Doi: 10.1016/j.fct.2017.11.042.
176. De Oliveira S.D., Araújo C.M., Borges G.D.S.C., Dos Santos Lima M., Viera V.B., Garcia E.F., De Souza E.L., and De Oliveira M.E.G., Improvement in physicochemical characteristics, bioactive compounds and antioxidant activity of acerola (*Malpighia emarginata* DC) and guava (*Psidium guajava* L.) fruit by-products fermented with potentially probiotic lactobacilli. *LWT-Food Science and Technology*, **134**, 110200 (2020). Doi: 10.1016/j.lwt.2020.110200.
177. Motohashi N., Wakabayashi H., Kurihara T., Fukushima H., Yamada T., Kawase M., Sohara Y., Tani S., Shirataki Y., and Sakagami H., Biological activity of barbados cherry (acerola fruits, fruit of *Malpighia emarginata* DC) extracts and fractions. *Phytotherapy Research*, **18**(3), 212-223 (2004). Doi: 10.1002/ptr.1426.
178. Righetto A., Netto F., and Carraro F., Chemical composition and antioxidant activity of juices from mature and immature acerola (*Malpighia emarginata* DC). *Food Science and Technology International*, **11**(4), 315-321 (2005). Doi: 10.1177/1082013205056785.
179. Hanamura T., Mayama C., Aoki H., Hirayama Y., and Shimizu M., Antihyperglycemic effect of polyphenols from Acerola (*Malpighia emarginata* DC.) fruit. *Bioscience, biotechnology, biochemistry*, **70**(8), 1813-1820 (2006). Doi: 10.1271/bbb.50592.
180. De Rosso V.V., Hillebrand S., Montilla E.C., Bobbio F.O., Winterhalter P., and Mercadante A.Z., Determination of anthocyanins from acerola (*Malpighia emarginata* DC.) and açai (*Euterpe oleracea* Mart.) by HPLC-PDA-MS/MS. *Journal of Food Composition and Analysis*, **21**(4), 291-299 (2008). Doi: 10.1016/j.jfca.2008.01.001.
181. Mezadri T., Villaño D., Fernández-Pachón M., García-Parrilla M., and Troncoso A., Antioxidant compounds and antioxidant activity in acerola (*Malpighia emarginata* DC.) fruits and derivatives. *Journal of Food Composition and Analysis*, **21**(4), 282-290 (2008). Doi: 10.1016/j.jfca.2008.02.002.
182. Delva L., Goodrich- Schneider R., and Technology, Antioxidant activity and antimicrobial properties of phenolic extracts from acerola (*Malpighia emarginata* DC) fruit. *International Journal of Food Science*, **48**(5), 1048-1056 (2013). Doi: 10.1111/ijfs.12061.
183. Leffa D.D., Rezin G.T., Daumann F., Longaretti L.M., Dajori A.L.F., Gomes L.M., Silva M.C., Streck E.L., and De Andrade V.M., Effects of acerola (*Malpighia emarginata* DC.) juice intake on brain energy metabolism of mice fed a cafeteria diet. *Molecular neurobiology*, **54**(2), 954-963 (2017). Doi: 10.1007/s12035-016-9691-y.
184. Barros B.R., Barboza B.R., Ramos B.A., Moura M.C., Coelho L.C., Napoleao T.H., Correia M.T.S., Paiva P., Maria G., and Cruz I.J.D., Saline extract from *Malpighia emarginata* DC leaves showed higher polyphenol presence, antioxidant and antifungal activity and promoted cell proliferation in mice splenocytes. *Anais da Academia Brasileira de Ciências*, **91**(1), e20180358 (2019). Doi: 10.1590/0001-3765201920180358.
185. Da Silva Barros B.R., Do Nascimento D.K.D., De Araújo D.R.C., Da Costa Batista F.R., De Oliveira Lima A.M.N., Da Cruz Filho I.J., De Oliveira M.L., and De Melo C.M.L., Phytochemical analysis, nutritional profile and immunostimulatory activity of aqueous extract from *Malpighia emarginata* DC leaves. *Biocatalysis Agricultural Biotechnology*, **23**, 101442 (2020). Doi: 10.1016/j.bcab.2019.101442.
186. Russo H.M., Queiroz E.F., Marcourt L., Rutz A., Allard P.-M., De Almeida R.F., Carvalho N.M., Wolfender J.-L., and Da Silva Bolzani V., Phytochemical analysis of the methanolic leaves extract of *Niedenzuella multiglandulosa* (Malpighiaceae), a plant species toxic to cattle in Brazil. *Phytochemistry Letters*, **37**, 10-16 (2020). Doi: 10.1016/j.phytol.2020.02.005.
187. Iyekowa O., Edema M., and Igbe I., Antimalarial Potential of Methanol Extract of *Stigmaphyllonovatum* in *Plasmodium falciparum* Infected Mice. *FUW Trends in Science & Technology Journal*, **6**(1), 239 – 242 (2021).
188. Nieto-Argüello A., Medina-Cruz D., Pérez-Ramírez Y.S., Pérez-García S.A., Velasco-Soto M.A., Jafari Z., De Leon I., González M.U., Huttel Y., and Martínez L., Composition-Dependent Cytotoxic and Antibacterial Activity of Biopolymer-Capped Ag/Au Bimetallic Nanoparticles against Melanoma and Multidrug-Resistant Pathogens. *Nanomaterials*, **12**(5), 779 (2022). Doi: 10.3390/nano12050779.
189. David J.M., Santos F.A., Guedes M.L.D.S., and David J.P., Flavonóide e triterpenos de *Stigmaphyllon paralias*. *Química Nova*, **26**(4), 484-487 (2003). Doi: 10.1590/S0100-40422003000400007
190. Mo A.M., A Chemical Study on the Constituents of *Stephania Rotunda* and *Tristellateia Australasiae*, *M. Thesis* (1996).
191. Randrianarivelosia M., Rasidimanana V.T., Rabarison H., Cheplogoi P.K., Ratsimbason M., Mulholland D.A., and Mauclère P., Plants traditionally prescribed to treat *tazo* (malaria) in the eastern region of Madagascar. *Malaria Journal*, **2**(1), 1-9 (2003).