



## Impact of Some Macro and Micro Nutrients on Productivity and Oil Properties of Coratina Olive Trees

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### Abstract

This experiment was conducted in two successive seasons (2018, 2019) on Coratina olive trees planted in a private orchard located at El Khatatbah district, El Behera governorate, Egypt; to study the effect of potassium dihydrogen phosphate ( $\text{KH}_2\text{PO}_4$ ), magnesium sulphate ( $\text{MgSO}_4$ ) and boric acid ( $\text{H}_3\text{BO}_3$ ) on olive yield per tree, fruit quality, also content and characteristics of fruit oil. The treatments were applied twice in September and November of each season. Generally, using any of the treatments under this investigation is led to increase the fruit yield per trees, and the fruit physical characteristics, also the oil accumulation percentage in the fruits, with maintaining its characteristics and quality as extra-virgin olive oil, compared to the untreated trees (control). However, treating the trees with magnesium sulphate alone recorded the highest fruit yield in the first season compared to the control. While, treating trees with potassium dihydrogen phosphate + boric acid + magnesium sulphate treatment, led to the highest oil percentage in the fruits, in the second season.

**Keywords:** Coratina; olive; fruit quality; olive oil; potassium dihydrogen phosphate; magnesium sulphate; boric acid.

### 1. Introduction

The worldwide olive oil consumption has increased up to 73% over the last 25 years which encourages the producers to increase its quality and quantity.

Low productivity of olive trees grown in sandy soil is a serious problem in Egypt, as evidenced by the new introduced varieties Koroneiki, Boutillan and Arbequina, which are related to water requirements, pruning and harvesting, planting density, and fertilization, particularly the method and doses of fertilizers [1]. The availability of nutrients is strongly influenced by soil properties. Clay soils, for example, have a high Potassium (K) fixing capacity and so show minimal reaction to K fertilizers added to the soil because much of the available K is quickly fixed to the clay particles [2]. If the mineral has been leached with excessive irrigation and/or rainfall in sandy soils, application has absolutely little impact [3].

Nutritional absorption is influenced by the availability of nutrients in the root system, as well as the nutrient need level and uptake period [4]. A high magnesium concentration in the soil inhibited plant calcium absorption, implying that plants grown in

such conditions may soon suffer from calcium deficiency [5]. Magnesium and calcium, as well as potassium and magnesium, have antagonistic relationships [6]. Excessive potassium application might have adverse or antagonistic effects on calcium and magnesium absorption due to potassium accumulation in the soil, and that the intensity of this impact on plants depended on plant variety and soil type [7]. It's possible that the negative interaction between potassium and magnesium consumption is due to their effects on leaves' sodium content [8].

Foliar application of fertilizers is generally beneficial in fulfilling plant needs and provides a quick way to restore nutrient deficits in plants. Furthermore, it is a convenient way to apply highly soluble fertilizers, especially in smaller doses; [9, 10].

Because of potassium, as a mineral osmosis, plays an important role in osmotic and pressure regulation, it is considered an essential element in higher plants. This element is crucial for cell expansion, plant development, and, lastly, the opening and shutting of stomata on leaves [11]. This element is also important for the activation of several photosynthetic enzymes, protein synthesis, oxidative metabolism,

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and the electrical balance of plant cell membranes [12]. Potassium, more than any other macro-element, is thought to enhance flowering [13], since it promotes the production of amino acids, which stimulate the creation of indole acetic acid (IAA) oxidase, which accelerates flower induction [14, 15]. As a result, potassium is critical to olive yield, quality and oil yield [16]. Olive needs a high K supply due to the high K concentration in the olive fruit [17]. Foliar treatment with potassium nitrate at 4% increased Picual olive fruit vegetative growth, nutritional status, productivity, fruit quality, and flesh oil content [18].

Magnesium is an important nutrient that aids in the development and creation of numerous sink organs, such as roots and seeds. Furthermore, in many horticultural crop species, fertilization has a significant influence on several physiological processes and productivity. Nonetheless, adequate foliar supply is critical for plant physiological and biochemical activities such as carbohydrate metabolism, enzyme activation, protein synthesis, and electricity transmission [19].

Magnesium foliar application plays an important role in the biochemical and physiological cycle of plants [20], for example, in the initiation of starch metabolism, protein synthesis, and electricity transfer. Magnesium acts as an impetus in the oxidation and reduction responses within plant tissues, and enhances plant resistance to drought [21].

Magnesium deficit during the plant development lowers yield and reduces quality [22]. Spraying magnesium sulphate four times on Manzanillo olive trees, led to a considerable increase in shoot and leaf development [23]. In January, olive trees receiving magnesium sulphate fertilizer (12-24 g/tree/year) showed an improvement in magnesium levels, as well as a substantial improvement in vegetative growth, flowering density, fruit set, and yield [24].

On the other hand, most soils have relatively low boron concentration, which is an issue. Plants can only use soluble boron in soils, which accounts for around 10% of the total soil boron concentration. Multiple factors contribute to boron deficit, including reduced soluble boron availability in the soil, weather conditions (drought, excessive precipitation), boron leaching, calcareous soils, and the cultivated crop species. Boron is involved in a variety of plant processes, including hormone transport, activated salt absorption, flowering and fruiting, and pollen germination, particularly its effects on pollen tube development directionality [25]. In addition, boron appears to play a significant role in fruit set [26], sugar and carbohydrate transport and synthesis. Boron deficiency reduces pollen production and results in poor fruit set in the olive plant's reproductive organs. Cuticle, cuticular cracks and

imperfections, stomata, trichomes, and specialized epidermal cells all play a role in foliar absorption of liquid solutions.

The consumption of extra virgin olive oil (EVOO) is currently gaining popularity due to its nutritional and health-promoting properties, which have been linked to the optimal balance of saturated, monounsaturated (MUFA), and polyunsaturated (PUFA) fatty acids, as well as minor components such as sterols, chlorophyll, polyphenols, and tocopherols; [27, 28].

Several studies showed that the beneficial properties of extra-virgin olive oil (EVOO) are due to the phenolic component [29] that confers to the EVOO free radical scavenging activity. Olive oils contain different classes of phenolic compounds such as cinnamic (caffeic acid and p-coumaric acid) and benzoic (vanillic acid) acids, phenyl ethyl alcohol (hydroxytyrosol and tyrosol), secoiridoids (oleuropein and ligstroside derivatives) and flavones (apigenin and luteolin), [30 - 32]. The main polyphenol in EVOO, hydroxytyrosol is a Reactive Oxygen Species (ROS) scavenger that reduces oxidized Low Density Lipoprotein (LDL) and platelets aggregations [33]. Oleuropein is an anti-inflammatory molecule that promotes nitric oxide production in macrophages [34]. Oleocanthal exerts anti-inflammatory properties similar to ibuprofen [35]. Polyphenols-rich EVOO is able to reduce heterocyclic amines and plasmatic C-reactive protein levels [36, 37], and ameliorate lipid metabolism and platelets function [38, 39], as well as glycaemia and insulin sensitivity [40]. Most of the phenolic compounds effects are due to their in vivo bioavailability [41].

The present study was carried out to improve the productivity and quality of olive fruit and olive oil of Coratina cultivar by foliar spray of potassium, magnesium and boron.

## 2. Experimental

### 2.1. Plant material

This experiment was carried out during 2018 and 2019 seasons on 36 years old Coratina olive trees in a private orchard located at El Behera governorate, Egypt. Trees were planted in a sandy soil under drip irrigation system, and they were almost similar in vigor and planted at 6.5 × 6.5 meters apart.

The experimental trees had received the same horticultural practices that suitable for tree age and the area weather conditions according to Ministry of Agriculture and Land Reclamation recommendation.

### 2.2. Treatments

Foliar applications with potassium dihydrogen phosphate ( $\text{KH}_2\text{PO}_4$ ) [known as mono potassium phosphate (MKP)] at 1%, magnesium sulfate ( $\text{MgSO}_4$ ) at 0.5 % and boric acid ( $\text{H}_3\text{BO}_3$ ) at 500

ppm, were sprayed solely or in combinations. Trees were sprayed twice a year, in July and September. Eight treatments were arranged in three replicates on one tree plot as follows:

- 1- (Control): Water spray only
- 2- (Mg): MgSO<sub>4</sub> at 0.5 %
- 3- (MKP): KH<sub>2</sub>PO<sub>4</sub> at 1%
- 4- (B): H<sub>3</sub>BO<sub>3</sub> at 500 ppm
- 5- (Mg + MKP): MgSO<sub>4</sub> at 0.5 % + KH<sub>2</sub>PO<sub>4</sub> at 1%
- 6- (Mg + B): MgSO<sub>4</sub> at 0.5 % + H<sub>3</sub>BO<sub>3</sub> at 500 ppm
- 7- (MKP + B): KH<sub>2</sub>PO<sub>4</sub> at 1% + H<sub>3</sub>BO<sub>3</sub> at 500 ppm
- 8- (Mg + MKP + B): MgSO<sub>4</sub> at 0.5 % + KH<sub>2</sub>PO<sub>4</sub> at 1% + H<sub>3</sub>BO<sub>3</sub> at 500 ppm

### 2.3. Measurements

At harvest time (late December), the examined trees were separately harvested and the following measurements were carried out as follows:

#### 2.3.1. Yield and fruit quality

Fruits of each replicate (tree) were separately harvested, weighed and the yield as kg/tree was measured, then samples of 90 fruits from the sprayed trees (30 fruits from each replicate tree) were picked randomly around the tree to determine the following parameters: Average fruit weight (gm), fruit length (cm) and fruit diameter (cm), [42].

#### 2.3.2. Oil extraction and its characteristics

After harvesting, about 5 Kg of fruits from treated and untreated Coratina olive trees were washed, crushed and packed in cheese cloth then pressed using a laboratory hydraulic press (carver). The pressure was 12,000 lb/in<sup>2</sup> for 30 min/ each sample which was reached gradually. The obtained juice was collected and left in a separator funnel until complete separation. The upper layer was separated and centrifuged. The extracted oil was dried using anhydrous sodium sulfate, then filtered with cotton wool and stored in dark glass bottles at -5°C till analysis.

- **Total lipid content:** It was determined as a percentage on dry matter weight in the oil samples according to the methods of A.O.A.C. [43].
- **Quality characteristics:** Peroxide value (PV) and free fatty acids % (FFAs) were evaluated following the methodology proposed by A.O.A.C. [43]. UV spectrophotometric indices (K232, K270, and ΔK) were measured according EEC. regulation methods [44].
- **Fatty acids composition:** Analysis and determination were carried out by preparation of methyl ester according to Cossiganani *et al.*, [45] followed by the identification of methyl esters using an Agilent 6890 series gas chromatograph apparatus equipped with a DB23 column (60 m X 0.32 mm X 0.25 μm), one

micro liter of FAME mixture was injected into the GC system with split/splitless injector and flame ionization detector (FID). The inlet temperature was 250 °C and the split ratio was 50:1. The carrier gas was nitrogen at 1.6 ml/min constant flow. The oven temperature was programmed at initial 150 °C, held for 1 min, followed by increase of 10 °C/min up to 170 °C, held for 5 min, followed by increase to 220 °C during 10min. held for 3 min. The detector was set at 270 °C with 450 ml/min airflow, 40 ml/min hydrogen flow, and 25 ml/min nitrogen makeup flow. Fatty acids standards were used to identify the peaks [45].

- **Total phenols content:** It was determined according to the method described by Gutfinger [46], and the results were expressed as mg of caffeic acid per kg of oil.
- **Phenolic compounds** of oil samples were identified and determined using HPLC (HP 1050) equipped with auto-sampling injector, solvent degasser, ultraviolet (UV) detector set at 280 nm and quaternary HP pump (series 1100). The column temperature was maintained at 35°C. Gradient separation was carried out with methanol and acetonitrile as a mobile phase at flow rate of 1 ml. Phenolic acids standards were dissolved in a mobile phase and injected into HPLC. Retention time and peak area were used to calculate of phenolic compounds concentrations by the data analysis of HP software [47].
- **Total pigments** (Chlorophyll and Carotenoids): 7.5 g of olive oil was accurately weighed and dissolved in cyclohexane up to a final volume of 25 ml., then chlorophyll and carotenoids were determined by measuring absorption at 670 and 472 nm, respectively, using a spectrophotometer (JENWAY 6405 UV/Vis., England) [48]. Results were expressed (mg/kg oil) as pheophytin and β-carotene for chlorophyll and carotenoids, respectively.
- **The unsaponifiable matter percentage** was evaluated as stated in A.O.A.C. [43].
- **Organoleptic test:** The organoleptic test was determined for the extracted oil according to IOOC [49].

#### 2.3.3. Leaf mineral content

Leaf samples were picked from each replicate tree then washed and dried at 70°C till a constant weight to determine nitrogen (N), phosphorus (P), and potassium (K) as percentage of dry weight basis due to the method described by Cottenie *et al.*, [50]. Boron as ppm was determined by the curcumin method of Dible *et al.*, [51]. Calcium and magnesium were determined following the criteria described by Moody *et al.*, [52].

#### 2.4. Statistical Analysis

The experimental layout was a randomized complete block design (RCBD) with three replicates for each treatment; one tree was represented as a replicate. The obtained data were subjected to analysis of variance (ANOVA) according to Snedecor and Cochran [53] and the least significant

differences (LSD) at probability of 5% were applied to separate the means using Co-Stat 4.11 software.

### 3. Results and discussion

#### 3.1. Fruit physical characteristics and yield

The physical measurements of Coratina olive sprayed with various treatments which previously mentioned, gave strong evidence that all treatments significantly affected them compared to water sprays (control) in the two studied seasons as shown in Table (1).

Regarding average fruit weight, the heaviest fruits (3.43 and 3.44 g) were weighed with B sprays, followed by Mg treatment (3.42 and 3.36 g) in both seasons, successively with significant differences among treatments. Whereas, the finest fruits (2.28 and 2.26 g) were weighed with the control sprays in the two seasons, respectively.

Concerning fruit length, the maximum values (2.31 and 2.29 cm) were recorded with B sprays, while the minimum lengths (2.00 and 1.97 cm) were measured with the control in both seasons, by order without significant difference among them.

As for fruit diameter, results show that the highest value of fruit diameter (1.66 and 1.68 cm) was scored by B sprays, followed by Mg treatment (1.64 cm) in the first season and MKP spray (1.66 cm) in the second season without significant differences among them. Whereas the lowest significant fruit diameter (1.39 and 1.40 cm) were obtained due to MKP+B sprays in the two seasons, respectively.

Considering the average of yield/ tree, it differed significantly among treatments, since in the first season; the highest value (103.33 kg) was recorded with Mg spray followed by 98.33 kg with MKP+B spray. On the contrary in the second season, the highest value 102.33 kg was recorded with MKP+B treatment followed by 96.33 kg that obtained by Mg spray. However, the lowest yield (63.33 and 66.00 kg) was measured with the control in both seasons, successively.

The above effects regarding fruit physical properties (fruit weight, fruit length and fruit diameter) show that the treatments improved the fruit physical parameters compared to the control. So, spraying boron alone was the most effective treatment followed by magnesium one. However, the tremendous impact of boron on the physical fruit properties may be due to that boron performs an crucial roles in the plant functions which include activate salt absorption, hormone movement, flowering and fruiting, and its effects on the directionality of pollen tube growth and pollen germination [25]. Also, boron appears to play a critical function in achieving sufficient fruit set synthesis, transport of carbohydrate and sugars which could have an effect on fruit weight, length and

diameter [26]. These outcomes are in agreement with the ones acquired through Abd-El-Migeed *et al.*, [54] on olive, Merwad *et al.*, [55, 56] on date palms and Merwad *et al.*, [57] on mango trees.

The acquired effects concerning the advanced impact of magnesium on fruit yield/tree can be attributed to the essential role of magnesium, due to the fact that sufficient supply via foliar application plays an essential role in physiological and biochemical processes of the plants such as metabolism of carbohydrates, enzymes activation, proteins synthesis and energy shifting [19]. In the current observe, magnesium sulphate treatment enhanced N, K and Mg leaves content (Table 6) than the other treatments which might also affect positively the fruits number/tree, consequently the consistent final yield/tree.

The received results are in harmony with the ones of Mostafa *et al.*, [58] who investigated the effect of foliar Mg treatment on leaf chlorophyll, vegetative growth, mineral content, yield and fruit quality of banana and found that Mg application affected positively the growth parameters, fruit quality and yield. Also, applying olives with magnesium sulphate in each of January, April, June and August enhanced the carbohydrates and leaf mineral contents (N, P, K and Mg), photosynthetic pigments, vegetative growth, flowering, fruit quality and caused an increase in fruit oil content (18-20%) compared to (12%) within the untreated trees, also the yield was doubled (43 kg/tree) compared to (20 kg/tree) for the untreated trees [59].

#### 3.2. Olive oil Percentage and quality parameters

Results in Table (2) show the effect of different foliar applications of magnesium (Mg), mono potassium phosphate (MKP), boron (B), and their mixtures on oil contents and quality parameters; free fatty acids% (FFAs) as oleic acid, peroxide value (PV) meq O<sub>2</sub> Kg<sup>-1</sup> oil and ultraviolet (UV) absorbency of the oils obtained from Coratina fruits in 2018 and 2019 seasons.

Acidity values ranged between 0.14–0.16 and 0.15–0.2% during 2018 and 2019, respectively. Concerning PV, it ranged from 2.5-2.7 and 2.59–2.9 meq O<sub>2</sub> Kg<sup>-1</sup> oil in 2018 and 2019 seasons, respectively.

There was a slight increase in values of FFAs and PV in the second season, these changes may be due to the extraction and climate conditions. On the other hand, these changes were not significant in the two successive seasons.

The highest value for acidity was 0.2% and for the PV was 2.8 meq O<sub>2</sub> Kg<sup>-1</sup> which were less than 0.8% and ≤ 20 meq O<sub>2</sub> Kg<sup>-1</sup>, respectively which are considered as reference values for these qualities[60]. So, all oil samples in this study are classified as extra virgin olive oil.

Treatment	Fruit weight (gm)		Fruit length (cm)		Fruit diameter (cm)		Yield/ tree (Kg)	
	2018	2019	2018	2019	2018	2019	2018	2019
Control	2.28	2.26	2.00	1.97	1.43	1.40	63.33	66.00
Mg	3.42	3.36	2.24	2.26	1.64	1.63	103.33	96.33
MKP	3.19	3.22	2.16	2.23	1.64	1.66	68.33	71.33
B	3.43	3.44	2.31	2.29	1.66	1.68	91.67	94.67
MKP+Mg	2.59	2.74	2.22	2.13	1.52	1.49	73.33	80.00
B+Mg	3.09	3.16	2.23	2.21	1.58	1.56	83.33	89.33
MKP+B	2.30	2.29	2.02	1.99	1.39	1.40	98.33	102.33
MKP+B+Mg	3.10	3.25	2.28	2.24	1.61	1.57	88.33	92.33
LSD (0.05)	0.09	0.14	0.16	0.12	0.03	0.03	4.67	4.17

Table 1

Effect of different fertilizing treatments on fruit physical characteristics and yield/tree in 2018 and 2019 seasons  
Control: water spray only, Mg: MgSO<sub>4</sub> at 0.5 %, MKP: KH<sub>2</sub>PO<sub>4</sub> at 1% and B: H<sub>3</sub>BO<sub>3</sub> at 500 ppm

Regarding to K<sub>232</sub> and K<sub>270</sub> are mainly indicating the conjugated dienes and trienes and also, the presence of carbonylic compounds. The results in Table (2) show that the values of K<sub>232</sub> and K<sub>270</sub> in all fertilizing treatments were lower than the control value with no significant differences in the two successive seasons. However, ΔK in the first season (2018) had the lowest value with MKP+B treatment (-0.0081) followed by MKP + B + Mg treatment (-0.0072), MKP treatment (-0.0071) and MKP+Mg treatment (-0.0059). The results were significantly different when compared to the control. In season 2019, there was a slight increase in ΔK, since MKP+B treatment had the lowest value of ΔK (-0.0074) followed by MKP treatment (-0.0067), B+Mg treatment (-0.0055) with a significant difference when compared to the control. All values of K<sub>232</sub>, K<sub>270</sub> and ΔK of Coratina olive oil were lower than the maximum permitted values of K<sub>232</sub>, K<sub>270</sub> and ΔK for extra virgin olive oils (2.50, 0.22 and 0.01), respectively [60].

It is clear from Table (2) that all treatments significantly increased oil content on fresh weight basis than the control in both seasons. Spraying trees with MKP+B+Mg gave the highest oil content (24.89% and 25.02%) in the two consecutive seasons respectively, followed by Mg (24.75% in the first season and 24.95% in the second one) and B+Mg (24.45% in 2018 and 24.90% in 2019). On the other hand, the control trees exhibited the lowest oil

content (20.20% and 20.90%) in both seasons, respectively. The previous studies show that oil content in Coratina olive fruits based on dry weight ranged between 40% - 46.3% and 35.97% - 46.48% [61, 62], while, it was 18.5% on fresh weight basis [63]. This progress in oil content for the two successive seasons of the study could be attributed to the increase of total photo-assimilates (e.g., lipids) as a result of applying magnesium and potassium which may control the biosynthesis of oil by enzymes [64] during the main metabolic pathway. Also, potassium fertilization is considered as essential in olive, particularly because K is found in high concentration in the fruit [65]. Moreover, potassium deficiency was found to diminish photoprotection mechanisms due to reduced photosynthetic and photorespiratory capacity. The lower CO<sub>2</sub> and O<sub>2</sub> assimilation rate in K-deficient trees caused elevated levels of excited energy [66]. Boron deficiency is more common than any other micronutrient; it also causes a reduction in pollen viability, poor pollen germination and pollen tube development [67]. This deficit decreases crop yield, consequently the oil content decreases, by reducing the number of perfect flowers. This is due to fertilizer applied at the right time and throughout the vital phases of the olive tree's development. Indeed, the accumulation of oil in the olive is a process that is influenced by the amount of carbohydrates present in fruits and old leaves [68].

Table 2

Effect of different fertilizing treatments on oil content of Coratina olive fruits and quality parameters of their oils in 2018 and 2019 seasons

Treatment	FFA% as oleic acid		PV (meqO <sub>2</sub> /kg oil)		K232		K270		ΔK		Oil %	
	2018	2019	2018	2019	2018	2019	2018	2019	2018	2019	2018	2019
Control	0.14	0.18	2.59	2.90	1.622	1.732	0.072	0.085	-0.0026	-0.0013	20.02	20.90
Mg	0.14	0.16	2.57	2.80	1.612	1.632	0.067	0.071	-0.0039	-0.0027	24.75	24.95
MKP	0.14	0.20	2.60	2.59	1.259	1.342	0.039	0.045	-0.0071	-0.0067	22.60	23.62
B	0.15	0.20	2.61	2.72	1.416	1.690	0.056	0.077	-0.0045	-0.0020	23.50	24.34

MKP+Mg	0.14	0.18	2.59	2.80	1.317	1.510	0.045	0.061	-0.0059	-0.0043	24.21	24.80
B+Mg	0.15	0.15	2.59	2.74	1.399	1.388	0.051	0.053	-0.0053	-0.0055	24.45	24.90
MKP+B	0.16	0.15	2.70	2.61	1.113	1.210	0.031	0.035	-0.0081	-0.0074	23.90	23.80
MKP+B+Mg	0.15	0.19	2.57	2.70	1.243	1.530	0.038	0.068	-0.0072	-0.0035	24.89	25.02
LSD (0.05)	N.S	N.S	N.S	N.S	N.S	N.S	N.S	N.S	-0.00299	-0.00243	2.18	1.26

### 3.3. Olive oil minor components and sensory attributes

Table (3) shows the minor components unsaponifiable matters% (UNSAF %), total polyphenolic contents, chlorophyll and carotenoids (in mg/kg) and organoleptic attributes (fruity, bitter and pungent) of the olive oils in different fertilizing treatments. The bioactive components of extra virgin olive oil are resulted from a number of variables acting before extraction (such as variety, environmental, climatic, soil and cultivation conditions, olive ripeness and olive healthy) and during olive oil extraction and storage [69].

The unsaponifiable fraction that represents less than 2% of olive oil is composed of a very large number of minor compounds which are very important for the flavour and the nutritional properties of EVOOs [70]. Results in Table (3) show that UNSAF% ranged from 1.01–1.32% and 0.99–1.41% in the two successive seasons respectively. The highest percentage was found due to MKP+B+Mg treatment in the two successive seasons. The differences between all foliar applications were not significant.

The content of phenolic compounds is an important parameter in the evaluation of extra virgin olive oil quality due to its correlation with oxidative stability and sensorial quality [71, 72]. Our results (Table 3) revealed that total phenols content ranged between 749.7–1056.61 ppm and 793.9–990.15 ppm in the two successive seasons (2018 and 2019) respectively, which were in line with the results of five different extra virgin olive oils (253–1400 ppm), [73]. All treatments showed high significant differences of total phenol content compared with the control treatment in the two successive seasons except MPK and MPK+B treatments which were lower than the control treatment in the first season (2018/2019). B + Mg treatment foliar application has the highest phenolic content in the two successive seasons (1056.61 ppm in 2018 and 990.15 ppm in 2019) followed by Mg treatment (972.4 ppm in 2018 and 989.6 ppm in 2019). Extra-virgin olive oil contains a considerable amount of phenolic compounds that are responsible for peculiar taste and for its high stability. Recent findings demonstrate that olive oil phenolics are powerful antioxidants, both in vitro and in vivo, and possess other potent biological activities that

could partially account for the observed healthful effects of the Mediterranean diet [74].

The main carotenoids and chlorophylls in olive oil are lutein and pheophytin, respectively. They are mainly responsible for the colour of virgin olive oil, ranging from yellow-green to greenish gold as well as oxidative stability due to their antioxidant nature in the dark and pro-oxidant activity in the light, [73]. There are also some reports about the benefits of dietary chlorophylls and carotenoids for human health [75]. Also chlorophyll act as pro-oxidant under light storage, whereas  $\beta$ -carotene minimizing lipid oxidation by its light filtering effect [48].

It is obvious from Table (3) that chlorophyll content ranged from 2.15 to 3.36 mg/Kg and from 2.89 to 3.5 mg/Kg in the two consecutive seasons 2018 and 2019, respectively. There were no significant differences between all treatments but, it is noticed that all magnesium treatments had the highest chlorophyll content. This may be related to that magnesium is considered the main component of chlorophyll.

Regarding to carotenoids content in Table (3), it ranged from 1.89 to 2.9 ppm and from 1.95 to 2.7 ppm in the two successive seasons 2018 and 2019, respectively. Also our results showed that all treatments were higher than that in control treatments in the two seasons except MKP foliar application was (1.89 ppm) lower than control treatments (1.93 ppm) in 2018 with no significant difference. MKP+Mg and MKP+ B+Mg treatments had the highest carotenoids content in the two seasons. Our results agreed with those revealed that the chlorophyll and carotenoid content ranged from 1.2-6.2 mg kg<sup>-1</sup> and 1-3.8 mg kg<sup>-1</sup>, respectively for different types of extra virgin olive oils [73].

Results in Table (3) show the effect of different foliar applications on sensory attributes of Coratina olive oils. One of the most important aspects of olive oil classification and value determination is sensory analysis. Human sensory evaluation is much more accurate (100 times) for olive oil than laboratory equipment for certain characteristics. The first and primary objective in sensory evaluation for olive oil is to determine if oils contain one or more of the defects that commonly occur in oils from improper fruit storage, handling, pest infestation, oil storage, or processing problems. The second objective of oil-sensory evaluation is to describe the positive characteristics of the oil in relation to its intensity of

olive-fruity character. Bitterness and pungency are often present in olive oils, especially when newly made. These regulations lead to the classification of oil as extra virgin (EVOO), virgin (VOO) or Lampante[64].

Concerning the median of positive fruity attributes ranged from 3.5 – 5.33 in the first season (2018). B+Mg, Mg and MKP+Mg treatments recorded the highest values of fruity 5.33, 4.67 and 4.5, respectively; they have significant differences compared with control value (3.5). Also, values of the other treatments were higher than the control value but with insignificant differences. All foliar

Table 3

Effect of different fertilizing treatments on minor components and sensory attributes of Coratina olive oils in 2018 and 2019 seasons

Treatment	Total phenols				Chlorophyll		Carotenoids		Fruity		Bitterness		Pungency		Defects	
	Content (mg/kg as caffeic acid)		UNSAF %		content (mg/kg)		content (mg/kg)									
	2018	2019	2018	2019	2018	2019	2018	2019	2018	2019	2018	2019	2018	2019	2018	2019
Control	824.90	793.90	1.01	1.30	2.80	2.90	1.93	1.95	3.50	3.00	2.50	3.00	1.87	1.75	0.00	0.00
Mg	972.40	989.16	1.28	1.20	3.31	3.08	2.50	2.39	4.67	4.83	2.83	3.50	1.83	2.33	0.00	0.00
MKP	749.70	834.35	1.29	1.40	2.15	2.90	1.89	1.98	3.83	4.83	2.00	3.00	1.67	2.17	0.00	0.00
B	866.90	870.34	1.20	0.99	2.96	3.00	2.25	2.00	3.50	3.58	2.33	3.50	1.50	2.57	0.00	0.00
MKP+Mg	921.83	950.19	1.28	1.19	3.01	3.50	2.90	2.60	4.50	5.00	2.53	3.33	1.92	2.27	0.00	0.00
B+Mg	1056.61	990.15	1.31	1.23	3.11	3.34	2.20	2.30	5.33	4.83	2.67	2.80	1.90	1.87	0.00	0.00
MKP+B	794.37	820.23	1.30	1.31	2.82	2.89	2.50	1.97	3.83	3.88	1.93	2.27	1.67	1.92	0.00	0.00
MKP+B+Mg	875.33	946.50	1.32	1.41	3.36	3.40	2.70	2.70	3.58	4.40	2.33	2.73	1.83	2.37	0.00	0.00
LSD (0.05)	9.4	7.5	N.S	N.S	N.S	N.S	0.23	0.40	0.70	0.55	N.S	N.S	N.S	N.S	0.00	0.00

Concerning the fatty acids composition, virgin olive oil has a prominent and well-balanced chemical composition of its fatty acids leads to high oxidative stability and health properties [76]. In particular, high levels of MUFAs (mainly oleic acid), which have health benefits and important for human nutrition are among the major components of the Mediterranean diet [77]. The high content of oleic acid in olive oil serves to slow down penetration of fatty acids into arterial walls [78]. Oils with higher monounsaturated fatty acids (MUFAs) and lower saturated fatty acids (SFAs) are preferred because of the proven beneficial effect of MUFAs on serum cholesterol levels, in turn; prevention of cardiovascular disorders [79, 80].

#### 3.4. Olive oil fatty acids composition

Results in Table (4) clarified the changes in fatty acid composition of Coratina olive oil as a result of different foliar fertilizations for olive trees as, total saturated fatty acids (TSFA), total unsaturated fatty acids (TUSFA), polyunsaturated fatty acids (PUSFA), monounsaturated fatty acids (MUSFA), TUSFA/TSFA ratio, and MUSFA/PUSFA ratio. The results in Table (4) reveal that, palmitic acid as main saturated fatty acid (16:0) varied between (12.33–13.06%) and (13.01–14.02%) in the two successive seasons. The highest content of palmitic acid was

found in control treatments, this mean that different foliar fertilizations decreased palmitic acid. On the other hand, oleic acid (18:1) as the main MUSFA varies between (70.55–73.81%) and (70.70 – 73.15%) in 2018 and 2019 respectively. The highest contents of oleic acid were found with B+Mg treatment in 2018 and MKP+B in 2019 respectively, while the lowest values of oleic acid were detected with MKP treatment (70.55% and 71.04%) in the two successive seasons, respectively. All foliar applications in the two successive seasons increased the percentage of oleic acid compared with the control except B and MKP treatments in the first season. With regard to linoleic acid (18:2), Mg, B and MKP treatments in the first season have higher contents of linoleic acid. On the other hand, in the second season all fertilizing treatments decreased the percentage of linoleic acid in contrary with control. In addition, linolenic acid (18:3) (which is susceptible to oxidation) contents were found less than 1% in all treatments. Generally, TUSFA increased as a result of most foliar applications in the two successive seasons in contrary with TSFA, which decreased in most foliar applications. Also, MUSFA/PUSFA ratio increased as a result of most foliar applications in the two successive seasons

since, by increasing this ratio, the oxidative stability of oil increases [81]. The fatty acids composition of all olive oil samples under study was in the range of International Olive Council (IOC) trade standard for EVOO [60].

The slight changes in fatty acids composition of olive oil in the two seasons may be related to the climate changes

. Table 4

Effect of different fertilizing treatments on relative percentage of fatty acids composition of Coratina olive oils in 2018 and 2019 seasons

Treatment FA%	Control		Mg		MKP		B		MKP+Mg		B+Mg		MKP+B		MKP+B+Mg	
	2018	2019	2018	2019	2018	2019	2018	2019	2018	2019	2018	2019	2018	2019	2018	2019
C <sub>16:0</sub>	13.06	14.02	12.33	13.80	12.42	14.01	12.72	13.85	13.01	13.16	12.84	13.01	13.02	13.06	13.00	13.05
C <sub>16:1</sub>	0.43	0.55	0.40	0.51	0.40	0.57	0.41	0.59	0.43	0.55	0.44	0.52	0.41	0.51	0.42	0.52
C <sub>17:0</sub>	0.05	0.05	0.05	0.04	0.04	0.05	0.05	0.05	0.05	0.05	0.05	0.04	0.05	0.04	0.05	0.05
C <sub>17:1</sub>	0.07	0.07	0.06	0.07	0.07	0.06	0.07	0.08	0.07	0.07	0.09	0.07	0.07	0.06	0.07	0.07
C <sub>18:0</sub>	1.98	1.79	2.03	1.73	2.01	1.76	2.02	1.70	2.04	1.70	2.03	1.70	2.01	1.75	1.87	1.71
C <sub>18:1</sub>	71.51	70.7	71.73	72.01	70.55	71.04	70.57	72.11	72.41	73.13	73.81	72.60	72.31	73.15	72.71	72.75
C <sub>18:2</sub>	10.98	10.9	11.51	10.00	12.60	10.5	12.28	9.80	10.02	9.50	8.85	10.21	10.15	9.60	9.99	10.1
C <sub>18:3</sub>	0.87	0.90	0.83	0.71	0.87	0.80	0.85	0.75	0.87	0.76	0.88	0.72	0.88	0.73	0.84	0.71
C <sub>20:0</sub>	0.43	0.51	0.44	0.55	0.43	0.57	0.43	0.52	0.46	0.55	0.43	0.55	0.46	0.54	0.43	0.52
C <sub>20:1</sub>	0.49	0.40	0.50	0.45	0.50	0.50	0.49	0.43	0.51	0.40	0.48	0.45	0.51	0.43	0.50	0.40
C <sub>22:0</sub>	0.11	0.13	0.12	0.13	0.11	0.13	0.11	0.12	0.13	0.13	0.10	0.13	0.13	0.13	0.12	0.12
TSFA	15.63	16.48	14.97	16.25	15.01	16.53	15.33	16.24	15.69	15.59	15.45	15.43	15.67	15.52	15.47	15.45
TUSFA	84.36	83.52	85.03	83.75	84.99	83.47	84.67	83.76	84.31	84.41	84.55	84.57	84.33	84.48	84.53	84.55
MUSFA	72.50	71.72	72.69	73.04	71.52	72.17	71.54	73.21	73.42	74.15	74.82	73.64	73.3	74.15	73.70	73.74
PUSFA	11.85	11.80	12.34	10.71	13.47	11.3	13.13	10.55	10.89	10.26	9.73	10.93	11.03	10.33	10.83	10.81
USFA/SFA	5.40	5.07	5.68	5.15	5.66	5.05	5.52	5.16	5.37	5.41	5.47	5.48	5.38	5.44	5.46	5.47
MUSFA/PUS FA	6.12	6.08	5.89	6.82	5.31	6.39	5.45	6.94	6.74	7.23	7.69	6.74	6.65	7.18	6.81	6.80

### 3.5. Olive oil phenolic compounds concentration

Phenolic compounds of olive oil have been of major interest to researchers due to their positive effects on both human health and preservation of olive oil. Results in Table (5) show the influence of different foliar applications on the phenolic compounds of Coratina olive oil. It could be noticed that, 16 phenolic compounds were detected in olive oil samples; 3-OH tyrosol, oleuropin, catechol, and pyrogallol were the predominant phenolic compounds in all olive samples, followed by coumarin, caffeic acid, p-OH benzoic, and benzoic acid. It could be observed that most of phenolic compounds were increased as a result of foliar applications especially MKP+B and MKP+B+Mg treatments, this increase was higher in the second season than the first one. 3-OH tyrosol compound had the greatest content comparing with the other

phenolic compounds since; it ranged between 125.6-198.9 ppm and 145.0-212.1 ppm in the two successive seasons. The greatest content of it was found due to MKP+B treatment in the two seasons. The highest concentration of oleuropin compound was detected with MKP+B treatment (198.2 ppm) in the second season and MKP+B+Mg treatment (170.3 ppm) in the first season. MKP and MKP+B+Mg treatments had the highest content of catechol (183.6 ppm) in the first season and (153.0 ppm) in the second one. Pyrogallol ranged between 83.9-115.09 ppm and 75.3-113.7 ppm in the two successive seasons respectively, where B+Mg treatment in 2018 and MKP+B+Mg treatment in 2019 had the highest content of it. Coumarin, caffeic acid, p-OH-benzoic acid and benzoic acid had considerable concentrations compared to the remained phenolic compounds such as catechin, vanilic and salicylic.

Olive oil phenolic compounds are shown to possess antioxidant, anti-inflammatory, anticancer, antidiabetic, antiobesity, cardioprotective, neuroprotective, antimicrobial, antisteatotic and many other effects [82, 83]. According to numerous

investigations, these effects are mostly attributable to the main secoiridoid derivatives such as oleuropein, oleocanthal and oleacein, and simple phenols hydroxytyrosol and tyrosol [84, 85].

Table 5

Effect of different fertilizing treatments on Phenolic compounds concentration (ppm) of Coratina olive oils in 2018 and 2019 seasons

Treatment Compounds	Control		Mg		MKP		B		MKP+Mg		B+Mg		MKP+B		MKP+B+Mg	
	2018	2019	2018	2019	2018	2019	2018	2019	2018	2019	2018	2019	2018	2019	2018	2019
3-OH Tyrosol	109.5	147.0	143.2	153.7	137.9	145.0	193.9	153.4	125.6	159.5	175.2	189.0	198.9	212.1	187.3	203.6
Oleuropein	99.4	120.2	124.6	135.04	120.8	131.0	145.0	139.0	137.3	149.2	140.9	136.1	147.3	198.2	170.3	155.6
Catechol	91.0	78.0	119.2	117.5	183.6	126.7	99.4	121.2	105.1	132.4	130.7	116.8	142.9	97.6	135.3	153.0
Pyrogallol	77.6	83.0	83.9	89.9	91.9	75.3	104.0	85.6	92.0	90.1	115.09	110.3	101.9	95.9	104.6	113.7
Coumarin	35.8	26.2	44.3	35.2	36.1	44.1	52.9	40.2	46.1	29.1	55.4	61.9	43.5	53.1	54.4	65.2
Caffeic	32.7	63.4	54.7	54.1	42.1	62.9	45.4	55.0	59.3	29.5	45.0	47.8	48.9	48.5	41.7	28.6
P-OH- benzoic	27.9	19.3	25.7	27.0	29.0	25.2	38.5	35.0	43.1	40.6	49.5	55.7	31.4	31.2	52.8	45.1
Benzoic	24.6	35.0	36.8	24.2	40.5	55.1	20.7	39.7	33.2	22.8	44.8	45.5	35.1	43.2	38.1	29.1
Gallic	18.7	27.9	17.3	34.1	19.6	32.1	15.4	29.6	16.3	22.3	27.4	42.8	23.5	29.3	31.4	47.1
chlorogenic	14.5	7.50	15.5	11.9	13.6	5.90	19.2	17.6	14.0	13.2	18.1	15.8	17.3	19.0	20.8	14.1
Vanillic	7.75	5.34	4.85	4.50	9.94	10.1	5.26	4.23	2.46	3.41	5.12	6.73	7.61	8.75	8.44	10.5
Caffeine	3.60	1.30	0.78	0.66	0.35	0.39	0.91	0.41	0.65	0.43	0.99	1.20	1.40	0.89	1.01	0.93
4-NH2 benzoic	2.48	1.91	1.20	2.33	1.41	2.05	0.91	0.79	1.12	1.93	1.26	3.01	0.87	1.74	1.03	2.93
Ferulic	1.23	0.92	0.85	0.55	1.90	1.49	0.48	0.87	0.72	1.29	1.71	1.50	1.43	1.03	1.54	1.70
Salicylic	1.34	1.91	2.57	3.14	2.60	3.87	2.94	2.50	1.29	3.61	1.43	2.91	1.69	3.01	2.26	4.01
Catechin	1.09	3.29	0.83	3.62	1.24	9.11	0.74	4.87	1.14	6.84	0.50	16.1	1.42	7.79	0.41	6.34

### 3.6. Olive leaves mineral content

As for olive leaves mineral content shown in Table (6), it could be seen that Mg treatment scored the highest N content (2.12 and 1.98%) in the first and second seasons, respectively followed by MKP treatment (1.97 and 1.89%), successively without significant differences between them in both seasons. However, the lowest significant N% (1.52 and 1.61%) was evaluated with the control treatment in both seasons, by order.

Regarding P content, it's obvious from Table (6) that the effect of all treatments was differed significantly and MKP treatment had recorded the maximum values in the two seasons (0.225 and 0.234%), respectively. Whereas the minimum values (0.188 and 0.182%), successively were recorded with the control treatment in both seasons.

Table (6) point out that, K% differed significantly among various treatments. Though, Mg and MKP

treatments, recorded the same highest values (1.62%) in the first season, while in the second season recorded 1.67% and 1.65%, respectively. The lowest values (1.24 and 1.29%) were measured with the control in the two seasons, by order.

Considering Ca%, Table (6) show that various treatments did not affect it significantly were the highest values (1.82%) were detected with MKP+ Mg and MKP+ B+ Mg treatments in the first season and (1.86%) with B+ Mg in the second season, while the lowest ones (1.52 and 1.64%), successively were detected with the control treatment in both seasons.

Concerning B leaf content tabulated in Table (6), a significant effect for treatments containing B was recorded since the highest content (35.2 and 34.7 ppm) was scored with B treatment solely, followed by MKP+ B+ Mg treatment (34.8 and 34.3 ppm) without significant difference between them, while the least content (23.2 and 22.8 ppm) was detected with the control treatment in both seasons by order.

Looking to Mg leaf content at Table (6), results show that the treatment containing Mg led to a significant increase in Mg content where the maximum values (2.31 and 2.22 %) were recorded with Mg treatment, followed by MKP+ Mg treatment (2.22 and 2.17 %), respectively without significant difference between them in the two seasons. Whereas, the minimum significant value (1.21 and 1.31 %) was measured with the control treatment in both seasons, successively.

Table 6

Effect of different fertilizing treatments on leaves mineral content in 2018 and 2019 seasons

Treatment	N (%)		P (%)		K (%)		Ca (%)		B (ppm)		Mg (%)	
	2018	2019	2018	2019	2018	2019	2018	2019	2018	2019	2018	2019
Control	1.52	1.61	0.188	0.182	1.24	1.29	1.52	1.64	23.2	22.8	1.21	1.31
Mg	2.12	1.98	0.215	0.224	1.62	1.67	1.74	1.65	24.3	25.3	2.31	2.22
MKP	1.97	1.89	0.225	0.234	1.62	1.65	1.71	1.65	24.7	24.9	1.82	1.84
B	1.74	1.65	0.194	0.191	1.31	1.38	1.78	1.72	35.2	34.7	1.85	1.87
MKP+Mg	1.99	1.97	0.212	0.213	1.57	1.59	1.82	1.84	25.4	26.1	2.22	2.17
B+Mg	1.85	1.82	0.194	0.193	1.31	1.28	1.81	1.86	32.4	31.9	1.95	1.91
MKP+B	1.77	1.75	0.195	0.199	1.55	1.58	1.76	1.78	32.4	32.1	1.81	1.72
MKP+B+Mg	1.78	1.85	0.211	0.198	1.46	1.52	1.82	1.79	34.8	34.3	1.98	1.92
LSD (0.05)	0.19	0.21	0.023	0.022	0.14	0.15	N.S	N.S	3.1	2.8	0.21	0.20

#### 4. Conclusion

It is clear from the previous results that the use of any of the examined treatments led to a varied increase among the treatments, in the yield of fruits per tree, the length and width of the fruit (physical characteristics), and also led to an increase in the oil accumulation in the fruits, with maintaining its quality, compared to the untreated trees.

Coratina olive is cultivated specifically for the production of oil, and since all the treatments did not affect the quality of the oil negatively, as they all give the characteristics of extra virgin olive oil, and preference is given to olive growers to obtain the highest economic return of the oil crop. Therefore, when calculating the oil yield per trees by multiplying the yield with the percentage of oil obtained from each treatment as shown in the figure (1), it is observed that spraying Mg solely recorded the highest oil yield (24.8 kg oil/tree) as an average of the two seasons, and thus this treatment was the superior and achieved an increase in oil yield by approximately 87% compared to the control, followed by MKP+B treatment with an average of 23.9 kg in the two seasons (approximately increased by 80% than the control). In this concern, the untreated trees (control) gave an average of 13.2 kg of oil/tree in both seasons.

Thus, the above can be summarized that the use of any of the treatments in this experiment led to a

noticeable increase in the yield of the resulting oil, compared to the control treatment, with maintaining its properties as (EVOO). But the treatment of Mg alone was the superior for reaching the highest oil yield followed by B+MKP treatment.

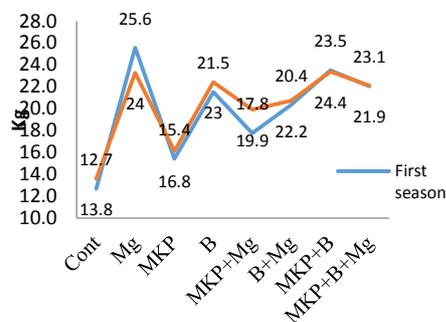


Fig. 1. Oil yeild/ tree (Kg)

#### 5. Conflicts of interest

“There are no conflicts to declare”.

#### 6. Formatting of funding sources

“There are no funding sources”.

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## 8. References

- [1] Desouky, I.M., L.F. Haggag, M.M.M.A. El-Migeed, Y.F.M. Kishk and E.S. El-Hady, (2009). Effect of boron and calcium nutrients sprays on fruit set, oil content and oil quality of some olive oil cultivars. *World J. Agric. Sci.*, 5: 180-185.
- [2] Brady, N.C. and R.R. Weil, (2002). *The Nature and Properties of Soils*. 13th Edn., Prentice Hall, New Jersey, ISBN: 0130167630, Pages: 960.
- [3] Mengel, K., (2002). Alternative or complementary role of foliar supply in mineral nutrition of plants. *Acta Hort.*, 594: 33-47.
- [4] Chapin, F.S., (1991). Effects of Multiple Environmental Stresses on Nutrient Availability and Use. In: *Response of Plants to Multiple Stresses*, Winner, W.E., E.J. Pell and J. Roy (Eds.). Academic Press, New York, ISBN: 978-0-08-092483-0, pp: 67-88.
- [5] Kene, H.K., S.T. Wankhade and B.N. Sagare, (1990). Influence of nutrients spray on yield and oil content of sunflower. *Ann. Plant Physiol.*, 4: 246-248.
- [6] Vafaie, A., A. Ebadi, B. Rastgou and S.H. Moghadam, (2013). The effects of potassium and magnesium on yield and some physiological traits of Safflower (*Carthamus tinctorius*). *Int. J. Agric. Crop Sci.*, 5: 1895-1900.
- [7] Jakobsen, S.T., (1993). Interaction between plant nutrients: III. Antagonism between potassium, magnesium and calcium. *Acta Agric. Scand. Sect. B: Soil Plant Sci.*, 43: 1-5.
- [8] Mirzapour, M.H., A.H. Khoshgoftar, S.K. Mirnia, H.A. Bahrami and M.R. Naeini, (2003). Interactive effects of potassium and magnesium on growth and yield of sunflower in a saline soil. *Iran. J. Soil Waters Sci.*, 17: 132-139.
- [9] Inglese, P., G. Gullo and L.S. Pace, (2002). Fruit growth and olive oil quality in relation to foliar nutrition and time of application. *Acta Hort.*, 586: 507-509.
- [10] Saykhul, A., C. Chatzissawidis, I. Therios, K. Dimassi and T. Chatzistathis, (2014). Growth and nutrient status of olive plants as influenced by foliar potassium applications. *J. Soil Sci. Plant Nutr.*, 14: 602-615.
- [11] Marschner, H., (1995). *Mineral Nutrition of Higher Plants*. 2nd Edn., Academic Press, London, Pages: 861.
- [12] Shabala, S., (2003). Regulation of potassium transport in leaves: From molecular to tissue level. *Ann. Bot.*, 92: 627-634.
- [13] Fabbri, A. and C. Benelli, (2000). Review article flower bud induction and differentiation in olive. *J. Hortic. Sci. Biotechnol.*, 75: 131-141.
- [14] Gonzalez-Garcia, F., M. Chaves, C. Mazuelos and A. Troncoso, (1972). Physiological aspects of the nutrition of the olive tree, 'Manzanillo' table variety. Cycle of nutrients in leaves and in growth of reproduction organs. *Physiol. Biochem. Hortic. Crops*, 32: 614-634.
- [15] Mazuelos, C., R. Romero, V. Valpuesta, R. Sarmiento and L. Catalina, (1983). Pyruvate kinase activity, levels of potassium and reducing sugars in vegetative and productive buds of *Olea europaea* L. trees. *Anales de Edafologia y Agrobiologia*, 42: 279-284.
- [16] Elloumi, O., M. Ghrab and M. Ben Mimoun, (2009). Responses of olive trees (cv. Chemlali) after five years of experiment to potassium mineral nutrition under rainfed condition. *Proceedings of the International Plant Nutrition Colloquium XVI*, August 26-30, 2009, Sacramento, California USA.
- [17] Fernandez-Escobar, R., R. Moreno and M.A. Sanchez-Zamora, (2004). Nitrogen dynamics in the olive bearing shoot. *HortScience*, 39: 1406-1411.
- [18] Hegazi, E.S., S.M. Mohamed, M.R. El-Sonbaty, S.K.M. Abd El-Naby and T.F. El-Sharony, (2011). Effect of potassium nitrate on vegetative growth, nutritional status, yield and fruit quality of olive cv. Picual. *J. Hort. Sci. Ornament. Plants*, 3: 252-258.
- [19] Muhammad A., Koko T., Fazal u. R., Muhammad S. S., Muhammad S. H., Muhammad I., Rohoma T., Jitendra M. (2021). Influence of Foliar Application of Magnesium on Horticultural Crops: A Review. *AGRINULA: Jurnal Agroteknologi dan Perkebunan*, 4 (1): 13-21.
- [20] Adnan, M., Hayyat, M. S., Imran, M., ur Rehman, F., Saeed, M. S., Mehta, J., and Tampubolon, K. (2020c). Impact of foliar application of magnesium fertilizer on agronomic crops: A review. *Indian Journal of Pure and Applied Biosciences*, 8 (6), 281-288. <http://dx.doi.org/10.18782/2582-2845.8465>.
- [21] Thaloorth, A. T., El-Zeiny, H. A., & Saad, A. O. M. (1990). Application of potassium fertilizer for increasing salt tolerance of broad bean (*Vicia faba* L.). *Bulletin of Egyptian Society for Physiological Sciences*, 10, 181-193.
- [22] Gerendas, J. and H. Fuhrs, (2013). The significance of magnesium for crop quality. *Plant Soil*, 368: 101-128.
- [23] El Khawaga, A.S., (2007). Improving growth and productivity of Manzanillo olive trees with foliar application of some nutrients and girdling under sandy soil. *J. Applied Sci. Res.*, 3: 818-822.
- [24] Wael, A.A., (2005). Improving growth and productivity of olive orchard under desert conditions. Ph.D. Thesis, Faculty of Agriculture, Cairo University, Egypt.
- [25] Robbertse, P.J., J.J. Lock, E. Stoffberg and L.A. Coetzer (1990). Effect of boron on directionality of pollen tube growth in *Petunia* and *Agapanthus*. *African J. Bot.*, 56: 487-492.
- [26] Khayyat, M.; Tafazoli, E.; Eshghi, S. and Rajae S. (2007). Effect of nitrogen, boron, potassium and zinc sprays on yield and fruit quality of date palm. *American-Eurasian J. Agric. & Environ. Sci.*, 2 (3): 289-296.
- [27] Cercaci, L., Passalacqua, G., Poerio, A., Rodriguez-Estrada, M. T., & Lercker, G. (2007). Composition of total sterols (4-desmethyl-sterols) in extravirgin olive oils obtained with different extraction technologies and their influence on the oil oxidative stability. *Food Chemistry*, 102, 66-76.
- [28] Lazzez, A., Perri, E., Caravita, M.A., Khlif, M., Cossentini, M., (2008). Influence of olive

- maturity stage and geographical origin on some minor components in virgin olive oil of the Chemlali Variety. *J. Agric. Food Chem.* 56 (3), 982–988.
- [29] Cicerale, S.; Conlan, X.A.; Sinclair, A.J.; Keast, R.S. (2009). Chemistry and health of olive oil phenolics. *Crit. Rev. Food Sci. Nutr.*, 49, 218–236.
- [30] Bianco, A.; Melchioni, C.; Ramunno, A.; Romeo, G.; Uccella, N. (2004). Phenolic components of *Olea europaea*—isolation of tyrosol derivatives. *Nat. Prod. Res.*, 18, 29–32.
- [31] Farag, R.S.; El-Baroty, G.S.; Basuny, A.M. (2003). Safety evaluation of olive phenolic compounds as natural antioxidants. *Int. J. Food Sci. Nutr.*, 54, 159–174.
- [32] Bendini, A., Cerretani, L., Carrasco-Pancorbo, A., Gómez-Caravaca, A. M., Segura-Carretero, A., Fernández-Gutiérrez, A., & Lercker, G. (2007). Phenolic molecules in virgin olive oils: a survey of their sensory properties, health effects, antioxidant activity and analytical methods. An overview of the last decade. *Alessandra. Molecules*, 12(8), 1679–1719.
- [33] Correa, J.A.; Lopez-Villodres, J.A.; Asensi, R.; Espartero, J.L.; Rodriguez-Gutierrez, G.; De La Cruz, J.P. (2009). Virgin olive oil polyphenol hydroxytyrosol acetate inhibits in vitro platelet aggregation in human whole blood: Comparison with hydroxytyrosol and acetylsalicylic acid. *Br. J. Nutr.*, 101, 1157–1164.
- [34] Virruso, C.; Accardi, G.; Colonna-Romano, G.; Candore, G.; Vasto, S.; Caruso, C. *Nutraceutical* (2014). Properties of extra-virgin olive oil: A natural remedy for age-related disease. *Rejuvenation Res.* 17, 217–220.
- [35] Beauchamp, G.K.; Keast, R.S.; Morel, D.; Lin, J.; Pika, J.; Han, Q.; Lee, C.H.; Smith, A.B.; Breslin, P.A. (2005). Phytochemistry: Ibuprofen-like activity in extra-virgin olive oil. *Nature*, 437, 45–46.
- [36] Monti, S.M.; Ritieni, A.; Sacchi, R.; Skog, K.; Borgen, E.; Fogliano, V., (2001). Characterization of phenolic compounds in virgin olive oil and their effect on the formation of carcinogenic/mutagenic heterocyclic amines in a model system. *J. Agric. Food Chem.* 49, 3969–3975.
- [37] Estruch, R.; Martinez-Gonzalez, M.A.; Corella, D.; Salas-Salvado, J.; Ruiz-Gutierrez, V.; Covas, M.I.; Fiol, M.; Gomez-Gracia, E.; Lopez-Sabater, M.C.; Vinyoles, E.; (2006). Effects of a Mediterranean-style diet on cardiovascular risk factors: A randomized trial. *Ann. Intern. Med.* 2006, 145, 1–11.
- [38] Covas, M.I.; de la Torre, K.; Farre-Albaladejo, M.; Kaikkonen, J.; Fito, M.; Lopez-Sabater, C.; Pujadas-Bastardes, M.A.; Joglar, J.; Weinbrenner, T.; Lamuela-Raventos, R.M.; (2006). Postprandial LDL phenolic content and LDL oxidation are modulated by olive oil phenolic compounds in humans. *Free Radic. Biol. Med.* 2006, 40, 608–616.
- [39] Dell’Aglì, M.; Maschi, O.; Galli, G.V.; Fagnani, R.; Dal Cero, E.; Caruso, D.; Bosisio, E. (2008). Inhibition of platelet aggregation by olive oil phenols via cAMP-phosphodiesterase. *Br. J. Nutr.* 99, 945–951.
- [40] D’Amore, S.; Vacca, M.; Cariello, M.; Graziano, G.; D’Orazio, A.; Salvia, R.; Sasso, R.C.; Sabba, C.; Palasciano, G.; Moschetta, A. (2016). Genes and miRNA expression signatures in peripheral blood mononuclear cells in healthy subjects and patients with metabolic syndrome after acute intake of extra virgin olive oil. *Biochim. Biophys. Acta*, 1861, 1671–1680.
- [41] Manna, C.; Galletti, P.; Maisto, G.; Cucciolla, V.; D’Angelo, S.; Zappia, V. (2000). Transport mechanism and metabolism of olive oil hydroxytyrosol in Caco-2 cells. *FEBS Lett.*, 470, 341–344.
- [42] A.O.A.C., (1995). *Official Methods of Analysis of AOAC International: Association of Official Analytical Chemists.* 13 th(Ed) Benjamin Franklin Station, Washington, D.C. USA. 495–510.
- [43] A.O.A.C., (2012). *Official Methods of Analysis of AOAC International: Association of Official Analytical Chemists.* 19 th(Ed). David Firestone, (chapter 41), pp.1-35.
- [44] EEC. (1991). Characteristics of olive and olive pomace oils and their analytical methods. Regulation EEC/2568. *Offic J Eur Commun*, 248, 1–82.
- [45] Cossiganani, L., Simonetti, M.S. and Damiani, P., (2005). Biocatalyzed acidolysis of olive oil triacylglycerols with 9c,11t and 10t,12c isomers of conjugated linoleic acid. *Eur. Food Res. Technol.*, 220, 267–271.
- [46] Gutfinger, T. (1981). Polyphenols in olive oils. *J. Am. Oil Chem. Soc.*, 61 (9): 966–968.
- [47] Goupy, P., Hugues, M., Biovin, P. & Amiot, M. J. (1999). Antioxidant composition and activity of barley (*Hordeum vulgare*) and malt extracts and of isolated phenolic compounds. *J. Sci. Food Agric.* 79:1625–1634.
- [48] Minguez-Mosquera, M.I., Rejano, L., Gandul, B., Sanchez, A. H. and Garrido, J. (1991). Color-Pigment correlation in virgin olive oil. *J. Am. Oil Chem. Soc.*, 68: 322–337.
- [49] IOOC. (2009). International Olive Oil Council. Trade Standard Applying to Olive Oil and Olive Pomace Oils. COI/T.15/NC no.3/Rev. 4 November.
- [50] Cottenie, A., Verloo M., Kiekens L., Velghe G. and Amerlynck R. (1982). *Chemical Analysis of Plant and Soil*, Laboratory of Analytical and Agroch. State Univ. of Belgium, Gent. pp. 63.
- [51] Dible WT, Troug E, Berger HC (1954). Boron determination in soils and plants. *Analytical Chemistry* 26:403–421
- [52] Moody, J. R., R. R. Greenberg, K. W. Pratt, and T. C. Rains (1988). Recommended inorganic chemicals for calibration, *Anal. Chem.*, 60, 1203A – 1218A.
- [53] Snedecor, G. W. and W.G Cochran (1980). *Statistical methods.* 7th ed. Iowa State Univ. Press, Ames, Iowa, USA pp. 507.
- [54] Abd-El-Migeed, M.M.M.; M.M.S. Saleh; E.A.M. Mostafa and M.S. Abou Raya (2002):

- Influence of soil and foliar applications of boron on growth, fruit set, mineral status, yield and fruit quality of Picual olive trees. *Egypt. J. Appl. Sci.*; 17 (1), 261 – 272.
- [55] Merwad, M.A., Eisa R.A., Ashour, N. E. and Saleh, M.M.S. (2014). Foliar spray of some growth regulators and nutrient elements for improving yield and fruit quality of Hayany date palm. *Middle East Journal of Agriculture Research*, 3(4): 751-756.
- [56] Merwad M.A.; E.A.M. Mostafa; N.E. Ashour and M.M.S. Saleh (2019): Effect of boron, zinc and seaweed sprays on yield and fruit quality of Barhee date palms. *Plant Archives*, 19 (2): 393-397.
- [57] Merwad, M.A., Eisa R.A., and Saleh, M.M.S. (2016). The beneficial effect of NAA, Zn, Ca and B on fruiting, yield and fruit quality of Alphonso mango trees. *International Journal of ChemTech Research*, 9(3): 147-157.
- [58] Mostafa, E. A. M., Sakeg, M. M. S., and El-Migeed Abd MM, M. (2007). Response of banana plants to soil and foliar applications of magnesium. *American-Eurasian Journal of Agricultural and Environmental Sciences*, 2(2), 141-146.
- [59] Hegazi, E.S., N.E. Kasim, T.A. Yehia, M.S. Abou Rayya and S.M. Thanana, (2016). Improving growth and productivity of olive trees through raising photosynthesis efficiency. *Res. J. Pharm. Biol. Chem. Sci.*, 7: 2697-2703.
- [60] IOC (2018). International olive council: trade standard applying to olive oils and olive-pomace oils. COT/T.15/NC3/Rev. 12.
- [61] Nissim Y, Shloberg M, Biton I, Many Y, Doron-Faigenboim A, Zemach H, et al., (2020). High temperature environment reduces olive oil yield and quality. *PLoS ONE* 15(4): e0231956. <https://doi.org/10.1371/journal.pone.0231956>
- [62] Morales A., Hurtado M.L. and Fichet T., (2014). Influence of Pruning Intensity of Olive Trees on the Oil Characteristics 'Coratina'. *Acta Horticulturae*. 1057: 717-723.
- [63] Eid, M. M. and M. E. I. Elsorady, (2012). Sterols compositions of some olive oil varieties cultivated under Egyptian conditions. *Egypt. J. Agric. Res.*, 90 (1) 263-276.
- [64] Mahmoud, T.S., N.E. Kassim and M.S. Abou Rayya, (2015). Effect of foliar application with dry yeast extract and benzyladenine on growth and yield of manzanillo olive trees. *J. Res. J. Pharmaceut. Biol. Chem. Sci.*, 6: 1573-1583.
- [65] López-Villalta, L.C. (1996). Production techniques. p.145-190. In: International Olive Oil Council (ed.), *World Olive Encyclopedia*. EGEDSA, Sabadell, Spain.
- [66] Erel, R., Yermiyahu, U., Ben-Gal, A., Dag, A., Shapira, O., and Schwartz, A. (2015). Modification of non-stomatal limitation and photoprotection due to K and Na nutrition of olive trees. *J. Plant Physiol.* 177, 1–10 <https://doi.org/10.1016/j.jplph.2015.01.005>. PubMed
- [67] Nyomora, A.M.S., (1995). The effect of boron deficiency on the reproductive processes of almond (*Prunus dulcis* [Mill] D.A. Webb) Ph.D. dissertation, Univ. California, USA.
- [68] Conde, C., S. Delrot and H. Geros, (2008). Physiological, biochemical and molecular changes occurring during olive development and ripening. *J. Plant Physiol.*, 165: 1545-1562.
- [69] Velasco, J. and Dobarganes, C. (2002). Oxidative stability of virgin olive oil, *Eur. J. Lipid Sci. Tech.*, 104:661-679.
- [70] Boskou D., Blekas G., Tsimidou M. (2006). Olive oil composition. In: Boskou D., editor. *Olive Oil Chemistry and Technology*. 2nd ed. Champaign: AOCS Press; p. 40–72.
- [71] Del Carlo, M. Sacchetti, G. Di Mattia, C. Compagnone, D. Mastrocola, D. Liberatore, L. and Cichelli, A. (2004). Contribution of the Phenolic Fraction to the Antioxidant Activity and Oxidative Stability of Olive Oil *J. Agric. Food Chem.*, 52,13, 4072-4079
- [72] Morello J.R., Vuorela S., Romero M.P., Motilva M.J. and Heinonen M., (2005). Antioxidant activity of olive pulp and olive oil phenolic compounds of the arbequina cultivar *J. Agric. Food Chem.*, 53 (6), 2002-2008.
- [73] Manai-Djebali, H. D. Krichène, Y. Ouni, L. Gallardo, J. Sánchez, E. Osorio, D. Daoud, F. Guido and M. Zarrouk (2012). Chemical profiles of five minor olive oil varieties grown in central Tunisia *J. Food Comp. Anal.* 27(2)109-119.
- [74] Visioli, F., & Galli, C. (2002). Biological properties of olive oil phytochemicals. *Critical reviews in food science and nutrition*, 42(3), 209-221.
- [75] Ferruzzi M.G., Blakeslee J. Digestion, (2007) absorption and cancer preventative activity of dietary chlorophyll derivatives. *Nutrition Research*. 27:1–12.
- [76] Guerfel, M., M.B. Mansour, Y. Ouni, D. Boujnah and M. Zarrouk, (2012). Compositional quality of virgin olive oils from cultivars introduced in two Tunisian locations. *African Journal of Agricultural Research*, 7(16): 2469-2474.
- [77] Sakouhi F, Herchi W, Sebei K, Ajsalon S, Kallel H, (2011) Accumulation of total lipids, fatty acids and triacylglycerols in developing fruits of *Olea europaea* L. *Sci Hort* 132:7-11.
- [78] Mailer, R., (2006). Chemistry and quality of olive oil. *PRIMEF ACT* 227, pp: 1-4.
- [79] Hashempour, A., R.F. Ghazvini, D. Bakhshi and S.A. Sanam, (2010). Fatty acids composition and pigments changing of virgin olive oil (*Olea europea* L.) in five cultivars grown in Iran. *AJCS* 4(4): 258-263 ISSN:1835-2707.
- [80] Aktas A. B., (2013) Chemical characterization of 'hurma' olive grown in karaburun peninsula. M.Sc. Thesis, Izmir Institute of Technology, Turkey.
- [81] Beltran, G., (2000). Influence of Ripening Process in *Olea europaea* L. Fruits on the Physicochemical Characteristics of the Oils. Ph.D. Thesis, Universidad de Jaen, Spain.
- [82] Cicerale, S.; Lucas, L.; Keast, R. (2012). Antimicrobial, antioxidant and anti-inflammatory phenolic activities in extra virgin olive oil. *Curr Opin. Biotech.*, 23, 129–135.

- 
- [83] Fabiani, R. (2016). Anti-cancer properties of olive oil secoiridoid phenols: A systematic review of in vivo studies. *Food Funct.* 7, 4145–4159.
- [84] Piroddi, M.; Albini, A.; Fabiani, R.; Giovannelli, L.; Luceri, C.; Natella, F.; Rosignoli, P.; Rossi, T.; Taticchi, A.; Servili, M.; (2017). Nutrigenomics of extra-virgin olive oil: A review. *BioFactors*, 43, 17–41.
- [85] Rigacci, S.; Stefani, M. Nutraceutical, (2016). Properties of Olive Oil Polyphenols. An Itinerary from Cultured Cells through Animal Models to Humans. *Int. J. Mol. Sci.*, 17, 843.