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Comparative Study of *Oxalis* L. Species Growing Wild in Egypt: GC-MS Analysis and Chemosystematic Significance

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Abstract

Five *Oxalis* L. plants belonging to three different sections were collected and subjected to GC-MS analysis to evaluate and understand the relationship between them from the chemosystematic point of view. The GC-MS analysis revealed the identification and characterization of thirty seven compounds belonging to different non-polar chemical classes of which nine compounds (two aliphatic hydrocarbons, one diterpene, one fatty acid, four fatty acid esters and one fatty alcohol) were identified before from the genus while, the rest compounds were identified for the first time. The GC-MS results were chemosystematically significant among *Oxalis* species and were found to be a good parameter in finding a tangible demarcation between *O. corniculata* and *O. corniculata* var. *repens*, on the other hand it is difficult to differentiate between *Oxalis* sections by the obtained data.

Keywords: Non-polar constituents; Oxalidaceae; Chemotaxonomy; Corniculatae; Cernuae; Ionoxalis.

1. Introduction

Genus Oxalis L. belongs to family Oxalidaceae R.Br. and comprising about 504 accepted species distributed worldwide specially in the area extending from South America to the southern part of North America and at the South-Western Cape region of South Africa [1-4]. The genus is represented in Egypt by five species and one variety viz; Oxalis anthelmintica A. Rich., O. corniculata L., O. corniculata var. repens (Thunb.) Zucc., O. corymbosa DC., O. latifolia Kunth and O. pes-caprae L. [2, 5, 6]. The data concerning isolation and identification of non-polar and volatile constituents from Oxalis species are limited. Only Karimzadeh et al. [7], Aruna et al. [8], Durgawale et al. [9] and Unni et al. [10] reported the chemical profile of O. corniculata using GC-MS technique and there were not any previous investigations for detecting the nonpolar constituents from the other species of the present study.

Some members of genus *Oxalis* has multiple therapeutic effects and used traditionally for treating various diseases such as *O. corniculata* which used as

anti-diabetic, anti-microbial, antioxidant and hepatoprotective [7-10].

Täckholm [5] and Kabuye [11] reported *repens* as a variety of *O. corniculata*. While, some other authors considered *O. corniculata* as a multiform species showing high variability of morphological expressions as a result of ecological variations [12] and they considered *O. corniculata* var. *repens* as a synonym of *O. corniculata* [2, 4]. The present study aims to evaluate the non-polar constituents of the five different *Oxalis* plants using GC/MS and studying the intraspecific relationships among the studied species.

2. Experimental

2.1. Plant material

In the present investigation, five plants (Four species & one variety) of genus *Oxalis* were identified by Dr. Eman M. Shamso in the Herbarium of Microbiology and Botany Dept., Faculty of Science, Cairo University. Samples were collected from different localities in Egypt and were deposited in the herbarium of Cairo University (CAI). *O. corniculata* L. (voucher No. A301), *O. corniculata*

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var. *repens* (voucher No. A302) and *O. corymbosa* DC. (voucher No. A303) were collected from National Research Centre gardens, Dokki, Giza in March, 2019, while *O. pes-caprae* L. (voucher No. A304) was collected from Bramly's grotto, Burg El-Arab in February, 2019 as well as *O. latifolia* Kunth (voucher No. A305) was collected from Faculty of Science gardens, Cairo University, Giza in April, 2019.

2.2. GC-MS analysis

The whole plants under investigation were dried and grounded to powder then 15 grams of each plant was soaked in 100 ml diethyl ether for 24 hrs. The extract was filtered using Whatman filter paper No.1 and the resulted filtrates were reduced under pressure using rotary evaporator then subjected to GC-MS analysis to identify their non-polar constituents. GC/MS analysis was carried out according to the methods of Hussein et al. [13] and Farid et al. [14] using Gas Chromatography (Thermo Scientific TRACE GC Ultra) directly coupled to ISQ Single Quadruple Mass spectrometer. GC/MS results are represented in (Table 1, Figure 1).

3. Results and discussion

3.1. GC-MS analysis

The GC-MS analysis revealed the identification and characterization of thirty seven compounds belonging to different non-polar chemical classes of which nine compounds were identified before from the genus while, the rest of compounds were identified for the first time (Table 1). It is apparent that almost all extracts have relatively the same concentration from aliphatic hydrocarbon compounds except for O. latifolia which has the lowest percent (17.86%) of alkene compounds and 6.63% of alkanes. Heptadecane is detected in O. corniculata and O. pes-caprae while triacontane is only present in O. corniculata var. repens and was isolated before from Leptadenia reticulate (Retz.) Wight & Arn. (Asclepiadaceae) [15]. In addition hentriacontane is absent from O. pes-caprae and O. corniculata. 1dodecene is lacking from O. corniculata var. repens and O. pes-caprae but present in the rest of species while in contrast, 1-tetradecene is found in O. corniculata var. repens and O. pes-caprae extracts not in all of the other plants.

O. corymbosa is the only species which has the ability of synthesizing cyclic aliphatic hydrocarbon compounds (cyclotetradecane, 13.18%) and those compounds were isolated before from Pelargonium graveolens L'Hér. (Geraniaceae Juss.) [16]. O. corniculata and O. pes-caprae are distinguished from the rest of species by being specialized in the formation of naphthalene derivatives which are bicyclic aromatic compounds and those compounds

were previously isolated from Euphorbia prostrata Aiton (Euphorbiaceae Juss.), Geranium robertianum L. (Geraniaceae) and Elettaria cardamomum (L.) Maton (Zingiberaceae Martinov.) [16-18]. Three terpenes were reported in this study; (E)-phytol is not detected this time in O. corniculata but was previously isolated by Aruna et al. [8] and Karimzadeh et al. [7]. limonene and linalylanthranilate were identified for the first time in Oxalis species; the former was previously separated from Averrhoa bilimbi L. (oxalidaceae) [19] while the latter was reported before from family lamiaceae Martinov [20, 21]. All carboxylic acid esters in the present study were not isolated before from genus Oxalis or even from the family but were identified from other families such as Acanthaceae Juss., Rosaceae Juss., Apiaceae Lindl. and Myrtaceae Schauer [22-26]. The carboxylic acid ester contents in O. corniculata var. repen is rare (0.37%) with comparison to O. corniculata (5.41%). Palmitic acid is the only fatty acid being present in all of O. species except for O. latifolia. In addition, all investigated species have the ability to produce fatty acid methyl esters except O. corymbosa which characterized by lacking of such compounds.

3.2. Chemosystematic evaluation

According to the systems proposed by Lourteig [27, 28] and Salter [29], *Oxalis* was divided into three sections; Corniculatae DC. to which *O. corniculata* belongs, section Cernuae R. Knuth includes *O. pescaprae* and finally, section Ionoxalis (small) R. Knuth which comprises *O. corymbosa* and *O. latifolia*.

O. repens was firstly described by Thunberg and later, Zuccarini lowered it to be a variety (*Oxalis corniculata* var. *repens*) [30, 31] and this was accepted by Täckholm [5] and Kabuye [11] while Lourteig [32] synonymized it with *O. corniculata* which was accepted by Boulos [2] and The plant list [4].

By comparing the non-polar chemical constituents of O. corniculata var. repens and O. corniculata that were collected from the same habitat, we found that each plant has a different chemical pattern not only in the type of identified compounds but also in their percentage. O. corniculata var. repens was characterized by presence of nine different compounds in its extract that were not detected in O. corniculata (Table 1) and O. corniculata contain eight compounds not recognized in the var. repens (Table 1). In addition, the fatty acid methyl ester content in O. corniculata var. repens (12.14%) is much more than that of *O. corniculata* (1.97%), the fatty alcohols and aldehydes represents (8.96%, 14.08%) of O. corniculata var. repens total extract and (2.19%, 27.93%) of O. corniculata extract respectively and due to these variations we agree with

Täckholm [5] and Kabuye [11] in treating *O. corniculata* var. *repens* as a variety of *O. corniculata* not as a synonym or even as a multiform species as proposed by Malik *et al.* [12].

From the GC/MS data we can distinguish between each of the studied species, where *O. pes-caprae* can be recognized by the absence of dodecane and E-14hexadecenal compounds in its extract, *O. corymbosa* characterized by presence of cyclic aliphatic hydrocarbon compounds (cyclotetradecane) and absence of fatty acid methyl ester compounds; *O.* *latifolia* can be separated from the other species by lacking of fatty acids (Palmitic acid) in its extract.

Although the present study helps in understanding the chemical relationship among *Oxalis* species particularly in the differentiation between *O. corniculata* var. *repens* and *O. corniculata* but when studying the relationship on the section level the nonpolar constituent is difficult to distinguish them and more chemical investigations are needed for providing a complete image for understanding the variations among *Oxalis* sections.

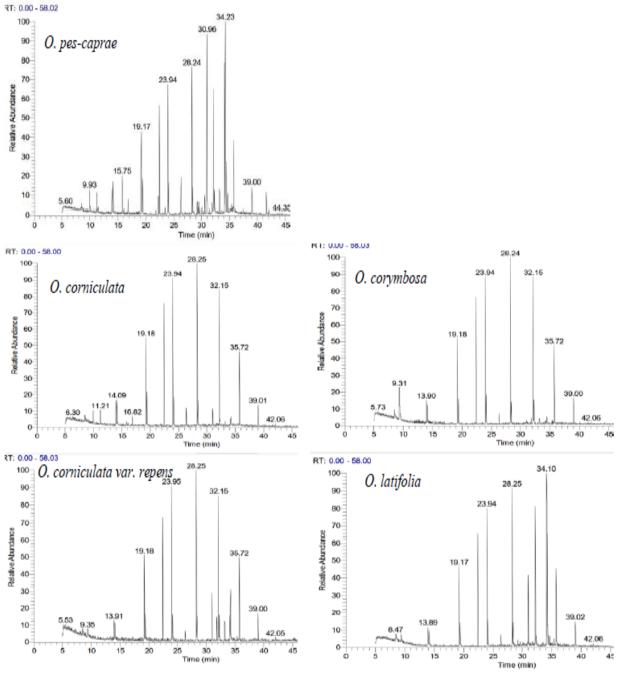


Figure 1. GC/MS analysis of non-polar fractions of Oxalis

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| Section | | | | Cernuae Corniculatae | | | Ionoxalis | |
|--|---------------|------------|--|----------------------|---------------------------------------|--|----------------|----------------|
| Oxalis | | | | pes-caprae | corniculata | <i>corniculata</i> var. <i>repens</i> | corymbosa | latifolia |
| Compound name | Rt/ min. | M.wt | M. formula | | | Area % | | |
| 1-Aliphatic hydrocarbons: | | | | | | | | |
| (A)- Acyclic aliphatic | hydrocarbo | ns: | | | | | | |
| - Alkanes: | 14.00 | 170 | C II | 1 77 | | | | r |
| Dodecane Tridecane | 14.09 14.1 | 170 184 | C ₁₂ H ₂₆ C ₁₃ H ₂₈ | 1.77 | - 1.05 | - | - | - |
| Tetradecane | 19.38 | 198 | C ₁₃ H ₂₈ C ₁₄ H ₃₀ | 1.75 | 3.21 | 2.33 | 2.28 | 1.85 |
| Hexadecane | 24.11 | 226 | C ₁₆ H ₃₄ | 1.6 | 2.58 | 2.1 | 2.53 | 1.78 |
| Heptadecane* | 26.3 | 240 | C ₁₇ H ₃₆ | 1.99 | 1.83 | - | - | - |
| Hentriacontane | 26.31 | 436 | C31H64 | - | - | 0.9 | 1.14 | 1.07 |
| Pentatriacontane | 28.38 | 492 | C35H72 | 1.41 | 2.37 | 2.17 | 2.39 | 1.93 |
| Triacontane | 32.26 | 422 | C ₃₀ H ₆₂ | - | - | 2.09 | - | - |
| -Alkenes: | 0.47 | 110 | C II | | 1.20 | 1.10 | 1.04 | 0.00 |
| 1-Octene 1-Dodecene | 8.47 13.89 | 112 168 | C ₈ H ₁₆ C ₁₂ H ₂₄ | - | 1.28 2.81 | 1.18 | 1.04 2.42 | 0.89 |
| 1-Tetradecene | 13.89 | 196 | C12H24 C14H28 | 1.52 | - | 2.23 | - | - |
| 4-Tetradecene | 19.19 | 196 | C14H28 | 4.51 | 7.4 | 6.99 | 7.58 | 5.45 |
| 1-Hexadecene | 23.94 | 224 | C16H32 | 6.92 | - | 11.73 | - | - |
| 1-Nonadecene* | 32.15 | 266 | C19H38 | 6.7 | 12.52 | 11.75 | 12.95 | 9.79 |
| 1-Docosene | 35.72 | 308 | C22H44 | 4.18 | 6.6 | - | 7.34 | - |
| Subtotal | | | | 32.35 | 41.65 | 43.47 | 39.67 | 24.49 |
| (B)- Cyclic aliphatic hy | | | a r | | 1 | | 10.10 | r |
| Cyclotetradecane | 23.95 | 196 | C14H28 | - | - | - | 13.18 | - |
| 2-Aromatic compounds: (A)- Monocyclic: | | | | | | | | |
| 2-tert-Butyl-4-isopropyl-5 – | | | | | | | | |
| methylphenol | 22.39 | 206 | $C_{14}H_{22}O$ | 5.86 | 11.11 | 9.87 | 10.69 | 7.99 |
| Subtotal | | | | 5.86 | 11.11 | 9.87 | 10.69 | 7.99 |
| (B)- Bicyclic: | | | | | • | | | |
| Naphthalene, decahydro, cis | 9.94 | 138 | C10H18 | 1.94 | 1.9 | - | - | - |
| Subtotal | | | | 1.94 | 1.9 | - | - | - |
| 3-Terpenes: | | | | | | | | |
| (A)- Acyclic diterpenes (E) Phytol * | 34.44 | 296 | C ₂₀ H ₄₀ O | 3.09 | - | - | 0.74 | 1.12 |
| (B)- Monocyclic terpen | | 290 | C201140O | 3.09 | | - | 0.74 | 1.12 |
| Limonene | 9.34 | 136 | C10H16 | - | - | 1.38 | 4.69 | 0.86 |
| Linalyl anthranilate | 15.75 | 273 | C ₁₇ H ₂₃ NO ₂ | 2.03 | 0.55 | _ | - | - |
| Subtotal | | | -17-232 | 5.12 | 0.55 | 1.38 | 5.43 | 1.98 |
| 4-Carboxylic acid esters: | | | | 5.12 | 0.55 | 1.50 | 5.45 | 1.70 |
| Carbazic acid, 3-(1- | 7.04 | 200 | G U NO | | 0.61 | 0.27 | | [|
| propylbutylidene)-ethyl ester | 7.24 | 200 | $C_{10}H_{20}N_2O_2$ | - | 0.61 | 0.37 | - | - |
| Oxalic acid, allyl nonyl ester | 14.1 | 256 | $C_{14}H_{24}O_4$ | - | 2.64 | - | - | - |
| Oxalic acid, allyl decyl ester | 14.12 | 270 | $C_{15}H_{26}O_4$ | - | - | - | 2.03 | 1.25 |
| Oxalic acid, allyl nonyl ester | 32.25 | 256 | $C_{14}H_{24}O_4$ | 1.22 | 2.16 | - | - | - |
| (isomer) Oxalic acid, allyl decyl ester | | | | | - | | - | - |
| (isomer) | 32.26 | 270 | $C_{15}H_{26}O_4$ | - | - | - | 2.13 | 1.53 |
| Subtotal | | | | 1.22 | 5.41 | 0.37 | 4.16 | 2.78 |
| 5-Fatty acids: | · | | | • | · · · · · · · · · · · · · · · · · · · | | - | · |
| Palmitic acid* | 33.2 | 256 | C16H32O2 | 1.39 | 0.5 | 1.47 | 0.67 | - |
| Subtotal | | | | 1.39 | 0.5 | 1.47 | 0.67 | - |
| 6- Fatty acid methyl esters: | 1 | | 1 | T | | | | 1 |
| Hexadecanoic acid, methyl ester* | 30.97 | 270 | C17H34O2 | 10.7 | 1.97 | 4.37 | - | 5.27 |
| Einolelaidic acid, methyl | | | | | | | 1 | |
| ester* | 34.11 | 294 | $C_{19}H_{34}O_2$ | 8.8 | - | 1.77 | - | 12.15 |
| Methyl linolenate * | 34.24 | 292 | C19H32O2 | 13.59 | - | 6 | - | 11.26 |
| Octadecanoic acid, methyl | 34.68 | 298 | C19H38O2 | 1.09 | _ | _ | _ | 1.08 |
| ester* | 54.00 | 290 | C19113802 | | - | | - | |
| Subtotal | | | | 34.18 | 1.97 | 12.14 | - | 29.76 |
| 7-Esters: | 21.9 | 270 | Culture | 1 | | 2.14 | - | |
| Butyl phthalate Diisooctyl phthalate | 31.8 41.55 | 278 390 | C ₁₆ H ₂₂ O ₄ C ₂₄ H ₃₈ O ₄ | - 1.57 | - | 2.14 | - | - |
| Subtotal | -11.55 | 570 | 024113604 | 1.57 | - | 2.14 | - | - |
| 8-Fatty alcohols | I | | | | | | | I |
| 1-Docosanol* | 35.73 | 326 | C22H46O | - | - | 6.55 | - | 5.82 |
| 1-Hexacosanol | 39.01 | 382 | C ₂₆ H ₅₄ O | 1.6 | 2.19 | 2.41 | 2.81 | 1.88 |
| Subtotal | | | | 1.6 | 2.19 | 8.96 | 2.81 | 7.7 |
| 9-Fatty aldhydes | | | | 1 | 1 4 8 9 5 | 1 | - | |
| E-14-Hexadecenal | 23.95 | 238 | C ₁₆ H ₃₀ O | - | 12.93 | - | - | 9.39 |
| E-15-Heptadecenal Subtotal | 28.25 | 252 | C17H32O | 8.04 8.04 | 15 27.93 | 14.08 14.08 | 15.67 | 11.43 |
| Total | | | | 93.27 | 93.21 | 93.88 | 15.67 92.28 | 20.82 95.52 |
| Total | I | | | | 93.21 | 23.00 | 72.20 | 75.52 |

Table (1): GC-MS analysis of four species and one variety of Oxalis L.

*Compounds identified before from genus *Oxalis* and detected in the present study.

4. Conclusion

Thirty seven non-polar compounds were recognized from five *Oxalis* species, twenty eight of them were reported for first time in the genus. The GC-MS profiling of *O. corniculata* var. *repens* and *O. corniculata* supported the treatment of *repens* as a variety of *corniculata*. The non-polar constituents of the studied plants are chemosystematically significant on the species level and difficult to be used on the section level of taxonomy.

5. Conflicts of interest

Authors declare that there are no conflicts of interest.

6. Acknowledgments

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7. References

- Denton M. F., A monograph of *Oxalis* section Ionoxalis (Oxalidaceae) in North America. Publication of the Michigan State University Museum, Biological Series 4, 445-615(1973).
- [2] Boulos L., Flora of Egypt; Al Hadara Publishing, Cairo - Egypt vol. 1 (1999).
- [3] Oberlander K.C., Dreyer L.L. & Esler K.J. Biogeography of Oxalis (Oxalidaceae) in South Africa: a preliminary study. Bothalia - African Biodiversity and Conservation 32(1) (2002).
- [4] The plant list. (2013) Version 1.1 published on http://www.theplantlist.org/tpl1.1/search?q=oxalis
- [5] Täckholm V., Students' Flora of Egypt, ed.2, Cairo University (1974).
- [6] El-khanagry S.S.G., New records of Dicotyledonous taxa to the Flora of Egypt. Bulletin of the Faculty of *Agriculture*, Cairo Univ., 56, 97 (2005).
- [7] Karimzadeh K., Bakhshi N. and Ramzanpoor M., Biogenic silver nanoparticles using Oxalis corniculata characterization and their clinical implications. Journal of Drug Delivery Science and Technology, 54, 101263(2019).
- [8] Aruna K., Rajeswari P. D. R., Ramkumar M., Prabu K., Chidambaram R. and Sankar S. R., GC-MS analysis of the ethanol extract of *Oxalis corniculata* L. (Oxalidaceae). *International Journal of Green and Herbal Chemistry*, 3(3), 912-917 (2014).
- [9] Durgawale P. P., Hendre A. S. and Phatak R. S., GC/MS characterization, antioxidant and free radical scavenging capacities of methanolic extract of *Oxalis* corniculata Linn: An ayurvedic herb. Rasayan Journal of Chemistry, 8(3), 271-278 (2015).
- [10] Unni B. G., Borah A., Wann S. B., Singh H. R., Devi B. and Bhattacharjee M., Phytochemical and antibacterial study of traditional medicinal plants of North East India on *Escherichia coli*. Asian Journal of *Experimental* Sciences, 23(1), 103-108 (2009).
- [11] Kabuye C.H.S., Oxalidaceae. In: E. Milne-Redhead & R. M. Polhill (eds.), Flora of Tropical East Africa. Balkema, Rotte (1971).
- [12] Malik M. I., Mahmood S., Yasin G. and Bashir N., Oxalis corniculata as a successful lawn weed: a study

of morphological variation from contrasting habitats. *Pakistan Journal of Botany*, *44*, 407-411 (2012).

- [13] Hussein S.R., Abdel Latif R.R., Marzouk M.M., Elkhateeb A., Mohammed R. S., Soliman A. A. F. and Abdel-Hameed E.S., Spectrometric analysis, phenolics isolation and cytotoxic activity of *Stipagrostis plumosa* (Family Poaceae). *Chemical Papers*, 72, 29–37 (2018).
- [14] Farid M. M., Ragheb A. Y., El-Shabrawy M., Marzouk M. M., Hussein S. R., Soliman A. A., Hussein T. and Kawashty S.A., GC-MS and LC-ESI-MS analysis of biologically active fractions from *Verbascum letourneuxii*; efficient protocol for in vitro propagation. *Biocatalysis and Agricultural Biotechnology*, 29, 101817 (2020).
- [15] Mallick S.S. and Dighe V.V., Detection and Estimation of alpha-Amyrin, beta-Sitosterol, Lupeol, and n-Triacontane in Two Medicinal Plants by High Performance Thin Layer Chromatography. *Advances in Chemistry*, 43948, 1-7 (2014).
- [16] Dashamiri S., Ghaedi M., Naghiha R., Salehi A. and Jannesar R., Antibacterial, anti fungal and E. coli DNA cleavage of *Euphorbia prostrata* and *Pelargonium* graveolens extract and their combination with novel nanoparticles. *Brazilian Journal of Pharmaceutical Sciences*, 54(4) (2018).
- [17] Graça V. C., Ferreira I. C. and Santos P. F., Phytochemical composition and biological activities of *Geranium robertianum* L.: A review. *Industrial Crops* and Products, 87, 363-378 (2016).
- [18] Rabu P., Edayadulla N. and Nnand s., In vitro antibacterial activity and gas chromatography-mass spectrometry analysis of ethanolic extract of leaves of *Elettaria cardamomum* L. Maton, *Asian Journal of Pharmaceutical and Clinical Research*, 12(1), 73-75 (2019).
- [19] Azeem A. K. and Vrushabendraswami B. M., Evaluation of physicochemical and volatile components of leaf ethanolic extract of *Averrhoa bilimbi*. *Journal of Innovations in Pharmaceuticals and Biological Sciences*, Vol 2(4), 646-648 (2015).
- [20] Adaszyńska M., Swarcewicz M., Dzięcioł M. and Dobrowolska A., Comparison of chemical composition and antibacterial activity of lavender varieties from Poland. *Natural product research*, 27(16), 1497-1501(2013).
- [21] Navarrete A., Ávila-Rosas N., Majín-León M., Balderas-López J. L., Alfaro-Romero A. and Tavares-Carvalho J. C., Mechanism of action of relaxant effect of Agastache mexicana ssp. mexicana essential oil in guinea-pig trachea smooth muscle. *Pharmaceutical biology*, 55(1), 96-100 (2016).
- [22] Zeidan S., Hijazi A., Rammal H., Al Bazzal A., Annan H. and Al-Rekaby A. A. N., Determination of bioactive compound content and antioxidant activity of the Labanese *Eryngium creticum*. Eu.Chem.Bul. 4(11), 498-502 (2015).
- [23] Granados-Chinchilla F., Villegas E., Molina A. and Arias C., Composition, Chemical Fingerprinting and Antimicrobial Assessment of Costa Rican Cultivated Guavas (*Psidium friedrichsthalianum* (O. Berg) Nied. and *Psidium guajava* L.) Essential Oils from Leaves and Fruits. *Natural Products Chemistry & Research*, 4, 236 (2016).
- [24] Leena P., Hukuman N. H. Z. and Jisha M., In vitro antimicrobial efficacy and GC-MS analysis of bioactive components from Lepidagathis keralensis

(Acanthaceae). World Journal of Pharmaceutical Research, 5(12), 937-948 (2016).

- [25] Sithara N. V., Komathi S. and Rajalakshmi G., Identification of bioactive compounds using different solvents through FTIR studies and GC-MS analysis. *Journal of Medicinal Plants Studies*, 5(2), 192-194 (2017).
- [26] Shaheen N., Qureshi N. A., Ashraf A., Hamid A., Iqbal A. and Fatima H., In vitro anti-leishmanial activity of Prunus armeniaca fractions on Leishmania tropica and molecular docking studies. *Journal of Photochemistry and Photobiology B: Biology*, 213, 112077 (2020).
- [27] Lourteig A., Oxalis L. subgénero Thamnoxys (Endl.) Reiche emend. Lourteig. Bradea, Boletim do Herbarium Bradeanum 7, 1–199 (1994).

- [28] Lourteig A., Oxalis L., subgenera Monoxalis (Small) Lourt., Oxalis y Trifidus Lourt. Bradea., 7, 201-629 (2000).
- [29] Salter T.M., The genus Oxalis in South Africa: a taxonomic revision. South African Journal of Botany, 1,1–355 (1944).
- [30] Nesom G. L., Again: taxonomy of yellow-flowered caulescent Oxalis (Oxalidaceae) in eastern North America. Journal of the Botanical Research Institute of Texas, 727-738 (2009).
- [31] Groom Q. J., Van der Straeten J. and Hoste I., The origin of *Oxalis corniculata* L. *Peer Journal*, 7, e6384 (2019).
- [32] Lourteig A., Oxalidaceae extra-austroamericanae: 2. Oxalis L. Sectio Corniculatae DC. Phytologia 42,57 (1979).