



# Utilization of seafood byproducts: biological activities and biotechnological applications

CrossMark

Shaymaa A Ismail<sup>\*</sup>, Amira A Hassan

Department of Chemistry of Natural and Microbial Products, Pharmaceutical and Drug Industries researches institute, National Research Centre, El Bohouth Street, Dokki, Giza 12622, Egypt.

#### Abstract

The global demands for seafood products are steadily increasing and consequently numerous amounts of solid and liquid byproducts are generated that usually discarded leading to serious problems including loss of nutrients and environmental pollution that consequently negatively affect the human health. These byproducts are considered as an undiscovered treasure having the potentiality for the production of various probiotics as well as several biomaterials possessing multiple functional and biological activities. Nitrogenous compounds, lipids, polysaccharides and minerals are the main constitutive components that can be recovered or converted to value added products with potential nutritional, biomedical and pharmaceutical applications. Biological conversions via microbial fermentation and/or enzymatic treatment are eco-friendly economic processes that are widely applied leading to the production of bioactive protein hydrolysates, chitin based products and various industrial enzymes. In addition, algae based bioconversion is an efficient method for the valorization of seafood industry effluents via the production of various bio-refineries.

Keywords: Seafood, solid byproducts, effluents, value added products

# 1. Introduction

The overgrowth in the human population enforces a global interest for the use of seas, oceans and marine resources for sharing a blueprint for the prosperity of people as well as sustainable development [1]. Seafood industry including either finfish or shellfish, has become one of the main sectors in food industry in which the global output of capture fisheries had been increased from 20 million ton in 1950 to 81.5 million ton in 2014 [2] reaching its highest recorded value (96.4 million ton live weight) in 2018 [1]. Additionally, the aquaculture sector provides 16% of the edible animal protein and has been estimated as a crucial component for providing food security for 9.8 billion people all over the world by 2050 [3].

In seafood industry, 80% of the total harvest has been processed into dried, smoked, frozen, marinated and other products in which several pre-processing operations including removal of heads, shells, scales, skin, gut and fins in addition to washing, filleting and others have been performed. These processes produce numerous amounts of solid and liquid byproducts that usually discarded causing environmental pollution and negatively affect the human health. In addition to incineration of these solid byproducts causing air pollution, huge amounts of by-catch are dumped in the oceans or nearby land. Microbial anaerobic fermentation of the dumped organic matters leads to the release of CO<sub>2</sub>, CH<sub>4</sub>, NH<sub>3</sub>, amines and H<sub>2</sub>S that significantly contributes in the climate changes. Also these solid byproducts negatively affect the aquatic life via alteration of the color and odor of the surrounding environment. Discharge of untreated effluents in the soil directly affect the inhabited microorganisms as it increases the moisture, salinity, carbon content and electric conductivity. Moreover, high levels of nitrogen, phosphorus, fat, oil and grease may lead to drinking water shortage, eutophication, biotic depletion, algal blooms, acidification of water, destruction of habitats, outbreaks of various diseases and corals siltation. Therefore, dealing with this huge amount of the produced wastes is a great challenge [4].

Solid byproducts including skin, viscera, bones, heads and other corporal structures reach about 65% of the total weight while liquid byproducts are the effluents produced during washing, cooking and thawing processes [5]. These byproducts are

\*Corresponding author e-mail: <a href="mailto:shaymaaabdallaismail@gmail.com">shaymaaabdallaismail@gmail.com</a>

Receive Date: 06 July 2022, Revise Date: 26 August 2022, Accept Date: 27 August 2022

DOI: 10.21608/EJCHEM.2022.149433.6457

<sup>©2022</sup> National Information and Documentation Center (NIDOC)

considered as an undiscovered treasure having the potentiality for the production of various biomaterials possessing several functional and biological activities. The valorization of these byproducts and their utilization in other industries is one of the main avenues that can close the loop in seafood production [6-10].

The problem of discarding the seafood byproducts is an emerging issue that indirectly increases the pressure on fisheries enforcing the search of an ecofriendly applicable solution to solve this problem via the adaptation of a circular economy based on business models [1]. Table (1) illustrated the negative and the positive issues that inherent the problem of seafood byproducts.

Table (1): The negative and the positive issue that inherent the problem of seafood byproducts (modified from Caruso *et al.*, [6]).

Negative issue	Positive issue
Environmental impact	The utilization of these
	byproducts contributes in
	their safe disposal.
Loss of nutrients	These nutrients can be
	exploited in nutritional,
	pharmaceutical, industrial
	sectors (i.e., biorefinery).
Operational cost of its	Biotechnological studies
exploitation	can enhance the
	operational quality as
	well as its efficiency.

# Solid byproducts

Fish composition is largely varied depending on several variables including the species, state of nutrition, health, age and season of collection [11]. The average percentage of the discarded seafood parts is illustrated in table (2).

Table (2): Average weight percentage of the discarded seafood parts.

Туре	Discarded part	Average weight percentage (%)	Reference
Finfish	Head	21	Caruso et
	Bones	14	al., [6]
	Fins	10	
	Gut	7	
	Liver	5	
	Skin	3	
	Ovaries	4	
Shellfish	Shell and	35-45	Suryawanshi
	head		<i>et al.</i> , [12]

#### Source for probiotics

Probiotics are defined according to the Food and Agriculture Organization of the United Nations and World Health Organization [13] as "live microorganisms which when administered in adequate amounts confer a health benefit on the host" via the direct or the indirect influence on the composition of the intestinal microbiota. Fish gastrointestinal tract, gills and skin microbiota represent a rich source of probiotic bacteria [14] in which their isolation and their synthesized bioactive compounds attract the research interest. Different activities including antimicrobial [15-16] and antitumor [17] have been reported for various biomolecules produced by probiotics isolated from fish organs. Additionally, Floris et al., [16] estimated seabream intestinal microflora gilthead as biosurfactant producers with antimicrobial activity. Microbial surfactants produced by marine microorganisms have attracted a great interest in the 21<sup>st</sup> century as they possess multiple therapeutic functions including antifouling, anti-adhesive, antimicrobial, antithrombotic and antitumor [18]. The strains that have been isolated from fish organs are summarized in table (3).

#### Fishmeal production

Seafood byproducts are rich in nutritionally valuable ingredients with average values illustrated in table (4). In the last two decades, the utilization of fishery by-catch or processing byproducts as a fishmeal (prepared by drying followed by grinding) participating in the production of feed for aquaculture, poultry, ruminants and pets had been examined and well established [19-20].

Table (3): The microbial strains isolated from fish organs.

Gram positive strain					
Lactobacillu	Lactobacillus	Lactobacillus	Enterococcus		
S	pentosus	delbrueckii	faecium		
fructivorans	-				
Bacillus sp.	Paenibacillus	Micrococcus	Macrococcus		
	sp.	sp.	sp.		
Leuconostoc	Brochothrix	Carnobacteriu	Arthrobacter		
sp.	sp	<i>m</i> sp.	arilaitensis		
Anoxybacillu	Staphylococc	Actinobacteri			
<i>s</i> sp.	us sp.	а			
	Gram neg	ative strain			
Citrobacter	Plesiomonas	Enterobacter	Shewanella		
freundii	shigelloides	sp.	xiamenensis		
Hafnia alvei	Vibrio sp.	Aeromonas	Psychrobacte		
		sp.	<i>r</i> sp		
Agrobacteriu	Photobacteriu	Acinetobacter	Azospirillum		
<i>m</i> sp.	<i>m</i> sp.	sp.	orizae		
Erwinia	Sphingomona	Pseudomonas	Cyanobacteri		
persicina	<i>s</i> sp.	sp.	а		
Yeast					
Candida sp.	Saccaromyces	Debaryomyce	Leucosporidiu		
	cerevisiae	s hansenii	<i>m</i> sp.		
Kodamea	Pichia sp.	Rhodotorula			
ohmeri		sp.			

viold

discards [4].	
Nutrient	Average percentage
	(%)
Crude protein	$57.9 \pm 5.3$
Fat	$19.1 \pm 6.1$
Crude fiber	$1.2 \pm 1.2$
Ash	$21.8 \pm 3.5$
Sodium	$0.6 \pm 0.1$
Potassium	$0.7 \pm 0.1$
Phosphorous	$2.0 \pm 0.6$
Calcium	5.8 ± 1.3

 Table (4): Average composition of seafood processing discards [4].

#### Source for their constitutive compounds

In the last few years, the extraction of seafood derived compounds and their applications in various fields of life including food, agricultural and medical sectors have attracted growing interests [4]. There are various compounds that can be extracted from either finfish or shellfish byproducts and can be classified under four main groups including nitrogenous compounds, lipids, polysaccharides and minerals.

#### Nitrogenous compounds

Diverse proteins, peptides and amino acids are the main constitutive components of the nitrogenous fraction possessing valuable nutritional and functional properties including emulsifying, foaming and texture improving agents in addition to possessing various biological activities including antioxidant and antimicrobial activities [20]. In general, fish proteins are easy digestible with high nutritional value. They are composed of wellbalanced composition of essential amino acids as valine, lysine and phenylalanine [22].

#### • Proteins

The increasing demands for protein either for meeting the human or animal requirements enforces the research interest for its recovery from novel sources [23]. Seafood solid byproducts consist of about 60% proteins with high nutritional value (containing essential amino acids) that can be recovered retaining its native properties or converted to amino acid and polypeptides [4]. Isoelectric solubilization precipitation is a mild technique in which dilute acid (pH 2.5-3.5) or dilute alkali (pH 10.8-11.5) are used in the homogenization and solubilization of seafood byproducts followed by precipitation and filtration of the dissolved proteins. It has been efficiently used in the recovery of sarcoplasmic and myofilbrillar proteins from different byproducts of various species of finfish and shellfish with up to 95% recovery yield (Table 5) [20].

yiciu.			
Source	Extraction	Yield	Reference
	method	(%)	
Tilapia	Alkaline	68	Chomnawang
frame			Yongsawatdigul,
			[24]
Bighead	Acidic	74.8	Chang et al., [25]
carp			
Common	Acidic	76.3	Tian <i>et al.</i> , [26]
carp			
	Alkaline	87.6	
Mackerel	sequential	95–	Álvarez et al.,
	acid/ alkaline	100	[27]
Pangas	Acidic	59	Surasani et al.,
processing			[28]
waste	Alkaline	69	
Catfish	Alkaline	36–55	Tan et al., [29]
heads			
	Alkaline	53	
Catfish			
frames			
Green crab	Alkaline	83	Khiari et al., [30]

Table (5): Seafood byproducts and protein recovery

Apart from myofibrillar protein, marine collagen represents a biocompatible alternative to the mammalian one. In general, collagen compromises 30% of the total protein exists in most organisms and it compromises 70% of the skin dry weight. Seafood byproducts are a collagen-rich source possessing various nutraceutical and functional properties [31-32]. Melgosa *et al.*, [33] reported the preparation of collagen-rich protein extract from cod frames possessing anti-inflammatory activity. In addition, fish scales collagen is a natural biomaterial find application in various fields including drug delivery [34], wound healing [35], corneal tissue engineering [36] and oral mucosa regeneration [37].

Gelatin is a more soluble protein that structurally constructed connective tissues. Dara *et al.*, [38] reported that the gelatin extracted from big eye tuna skin was suitable in nutraceutical and biomedical applications as it possessed good gelling properties. Valcarcel *et al.*, [39] indicated seabream skin byproducts as suitable raw materials for the production of gelatin.

#### • Enzymes

Enzymes are highly specific protein molecules that catalyze various biochemical reactions providing ecobenign, easy controllable and efficient processes. Viscera of fish are rich in enzymes including proteases (pepsin, trypsin, trypsin-like enzymes, collagenase, peptidase and elastase), chitinase, lipase and others. The isolated enzymes mainly exhibit high thermal activity and stability with high stability at wide pH range that make them suitable candidates in various food, detergent and pharmaceutical industries [40-42]. The isolated enzymes and their potential applications are illustrated in table (6).

Table (6): Isolated enzymes and their potentialapplications [20].

	Enzyme	Source	Application
Ī	Proteases	Fish	Recovery of
		intestines and	seafood
		shrimp heads	byproducts
			constituents and
			production of fish
			protein
			hydrolysate
	Lipases	Fish discards	Production of
			omega-3-enriched
			triglycerides
	Chitinases	Shellfish	Production of
l			chitin hydrolysis
ļ			products
	Lysozyme	Arctic scallop	Bacteriostatic
		shell, crab	agent
ì		shell	
	Catalase,	Marine	Antioxidants
	glutathione	mussel	
ļ	peroxidase		<i>a</i>
	5'-	Fishery	Construction of
	nucleotidase,	byproducts	biosensors to
	Nucleoside		measure amines,
	phosphorylase		nucleotides, and
			others

# Lipids and co-products

# • Oil production

Fishery byproducts contain a varied amount of lipids (up to 30%) that present in the form of fish oil. The oil extracted from fish byproducts is a good quality one that can be exploited in pharmaceutical and food industries. It contains two main polyunsaturated fatty acids, eicosapentaenoic acid and docosahexaenoic acid that are classified as omega-3 fatty acids [43]. Bio-oils gained a global interest for either their lonely use or in blend with petroleum fuel. The use of biomasses in the production of bio-fuels has attracted the research focus from the prospective of their valorization and environmental protection. Fish oil produced from seafood byproducts can be considered as a convenient source for fuel production [44-45].

#### • Carotenoids

They are red orange pigments present in crustaceans as well as salmon and trout. They are either hydrocarbon in nature as  $\beta$ -carotene and xanthophylls or oxygenated derivatives as astaxanthin and canthaxanthin. Crustacean shells are an important source of natural astaxanthin that has been reported as potent antioxidant [46]. In general, antioxidants are compounds that can be used to

overcome the deleterious effects of free radicals in the biological systems [47]. Solvent extraction is the commonly applied method for the extraction of astaxanthin from crustaceans. Recently, the searching for a new eco-friendly sustainable technique as well as adjuvant treatments for the extraction process has attracted the research focus for example; microbial fermentation [48], ultrasound-assistance [49], enzymatic treatment [50] and microwave pretreatment [51].

#### Polysaccharides

Crustacean shell is the primary source of chitin, linear polysaccharide of N-acetyl-D-glucosamine units linked by  $\beta$ -1,4-glycosidic bonds [52], prepared manipulation steps including via several deproteinization, demineralization and discoloration with a recovery yield of about 25% [53]. Chitin has attracted a growing interest due to its various food, agricultural and pharmaceutical applications in addition to its applicability in the production of valuable products including chitosan (de-acetylated derivative). N-acetyl glucosamine, chitooligosaccharide and various biologically active chitinolytic enzymes [54]. Chemical extraction is the commercial process that still applied for the preparation of chitin [55] but the development of green extraction processes gained more attention [56-57] preferring the use of enzymes and acid producing bacteria since the biological process produce product of better quality under mild and economic conditions [58-59] (Figure 1).



Figure (1): Recovery of chitin and its application.

Glycosaminoglycans are linear polysaccharides composed of disaccharide repeating units of amino sugars covalently linked to uronic acid. In the last two decade, the researchers investigate their potential applications in which they have been reported to possess various structural and functional properties including antitumor, anticoagulant and antiinflamatory activities in addition to their applicability in tissue engineering [60-61]. Among them, hyalouronic acid and chondroitin sulphate are the main groups that have been extracted from seafood byproducts [62]. Murado *et al.*, [63] reported the extraction of hyalouronic acid from fish eyeball and Vazquez *et al.*, [64] reported the extraction of chondroitin sulphate from the cartilage of blackmouth catfish.

# Minerals

Bone composed of about 70% minerals so bone-rich byproducts are a significant source of minerals. Tang *et al.*, [65] reported the production of fermentable solution rich in calcium salts by the fermentation of grass fish bones using *Leuconostoc mesenteroides* suggesting its applicability as calcium supplement.

Hydroxyapatite is an inorganic material widely distributed in hard tissues for supporting their structure [66]. It has been reported to be applied in some biomedical field including bone tissue engineering, periodontal repair and dyes biosorbant [9]. Fish scales are efficient byproducts used for its preparation [67-68].

# Conversion of the constitutive components

Protein hydrolysates, chitin based products and enzymes are the major value added products that resulted from the conversion of the components of seafood byproducts. The use of microorganisms or enzymes is the main employed conversion process.

# Major conversion techniques

# • Microbe-mediated conversion

The use of microorganisms in the conversion processes is named as fermentation in which several products have been produced. In the fermentation process, the cultural and the nutritional conditions are crucial variables that influence the growth of the microorganisms as well as their released metabolic products. In the last two decade, statistical models have been widely employed in fermentation technology to adjust its condition for optimizing the productivity of the desired product [69]. Response surface methodology that described by Box and Wilson, [70] as well as artificial neural networks are the most popular mathematically based techniques that has been applied in the optimization processes [71]. The use of microorganisms in the fermentation of seafood byproducts is estimated as an efficient technique for their bioremediation that resulted in the production of valuable products including enzymes, antioxidant compounds, protein hydrolysate and others [72]. Enzymes, liquid fertilizers, glutamic acids, pigments and biologically active oligosaccharides are the main valuable products produced by the microbial conversion of seafood byproducts (Table 7).

• Enzyme- mediated conversion

In general, hydrolases are the most famous group of enzymes used in biotechnological applications. Specifically, proteases are widely applied in the conversion of seafood byproducts [31]. In addition, glycoside hydrolases and lipases give rise to various biologically active hydrolysates by the conversion of seafood constitutive ingredients [83-84]. Proteins, pigment, chitin, chitooligosaccharides and deodorized oil are the main products produced by the enzymatic conversion of some seafood byproducts (Table 8).

# Major conversion products

## • Protein hydrolysate and bioactive peptides

The protein content in seafood byproduct is an efficient source for the production of peptide-rich hydrolysates possessing various functional properties including emulsifying, foaming, rheological, textural and physical properties in addition to various biological activities including antimicrobial, antioxidant, anticancer, antidiabetic, anticoagulant and antihypertensive in addition to hepato and cardio protective agents [91].

# • Chitin based products

Chitosan is the de-acetylated derivative of chitin with the presence of less than 20% N-acetyl-Dglucosamine units. It is a non-toxic biopolymer extensively applied in various fields including biomedical, pharmaceutical, agricultural, food and feed industries in addition to possess various biological activities including anti-inflammatory, immune-modulatory [92], antitumor [93] and antimicrobial [94].

N-acetyl glucosamine (chitin monoconstituent sugar) is a clinical drug for the treatment of rheumatoid arthritis. In addition, it possesses antimicrobial, antioxidant and anticancer activities with potential food, agricultural, medical and pharmaceutical applications [95]. Currently, the conversion of crustacean byproducts to N-acetyl glucosamine attracts a growing interest as it can be exploited in the production of bioethanol [10].

Chitooligosachharides are water soluble homo- or hetero-oligomers of D-glucosamine and Nacetyl-D-glucosamine with an average molecular weight less than 3900Da [96]. They have been estimated to possess various biological activities including prebiotic, antioxidant [82, 97], antitumor [98], neuroprotective [99-100], antifungal [101-102], antibacterial [103], immuno-modulatory [92], hepatoprotective [104] and hypolipidemic effect [105].

# • Enzymes

Nowadays, green chemistry is attracting a great interest. Industrial enzymes have been considered as a green route for protecting the environment and popular health but the cost is a significant barrier that restricted its application. The reduction of the cost via the use of seafood byproducts as a substrate for the production of enzymes instead of using the refined one is attracting the research focus [54].

Proteases are the most popular class of various that widely applied in enzymes biotechnological processes including amino acid analysis, detergent, cosmetics, food and feed production [106]. Proteases in general are a complex group of enzymes that specifically catalyze the hydrolysis of proteins converting them to peptide chains and/or amino acids [107]. Ramkumar et al., [77] reported the use of fish gut waste as an efficient substrate for the microbial production of protease using Bacillus licheniformis.

Shrimp byproducts have also been reported as an efficient substrate for inducing the microbial production of various chitinolytic [54] and chitosanolytic [81] enzymes as well as chitin deacetylase [108]. Chitinases and chitosanases are chitin and chitosan specific hydrolytic enzymes that lead to the production of their monomer constituents as well as chitooligosaccharides while chitin deacetylases are the enzymes responsible for the deacetylation of chitin for chitosan production [109].

#### Liquid byproduct

The operations carried out during the processing of seafood result in the production of wastewater or effluents reach in soluble organic matters, salts and colloidal substances with high content of Chemical Oxygen Demand generated mainly from biodegradable lipids and proteins [110]. A variation in the volume of wastewater results from different processing operations was estimated and the average volumes are illustrated in table (9).

Table	e (9):	The	average	volume	of	wastewater	results
from	differ	ent s	eafood pr	ocessing	op	erations.	

Operating process	Volume of wastewater (m <sup>3</sup> /ton raw material)	Reference
Precooking of fish to be	0.07-0.27	Arvanitoyannis
canned		and Kassaveti,
Unloading fish for canning	2-5	[111]
Sterilization of cans	3-7	
Handling and storage of fish	10-12	
Scaling of white fish	10-15	
Oily fish skinning	0.2-0.9	
Marine finfish	14.0	
Frozen fish thawing	5.0	
Shrimp freezing	7.0	
Blue crab, mechanized plant	29-44	
Processing of tuna	3.0	Fluence, [112]
Canning of sardine	15.0	Venugopal and
White fish filleting	5-11	Sasidharan,
Oily fish filleting	5-8	[113]
Skinning of knobbed fish	17.0	
Filleting of un-gutted oily fish	1-2	

Egypt. J. Chem. 65, No. SI13B (2022)

#### Algae based bioconversion

Microalgae is one of the resources that attracted the research focus for its economic biomass production as it can incorporate in the production of biofuels, animal feed, pharmaceutical and health products [114]. Gao et al., [115] indicated the feasibility of the production of algal biomass (*Chlorella* sp.) using seafood processing wastewater.

#### Source of biologically active compounds

Seafood liquid byproducts (cooking juice and stickwater) can be used as a source for the production of various bio-molecules including nitrogenous compounds, carotenoids, lipids and flavors that can be exploited in several biotechnological processes.

#### • Cooking juice

Cooking juice or cooking wastewater is the effluent resulted from fish cooking operations performed mainly during canning processes. Tuna or small pelagic fish (sardine and anchovy) are the main traditional raw material. Hsu *et al.*, [116] reported that the yield of cooking juice (with about 4% of water-soluble protein) produced every day was in the range of 15 to 27 ton for each fish canning plant. Tang *et al.*, [117] reported the production of 1.5 ton of cooking wastewater for each ton of processed anchovy containing 5g/L of crude protein in addition to essential amino acids. Additionally, P'erez-Santín *et al.*, [118] reported that the industrial shrimp cooking juice contains 13.5% protein.

#### • Stickwater

Stickwater is the effluent resulted during the preparation of fishmeal. It represents 60% of the fish weight and composed of 5-9% protein content [119].

#### Nitrogenous compounds

Seafood effluents contain a considerable amount of soluble proteins that can be recovered and concentrated or used for the production of protein hydrolysates containing biologically active peptides.

#### Production of bioactive peptides

#### Enzymatic hydrolysis

The hydrolysis of protein-reach seafood byproducts leads to the production of peptides varied in their size as well as their composition and consequently their bioactivity [120]. The selection of the hydrolysis conditions is the crucial step in the application of enzymes in which the enzymesubstrate ratio, hydrolysis period and the temperature of the reaction have been reported as the main influencing variables [121]. The application of proteolytic enzymes in the hydrolysis of seafood liquid byproducts has been previously studied manifesting its applicability as a valuable source for the production of bioactive peptides. P'erez-Santín et al., [118] reported the use of proteolytic alcalase in the hydrolysis of shrimp cooking juice and production of bioactive peptides. Hung et al., [122] reported the production of biologically active protein hydrolysate from the hydrolysis of cooking juice of the industrial manufacturing of tuna by applying ultrafilteration. protease followed by The combination of enzymatic hydrolysis and ultrafilteration had also been reported for the production of protein hydrolysate results from the hydrolysis of shrimp cooking water [123]. Mahdabi and Shekarabi, [124] reported the production of protein hydrolysate by the enzymatic hydrolysis of stickwater resulted from the preparation of kilka fishmeal using alcalase.

# • Membrane technology

The use of semi-permeable membranes in the separation of valuable molecules from seafood liquid byproducts is one of the efficient techniques that have been applied in the preparation of biologically active peptides. It possesses several advantages as it minimizes the denaturation of protein and it can be utilized for obtaining specific molecular weight peptides [125].

# Functional and biological activities

The recovered protein as well as the produced peptides can be widely applied depending on the base of their structural features including molecular weight, amino acid composition, sequence and hydrophobicity.

# • Emulsifying and foaming agent

The protein recovered from herring industry wastewater with a molecular weight of 50 KDa can be used as a natural emulsifying agent [126]. In addition, Gringer *et al.*, [127] indicated that the foaming as well as the emulsifying property of the proteins recovered by ultra-filtration of herring industry wastewater was not affected.

# • Antioxidant activity

Antioxidant peptides have been previously prepared from tuna cooking juice [116], shrimp cooking juice [118], herring industry wastewater [126] and from kilka stickwater [124]. Hsu *et al.*, [116] attributed the antioxidant activity of the produced peptides to the presence of proton donating amino acids, histidine and proline. Additionally, Tremblay *et al.*, [128] indicated that the cooking effluent of snow crab was composed of 59% protein that possessed antioxidant activity.

## • Antihypertensive activity

The peptides prepared by the enzymatic hydrolysis of the protein recovered by ultrafilteration from cuttlefish processing wastewater had been reported to possess antihypertensive activity [129].

# • Antiproliferative activity

Mutations in general encourage carcinogenesis proliferation of and cells. Antiproliferative activity of peptides prepared from tuna cooking juice had been estimated against breast cancer cell line (MCF-7) without any cytotoxic activity against mammary epithelial cells [122]. The molecular weight of the produced peptide was greater than 2.5 KDa and composed mainly of hydrophobic amino acids. Huang et al., [130] indicated that hydrophobic peptides could penetrate into the hydrophobic core of the cell membrane participating in antiproliferative activity. In addition, Hung et al., [122] attributed the antiproliferative activity of the produced peptide to the induction of the expression of caspase 3 that activated cancer cell apoptosis.

# Carotenoids

Shrimp cooking fluids can be used to isolate the carotenoid, astaxanthin that can exist freely or with esterified derivative possessing antioxidant activity [118].

# Lipids

Fish processing effluents contain considerable amount of lipids that can be isolated and exploited in various applications. Bechtel, [131] reported that pollock, cod and salmon stickwater contain variable amount of lipids and Garcia-Sifuentes *et al.*, [132] concentrated sardine stickwater in which the fat content reached 18%. Additionally, Alkaya and Demirer, [133] reported that gutting process water recycling system contains valuable fish oil/grease by-product and Monteiro *et al.*, [134] reported the application of High hydrostatic pressure for the extraction of polyunsaturated fatty acid from fish canning effluents.

# Flavor compounds

Flavor compounds are mainly used either to add aroma or taste. Aromatic compounds are usually volatile in nature with molecular weight less than 400Da as aldehydes and ketones while taste-adding compounds are mostly water soluble consisting of organic acids, amino acids and sugars [128]. Crustacean effluents including shrimp and crab are attractive proposition for the production of natural flavoring products [135, 128].

### **Bio-refinery approaches**

Seafood byproducts are a treasure rich with various valuable products that can be a promising renewable biomass for bio-refineries. The International Energy Agency defined bio-refinery as the "sustainable processing of biomass into a spectrum of bio-based products (food, feed, chemicals, and materials) and bio-energy [136]. Several bio-refinery approaches have been designed for the utilization of seafood byproducts as illustrated in table (10).

Table (10): Bio-refinery approaches for seafoodbyproducts.

<b>Bio-refinery</b>	Products	Refrences
Cultivation of alga	Astaxanthin, single cell protein (SCP)	Khoo <i>et al.</i> , [137]
	Various products	Mitra and Mishra, [138]
Lactic fermentation	Astaxanthin, hydrolyzed protein and chitin	Routray <i>et al.</i> , [78]
Anaerobic fermentation with cow dung	Methane, Liquid mineral fertilizer	Kratky and Zamazal, [139]
Coupled alcalase hydrolysis and bacterial fermentation	Gelatin, oils, FPH, bioactive peptides, and fish peptones	Vázquez <i>et al.</i> , [58]
Sequential extraction by ISP followed by enzyme	Collagen, myofibrillar proteins	Abdollahi <i>et</i> <i>al.</i> , [140]
Sequential treatment of crustacean shells	Chitin, proteins, lipids, carotenoids and CaCO3	Hülsey, [141]

#### Conclusion

In a world of stagnating oceanic resources and increasing of environmental problems, it is imperative that seafood processing needs to be ecofriendly and economic. Management of the produced byproducts can significantly help seafood industry realize objectives of food security and environmental protection. In this article, seafood byproducts have been estimated as a potential resource for the production of various biomaterials that find applications in food, agricultural and biomedical fields. These byproducts either solid discards including skin, viscera, bones, heads and other corporal structures or liquid effluents produced during washing, cooking and thawing processes are

Egypt. J. Chem. 65, No. SI13B (2022)

source for diverse proteins, peptides, amino acids, oils, pigments, polysaccharides and minerals that possess various nutritional, functional and biological activities. In addition, Fish gastrointestinal tract, gills and skin microbiota represent a rich source of probiotic bacteria capable for the production of various bioactive compounds. For more economic approaches, integrated refinery-type processes for the extraction of multiple products are widely applied.

#### References

- Ruiz-Salmón, I., Laso, J., Margallo, M., 1. Villanueva-Rey, Р., Rodríguez, Е., Quinteiro, P., Dias, A.C., Almeida, C., Nunes, M.L., Marques, A. and Cortés, A., Life cycle assessment of fish and seafood processed products-a review of methodologies and new challenges. Science of The Total Environment, 761, 144094 (2021).
- Han, D., Chen, Y., Zhang, C., Ren, Y., Xu, B. and Xue, Y., Evaluation of effects of shellfish aquaculture and capture fishery on a semi-closed bay ecosystem. *Estuarine*, *Coastal and Shelf Science*, 207, 175-182 (2018).
- Willer, D.F. and Aldridge, D.C., Microencapsulated diets to improve bivalve shellfish aquaculture for global food security. *Global Food Security*, 23, 64-73 (2019).
- Venugopal, V., &Sasidharan, A., Seafood industry effluents: environmental hazards, treatment and resource recovery. *Journal of Environmental Chemical Engineering*, 9(2), 104758 (2021).
- Ferraro, V., Carvalho, A.P., Piccirillo, C., Santos, M.M., Castro, P.M. and Pintado, M.E., Extraction of high added value biological compounds from sardine, sardinetype fish and mackerel canning residues—A review. *Materials Science and Engineering: C*, 33(6), 3111-3120 (2013).
- de la Caba, K., Guerrero, P., Trung, T.S., Cruz-Romero, M., Kerry, J.P., Fluhr, J., Maurer, M., Kruijssen, F., Albalat, A., Bunting, S. and Burt, S., 2019. From seafood waste to active seafood packaging: An emerging opportunity of the circular economy. *Journal of Cleaner Production*, 208, 86-98 (2019).
- Caruso, G., Floris, R., Serangeli, C. and Di Paola, L., Fishery wastes as a yet undiscovered treasure from the sea:

Biomolecules sources, extraction methods and valorization. *Marine Drugs*, **18**(12), 622 (2020).

- Suryawanshi, N. and Eswari, J.S., Shrimp shell waste as a potential raw material for biorefinery—a revisit. *Biomass Conversion and Biorefinery*, 1-8 (2021).
- Mahari, W.A.W., Waiho, K., Azwar, E., Fazhan, H., Peng, W., Ishak, S.D., Tabatabaei, M., Yek, P.N.Y., Almomani, F., Aghbashlo, M. and Lam, S.S., A state-ofthe-art review on producing engineered biochar from shellfish waste and its application in aquaculture wastewater treatment. *Chemosphere*, 288, 132559 (2022).
- Qin, D., Bi, S., You, X., Wang, M., Cong, X., Yuan, C., Yu, M., Cheng, X. and Chen, X.G., 2022. Development and application of fish scale wastes as versatile natural biomaterials. *Chemical Engineering Journal*, 428, 131102 (2022).
- Ghaly, A.E., Ramakrishnan, V.V., Brooks, M.S., Budge, S.M. and Dave, D., Fish processing wastes as a potential source of proteins. *Amino acids and oils: a critical review. Journal of Microbial and Biochemical Technology*, 5(4), 107-129 (2013).
- 12. Suryawanshi, N., Jujjavarapu, S.E. and Ayothiraman, S., Marine shell industrial wastes–an abundant source of chitin and its derivatives: constituents, pretreatment, fermentation, and pleiotropic applications-a revisit. *International journal of environmental science and technology*, 1-22 (2019).
- 13. FAO/WHO., Probiotics in food: Health and nutritional properties and guidelines for evaluation, 1–29 (2001).
- 14. Merrifield, D.L. and Rodiles, A., The fish microbiome and its interactions with mucosal tissues. In *Mucosal health in aquaculture*, 273-295 (2015).
- 15. Burbank, D.R., LaPatra, S.E., Fornshell, G. and Cain, K.D., Isolation of bacterial probiotic candidates from the gastrointestinal tract of rainbow trout, *Oncorhynchusmykiss* (W albaum), and screening for inhibitory activity against *Flavobacterium psychrophilum. Journal of Fish Diseases*, **35**(11), 809-816 (2012).
- Floris, R., Scanu, G., Fois, N., Rizzo, C., Malavenda, R., Spanò, N. and Lo Giudice, A., Intestinal bacterial flora of Mediterranean gilthead sea bream (*Sparusaurata Linnaeus*) as a novel source

of natural surface active compounds. *Aquaculture Research*, **49**(3), 1262-1273 (2018).

- Chen, J., Zhao, K.N. and Vitetta, L., Effects of intestinal microbial–elaborated butyrate on oncogenic signaling pathways. *Nutrients*, 11(5), 1026 (2019).
- Gudiña, E.J., Teixeira, J.A. and Rodrigues, L.R., Biosurfactants produced by marine microorganisms with therapeutic applications. *Marine drugs*, 14(2), 38 (2016).
- 19. Malaweera, B.O. and Wijesundara, W.N.M., Use of seafood processing by-products in the animal feed industry. In *Seafood Processing By-Products*, 315-339 (2014). Springer, New York, NY.
- 20. Afreen, M. and Ucak, I., Fish processing wastes used as feed ingredient for animal feed and aquaculture feed. *Survey in Fisheries Sciences*, **6**(2), 55-64 (2020).
- 21. Sasidharan, A. and Venugopal, V., Proteins and co-products from seafood processing discards: Their recovery, functional properties and applications. *Waste and Biomass Valorization*, **11**(11), 5647-5663 (2020).
- 22. Hayes, M. and Flower, D., Bioactive peptides from marine processing byproducts. *Bioactive compounds from marine foods: Plant and animal sources*, 57-71 (2013).
- Henchion, M., Hayes, M., Mullen, A.M., Fenelon, M. and Tiwari, B., Future protein supply and demand: strategies and factors influencing a sustainable equilibrium. *Foods*, 6(7), 53 (2017).
- 24. Chomnawang, C. and Yongsawatdigul, J., Protein recovery of tilapia frame byproducts by pH-shift method. *Journal of Aquatic Food Product Technology*, **22**(2), 112-120 (2013).
- Chang, T., Wang, C., Yang, H., Xiong, S., Liu, Y. and Liu, R., Effects of the acid-and alkali-aided processes on bighead carp (*Aristichthysnobilis*) muscle proteins. *International Journal of Food Properties*, 19(8), 1863-1873 (2016).
- 26. Tian, Y., Wang, W., Yuan, C., Zhang, L., Liu, J. and Liu, J., Nutritional and digestive properties of protein isolates extracted from the muscle of the common carp using pH-shift processing. *Journal of food processing and preservation*, **41**(1), e12847 (2017).
- 27. Álvarez, C., Lélu, P., Lynch, S.A. and Tiwari, B.K., Optimised protein recovery from mackerel whole fish by using

sequential acid/alkaline isoelectric solubilization precipitation (ISP) extraction assisted by ultrasound. *LWT*, **88**, 210-216 (2018).

- Surasani, V.K.R., Kudre, T. and Ballari, R.V., Recovery and characterization of proteins from pangas (*Pangasiuspangasius*) processing waste obtained through pH shift processing. *Environmental Science and Pollution Research*, **25**(12), 11987-11998 (2018).
- 29. Tan, Y., Gao, H., Chang, S.K., Bechtel, P.J. and Mahmoud, B.S., Comparative studies on the yield and characteristics of myofibrillar proteins from catfish heads and frames extracted by two methods for making surimi-like protein gel products. *Food chemistry*, **272**, 133-140 (2019).
- 30. Khiari, Z., Kelloway, S. and Mason, B., Turning Invasive Green Crab (Carcinusmaenas) into **Opportunity:** Recovery of Chitin and Protein Isolate Through Isoelectric Solubilization/Precipitation. Waste and Biomass Valorization, 11(1), pp.133-142 (2020).
- 31. Pal, G. K., & Suresh, P. V., Sustainable valorisation of seafood by-products: Recovery of collagen and development of collagen-based novel functional food ingredients. *Innovative food science & emerging technologies*, **37**, 201-215 (2016).
- Raman, M. and Gopakumar, K., Fish collagen and its applications in food and pharmaceutical industry: a review. *EC Nutr*, 13(12), 752-67 (2018).
- Melgosa, R., Marques, M., Paiva, A., Bernardo, A., Fernández, N., Sá-Nogueira, I., &Simões, P., Subcritical Water Extraction and Hydrolysis of Cod (*Gadusmorhua*) Frames to Produce Bioactive Protein Extracts. *Foods*, 10(6), 1222 (2021).
- 34. Pathan, I.B., Munde, S.J., Shelke, S., Ambekar, W. and MallikarjunaSetty, C., Curcumin loaded fish scale collagen-HPMC nanogel for wound healing application: Exvivo and In-vivo evaluation. *International Journal of Polymeric Materials and Polymeric Biomaterials*, 68(4), pp.165-174 (2019).
- 35. Shalaby, M., Agwa, M., Saeed, H., Khedr, S.M., Morsy, O. and El-Demellawy, M.A., Fish scale collagen preparation, characterization and its application in wound healing. *Journal of Polymers and the Environment*, 28(1), 166-178 (2020).

- 36. Hsueh, Y.J., Ma, D.H.K., Ma, K.S.C., Wang, T.K., Chou, C.H., Lin, C.C., Huang, M.C., Luo, L.J., Lai, J.Y. and Chen, H.C., Extracellular matrix protein coating of processed fish scales improves human corneal endothelial cell adhesion and proliferation. *Translational vision science & technology*, 8(3), 27-27 (2019).
- 37. Suzuki, A., Kato, H., Kawakami, T., Kodama, Y., Shiozawa, M., Kuwae, H., Miwa, K., Hoshikawa, E., Haga, K., Shiomi, A. and Uenoyama, A., Development of microstructured fish scale collagen scaffolds to manufacture a tissue-engineered oral mucosa equivalent. *Journal of Biomaterials Science, Polymer Edition*, **31**(5), 578-600 (2020).
- 38. Dara, P. K., Raghavankutty, M., Sebastian, N., Chatterjee, N. S., Mathew, S., Ravishankar, C. N., &Anandan, R., Rheological, physico-chemical, and surfaceactive properties of gelatin extracted from bigeye tuna (*Thunnusobesus*) skin waste. *Journal of Aquatic Food Product Technology*, **29**(5), 428-444 (2020).
- Valcarcel, J., Hermida-Merino, C., Piñeiro, M. M., Hermida-Merino, D., &Vázquez, J. A., Extraction and Characterization of Gelatin from Skin By-Products of Seabream, Seabass and Rainbow Trout Reared in Aquaculture. *International Journal of Molecular Sciences*, 22(22), 12104 (2021).
- 40. Venugopal, V., Enzymes from seafood processing waste and their applications in seafood processing. *Advances in food and nutrition research*, **78**, 47-69 (2016).
- Mardina, V., Fitriani, F., Harmawan, T., & Hildayani, G. M., Valorisasi Pankreas Ikan Tongkol (*EutynnusAffinis*) Untuk Produksi Enzim Lipase. *Elkawnie: Journal of Islamic Science and Technology*, 4(2), 189-197 (2018).
- Murthy, L. N., Phadke, G. G., Unnikrishnan, P., Annamalai, J., Joshy, C. G., Zynudheen, A. A., & Ravishankar, C. N., Valorization of fish viscera for crude proteases production and its use in bioactive protein hydrolysate preparation. *Waste and Biomass Valorization*, 9(10), 1735-1746 (2018).
- Šimat, V., Vlahović, J., Soldo, B., Mekinić, I. G., Čagalj, M., Hamed, I., &Skroza, D., Production and characterization of crude oils from seafood processing by-products. *Food Bioscience*, 33, 100484 (2020).

- 44. Adeoti, I. A., &Hawboldt, K., A review of lipid extraction from fish processing by-product for use as a biofuel. *Biomass and Bioenergy*, **63**, 330-340 (2014).
- 45. de Medeiros, E. F., Vieira, B. M., de Pereira, C. M. P., Nadaleti, W. C., Quadro, M. S., &Andreazza, R., Production of biodiesel using oil obtained from fish processing residue by conventional methods assisted by ultrasonic waves: Heating and stirring. *Renewable Energy*, **143**, 1357-1365 (2019).
- 46. Pereira, C. P. M., Souza, A. C. R., Vasconcelos, A. R., & Prado, P. S., Antioxidant and anti-inflammatory mechanisms of action of astaxanthin in cardiovascular diseases. *International Journal of Molecular Medicine*, **47**(1), 37-48 (2021).
- Aliakbarlu, J., Mohammadi, S., &Khalili, S., A Study on Antioxidant Potency and Antibacterial Activity of Water Extracts of Some Spices Widely Consumed in Iranian Diet. *Journal of Food Biochemistry*, 38(2), 159-166 (2014).
- 48. El-Bialy, H. A. A., &Abd El-Khalek, H. H., A comparative study on astaxanthin recovery from shrimp wastes using lactic fermentation and green solvents: An applied model on minced Tilapia. *Journal of Radiation Research and Applied Sciences*, 13(1), 594-605 (2020).
- Gao, J., You, J., Kang, J., Nie, F., Ji, H., & Liu, S., Recovery of astaxanthin from shrimp (*Penaeusvannamei*) waste by ultrasonic-assisted extraction using ionic liquid-in-water microemulsions. *Food chemistry*, **325**, 126850 (2020).
- Wang, L., Hu, J., Lv, W., Lu, W., Pei, D., Lv, Y., ... &Lv, M., Optimized extraction of astaxanthin from shrimp shells treated by biological enzyme and its separation and purification using macroporous resin. *Food Chemistry*, 363, 130369 (2021).
- 51. Nunes, A. N., Roda, A., Gouveia, L. F., Fernández, N., Bronze, M. R., &Matias, A. A., Astaxanthin extraction from marine crustacean waste streams: An integrate approach between microwaves and supercritical fluids. ACS Sustainable Chemistry & Engineering, 9(8), 3050-3059 (2021).
- 52. Blaak, H., Schnellmann, J., Walter, S., Henrissat, B., & Schrempf, H., Characteristics of an exochitinase from

*Streptomyces olivaceoviridis*, its corresponding gene, putative protein domains and relationship to other chitinases. *European journal of biochemistry*, **214**(3), 659-669 (1993).

- 53. Shahidi, F., Arachchi, J. K. V., &Jeon, Y. J., Food applications of chitin and chitosans. *Trends in food science & technology*, **10**(2), 37-51 (1999).
- 54. Hassan, A. A., & Ismail, S. A., Production of antifungal N-acetyl-β-glucosaminidase chitinolytic enzyme using shrimp byproducts. *Biocatalysis and Agricultural Biotechnology*, **34**, 102027 (2021).
- 55. Mohan, K., Muralisankar, T., Jayakumar, R., & Rajeevgandhi, C., A study on structural comparisons of α-chitin extracted from marine crustacean shell waste. *Carbohydrate Polymer Technologies and Applications*, 2, 100037 (2021).
- Huang, W. C., Zhao, D., Guo, N., Xue, C., & Mao, X., Green and facile production of chitin from crustacean shells using a natural deep eutectic solvent. *Journal of agricultural and food chemistry*, **66**(45), 11897-11901 (2018).
- Bradić, B., Novak, U., &Likozar, B., Crustacean shell bio-refining to chitin by natural deep eutectic solvents. *Green Processing and Synthesis*, 9(1), 13-25 (2020).
- 58. Vázquez, J. A., Meduíña, A., Durán, A. I., Nogueira, M., Fernández-Compás, A., Pérez-Martín, R. I., & Rodríguez-Amado, I., Production of valuable compounds and bioactive metabolites from by-products of fish discards using chemical processing, enzymatic hydrolysis, and bacterial fermentation. *Marine drugs*, **17**(3), 139 (2019).
- Yadav, M., Goswami, P., Paritosh, K., Kumar, M., Pareek, N., & Vivekanand, V., Seafood waste: a source for preparation of commercially employable chitin/chitosan materials. *Bioresources and Bioprocessing*, 6(1), 1-20 (2019).
- Kovensky, J., Grand, E., & Uhrig, M. L., Applications of glycosaminoglycans in the medical, veterinary, pharmaceutical, and cosmetic fields. In *Industrial Applications of Renewable Biomass Products*, 135-164 (2017). Springer, Cham.
- 61. Morla, S. Glycosaminoglycans and glycosaminoglycan mimetics in cancer and

Egypt. J. Chem. 65, No. SI13B (2022)

inflammation. *International journal of molecular sciences*, **20**(8), 1963 (2019).

- Abdallah, M. M., Fernández, N., Matias, A. A., & do Rosário Bronze, M., Hyaluronic acid and Chondroitin sulfate from marine and terrestrial sources: Extraction and purification methods. *Carbohydrate Polymers*, 243, 116441 (2020).
- Murado, M. A., Montemayor, M. I., Cabo, M. L., Vázquez, J. A., & González, M. P., Optimization of extraction and purification process of hyaluronic acid from fish eyeball. Food and Bioproducts Processing, **90**(3), 491-498 (2012).
- Vázquez, J. A., Fraguas, J., Novoa-Carvallal, R., Reis, R. L., Antelo, L. T., Pérez-Martín, R. I., &Valcarcel, J., Isolation and chemical characterization of chondroitin sulfate from cartilage by-products of blackmouthcatshark (*Galeusmelastomus*). *Marine drugs*, **16**(10), 344 (2018).
- 65. Tang, S., Dong, S., Chen, M., Gao, R., Chen, S., Zhao, Y., ... & Sun, B., Preparation of a fermentation solution of grass fish bones and its calcium bioavailability in rats. *Food & function*, 9(8), 4135-4142 (2018).
- Dey, A., Bomans, P. H., Müller, F. A., Will, J., Frederik, P. M., de With, G., &Sommerdijk, N. A., The role of prenucleation clusters in surface-induced calcium phosphate crystallization. *Nature materials*, 9(12), 1010-1014 (2010).
- 67. Deb, P., & Deoghare, A. B., Effect of Acid, Alkali and Alkali–Acid Treatment on Physicochemical and Bioactive Properties of Hydroxyapatite Derived from CatlaFish Scales. Arabian Journal for Science & Engineering (Springer Science & Business Media BV), 44(9) (2019).
- Sathiskumar, S., Vanaraj, S., Sabarinathan, D., Bharath, S., Sivarasan, G., Arulmani, S., ... & Ponnusamy, V. K., Green synthesis of biocompatible nanostructured hydroxyapatite from Cirrhinusmrigala fish scale–A biowaste to biomaterial. *Ceramics International*, 45(6), 7804-7810 (2019).
- Desai, K. M., Survase, S. A., Saudagar, P. S., Lele, S. S., &Singhal, R. S., Comparison of artificial neural network (ANN) and response surface methodology (RSM) in fermentation media optimization: case study of fermentative production of scleroglucan. *Biochemical Engineering Journal*, **41**(3), 266-273 (2008).

- 70. Box GE, and Wilson KB., On the experimental attainment of optimum conditions. *J Royal Statistical Society: Series B (Methodological)*, **13**(1), 1-38 (1951).
- 71. Astray, G., Gullón, B., Labidi, J., &Gullón, P., Comparison between developed models using response surface methodology (RSM) and artificial neural networks (ANNs) with the purpose to optimize oligosaccharide mixtures production from sugar beet pulp. *Industrial Crops and Products*, **92**, 290-299 (2016).
- 72. Marti-Quijal, F. J., Remize, F., Meca, G., Ferrer, E., Ruiz, M. J., &Barba, F. J., Fermentation in fish and by-products processing: An overview of current research and future prospects. *Current Opinion in Food Science*, **31**, 9-16 (2020).
- Esakkiraj, P., Dhas, G. A. J., Palavesam, A., & Immanuel, G., Media preparation using tuna-processing wastes for improved lipase production by shrimp gut isolate *Staphylococcus epidermidis* CMST Pi 2. *Applied biochemistry and biotechnology*, 160(4), 1254-1265 (2010).
- 74. Sanchart, C., Watthanasakphuban, N., Boonseng, O., Nguyen, T. H., Haltrich, D., &Maneerat, S., Tuna condensate as a promising low-cost substrate for glutamic acid and GABA formation using *Candida rugosa* and *Lactobacillus futsaii. Process biochemistry*, **70**, 29-35 (2018).
- 75. Dao, Y. T., and Kim, J. K. Scaled-up bioconversion of fish waste to liquid fertilizer using a 5 L ribbon-type reactor. *Journal of environmental management*, **92**(10), 2441-2446 (2011).
- 76. Vázquez, J. A., Durán, A. I., Menduíña, A., Nogueira, M., Gomes, A. M., Antunes, J., ... &Valcarcel, J., Bioconversion of fish discards through the production of lactic acid bacteria and metabolites: sustainable application of fish peptones in nutritive fermentation media. *Foods*, **9**(9), 1239 (2020).
- 77. Ramkumar, A., Sivakumar, N., Gujarathi, A. М., & Victor, R., Production of thermotolerant, detergent stable alkaline gut protease using the waste of Sardinellalongiceps as а substrate: Optimization and characterization. Scientific reports, 8(1), 1-15 (2018).
- 78. Routray, W., Dave, D., Cheema, S. K., Ramakrishnan, V. V., &Pohling, J., 2019.

Egypt. J. Chem. 65, No. SI13B (2022)

Biorefinery approach and environmentfriendly extraction for sustainable production of astaxanthin from marine wastes. *Critical reviews in biotechnology*, **39**(4), 469-488 (2019).

- Wang, S. L., Nguyen, V. B., Doan, C. T., Tran, T. N., Nguyen, M. T., & Nguyen, A. D., Production and potential applications of bioconversion of chitin and proteincontaining fishery byproducts into prodigiosin: A Review. *Molecules*, 25(12), 2744 (2020).
- Doan, C. T., Tran, T. N., Nguyen, V. B., Tran, T. D., Nguyen, A. D., & Wang, S. L., Bioprocessing of squid pens waste into chitosanase by *Paenibacillus* sp. TKU047 and its application in low-molecular weight chitosan oligosaccharides production. *Polymers*, **12**(5), 1163 (2020).
- Ismail, S. A., Microbial valorization of shrimp byproducts via the production of thermostable chitosanase and antioxidant chitooligosaccharides. *Biocatalysis and Agricultural Biotechnology*, 20, 101269 (2019).
- Ismail, S., & Emran, M., Direct microbial production of prebiotic and antioxidant chitin-oligosaccharides from shrimp byproducts. *Egyptian Journal of Aquatic Biology and Fisheries*, **24**(4), 181-195 (2020).
- Ismail, S. A., Hassan, M. E., & Hashem, A. M., Single step hydrolysis of chitin using thermophilic immobilized exochitinase on carrageenan-guar gum gel beads. *Biocatalysis and Agricultural Biotechnology*, 21, 101281 (2019).
- Liu, Y., & Dave, D., Recent progress on immobilization technology in enzymatic conversion of marine by-products to concentrated omega-3 fatty acids. *Green Chemistry*, (2021).
- 85. de Oliveira, D. A., Minozzo, M. G., Licodiedoff, S., &Waszczynskyj, N. Physicochemical and sensory characterization of refined and deodorized tuna (*Thunnusalbacares*) by-product oil obtained by enzymatic hydrolysis. *Food Chemistry*, **207**, 187-194 (2016).
- 86. Hamdi, M., Hammami, A., Hajji, S., Jridi, M., Nasri, M., &Nasri, R., Chitin extraction from blue crab (*Portunussegnis*) and shrimp (*Penaeuskerathurus*) shells using digestive alkaline proteases from *P. segnis* viscera.

International journal of biological macromolecules, **101**, 455-463 (2017).

- Doan, C. T., Tran, T. N., Vo, T. P. K., Nguyen, A. D., & Wang, S. L., Chitin extraction from shrimp waste by liquid fermentation using an alkaline proteaseproducing strain, *Brevibacillus parabrevis*. *International journal of biological macromolecules*, **131**, 706-715 (2019).
- Wang, C. H., Doan, C. T., Nguyen, V. B., Nguyen, A. D., & Wang, S. L., Reclamation of fishery processing waste: A mini-review. *Molecules*, 24(12), 2234 (2019).
- Zhang, Y., Dong, Y., & Dai, Z., Antioxidant and Cryoprotective Effects of Bone Hydrolysates from Bighead Carp (*Aristichthysnobilis*) in Freeze-Thawed Fish Fillets. *Foods*, **10**(6), 1409 (2021).
- Fu, X., Guo, Y., Jin, Y., & Ma, M., Bioconversion of chitin waste using a coldadapted chitinase to produce chitin oligosaccharides. *LWT*, **133**, 109863 (2020).
- Phadke, G. G., Rathod, N. B., Ozogul, F., Elavarasan, K., Karthikeyan, M., Shin, K. H., & Kim, S. K., Exploiting of Secondary Raw Materials from Fish Processing Industry as a Source of Bioactive Peptide-Rich Protein Hydrolysates. *Marine Drugs*, 19(9), 480 (2021).
- 92. Mohamed, F. H., El-sissi, A. F., Ismail, S. A., Ismail, S. A., &Hashem, A. M., The potentiality of using chitosan and its enzymatic depolymerized derivative chitooligosaccharides as immunomodulators. *Journal of Applied Pharmaceutical Science*, 8(12), 132-139 (2018).
- 93. Adhikari, H. S., & Yadav, P. N., Anticancer activity of chitosan, chitosan derivatives, and their mechanism of action. *International Journal of Biomaterials*, 2018.
- Hosseinnejad, M., &Jafari, S. M., Evaluation of different factors affecting antimicrobial properties of chitosan. *International journal of biological macromolecules*, 85, 467-475 (2016).
- 95. Cao, S., Liu, Y., Shi, L., Zhu, W., & Wang, H., N-Acetylglucosamine as a platform chemical produced from renewable resources: opportunity, challenge, and future prospects. *Green Chemistry*, (2022).
- 96. Muzzarelli, R. A., Biochemical significance of exogenous chitins and chitosans in

Egypt. J. Chem. 65, No. SI13B (2022)

animals and patients. *Carbohydrate Polymers*, **20**(1), 7-16 (1993).

- Ismail, S. A., El-Sayed, H. S., & Fayed, B., Production of prebiotic chitooligosaccharide and its nano/microencapsulation for the production of functional yoghurt. *Carbohydrate polymers*, 234, 115941 (2020).
- Hashem, A. M., Ismail, S., Hosny, A. E. D., Awad, G., & Ismail, S., Optimization of *Dothideomycetes* sp. NrC-SSW chitosanase productivity and activity using response surface methodology. *Egyptian Journal of Chemistry*, **61**(6), 973-987 (2018).
- Nidheesh, T., Salim, C., Rajini, P. S., & Suresh, P. V., Antioxidant and neuroprotective potential of chitooligomers in *Caenorhabditiselegans* exposed to Monocrotophos. *Carbohydrate polymers*, 135, 138-144 (2016).
- 100. Santos-Moriano, P., Fernandez-Arrojo, L., Mengibar, М., Belmonte-Reche, Е., Peñalver, P., Acosta, F. N., ... & Plou, F. J., Enzymatic production of fullv deacetylatedchitooligosaccharides and their and neuroprotective anti-inflammatory properties. **Biocatalysis** and Biotransformation, 36(1), 57-67 (2018).
- 101. Cui, D., Yang, J., Lu, B. and Shen, H., Efficient preparation of chitooligosaccharide with a potential chitosanase Csn-SH and its application for fungi disease protection. *Frontiers in Microbiology*, **12**, (2021).
- 102. Villalobos Solis, I., Engle, N.L., Spangler, M., Cottaz, S., Fort, S., Maeda, J., Ane, J.M., Tschaplinski, T., Labbe, J., Hettich, R.B. and Abraham, P., Expanding the Biological Role of Lipo-Chitooligosaccharides and Chitooligosaccharides in Laccaria bicolor Growth and Development. *Frontiers in Fungal Biology*, 3(1), (2022).
- 103. Sánchez, Á., Mengíbar, M., Rivera-Rodríguez, G., Moerchbacher, B., Acosta, N., & Heras, A., The effect of preparation processes on the physicochemical characteristics and antibacterial activity of chitooligosaccharides. *Carbohydrate polymers*, **157**, 251-257 (2017).
- 104. Liu, P., Li, H., Gong, J., Geng, Y., Jiang, M., Xu, H., Xu, Z. and Shi, J., Chitooligosaccharides alleviate hepatic fibrosis by regulating the polarization of M1 and M2 macrophages. *Food and Function*. 13(2), 753-768 (2022).
- 105. Cao, P., Huang, G., Yang, Q., Guo, J., & Su, Z., The effect of chitooligosaccharides on oleic acid-induced lipid accumulation in

HepG2 cells. *Saudi Pharmaceutical Journal*, **24**(3), 292-298 (2016).

- 106. Tavano, O. L., Berenguer-Murcia, A., Secundo, F., & Fernandez-Lafuente, R., Biotechnological applications of proteases in food technology. *Comprehensive Reviews in Food Science and Food Safety*, **17**(2), 412-436 (2018).
- 107. Barrett, A. J., & McDONALD, J. K., Nomenclature: protease, proteinase and peptidase. *Biochemical Journal*, 237(3), 935-935 (1986).
- 108. Suresh, P. V., Sakhare, P. Z., Sachindra, N. M., &Halami, P. M., Extracellular chitin deacetylase production in solid state fermentation by native soil isolates of *Penicillium monoverticillium* and *Fusarium* oxysporum. Journal of food science and technology, **51**(8), 1594-1599 (2014).
- 109. Beygmoradi, A., Homaei, A., Hemmati, R., Santos-Moriano, P., Hormigo, D., &Fernández-Lucas, J., Marine chitinolytic enzymes, a biotechnological treasure hidden in the ocean?. *Applied microbiology and biotechnology*, **102**(23), 9937-9948 (2018).
- 110. Cristóvao, R. O., Botelho, C. M., Martins, R. J., Loureiro, J. M., &Boaventura, R. A., Fish canning industry wastewater treatment for water reuse–a case study. *Journal of Cleaner Production*, 87, 603-612.
- 111. Arvanitoyannis, I. S., &Kassaveti, A., Fish industry waste: treatments, environmental impacts, current and potential uses. *International journal of food science & technology*, **43**(4), 726-745 (2008).
- 112. Fluence, Waste-to-energy Technology Helps Fish Processor Save on Operating Costs, in: Https://Www.Fluencecorp.Com/Case/Waste -to-Energy-for-Fish- Processing-Plant (2019).
- 113. Venugopal, V., & Sasidharan, A., Seafood industry effluents: environmental hazards, treatment and resource recovery. *Journal of Environmental Chemical Engineering*, 9(2), 104758 (2021).
- 114. Katiyar, R., Gurjar, B. R., Biswas, S., Pruthi, V., Kumar, N., & Kumar, P., Microalgae: an emerging source of energy based bio-products and a solution for environmental issues. *Renewable and Sustainable Energy Reviews*, **72**, 1083-1093 (2017).
- 115. Gao, F., Peng, Y. Y., Li, C., Yang, G. J., Deng, Y. B., Xue, B., &Guo, Y. M.,

Egypt. J. Chem. 65, No. SI13B (2022)

Simultaneous nutrient removal and biomass/lipid production by Chlorella sp. in seafood processing wastewater. *Science of the Total Environment*, **640**, 943-953 (2018).

- 116. Hsu, K. C., Lu, G. H., &Jao, C. L., peptides Antioxidative properties of from prepared tuna cooking juice hydrolysates with orientase (Bacillus subtilis). Food Research International, 42(5-6), 647-652 (2009).
- 117. Tang, W., Zhang, H., Wang, L., Qian, H., & Qi, X., Targeted separation of antibacterial peptide from protein hydrolysate of anchovy cooking wastewater by equilibrium dialysis. *Food chemistry*, **168**, 115-123 (2015).
- 118. Pérez-Santín, E., Calvo, M. M., López-Caballero, M. E., Montero, P., & Gómez-Guillén, M. C., Compositional properties and bioactive potential of waste material from shrimp cooking juice. *LWT-Food Science and Technology*, **54**(1), 87-94 (2013).
- 119. Pacheco-Aguilar, R., Leyva-Soto, P., Carvallo-Ruiz, G., García-Carreño, L. F., &Márquez-Ríos, E., Efecto de la concentración de quitosano y pH sobre la remoción de sólidos en agua de cola de la industriasardinera. *Interciencia*, **34**(4), 274-279 (2009).
- 120. Chalamaiah, M., Hemalatha, R., &Jyothirmayi, T., Fish protein hydrolysates: proximate composition, amino acid composition, antioxidant activities and applications: a review. *Food chemistry*, 135(4), 3020-3038 (2012).
- 121. Zamora-Sillero, J., Gharsallaoui, A., & Prentice, C., Peptides from fish by-product protein hydrolysates and its functional properties: An overview. *Marine Biotechnology*, **20**(2), 118-130 (2018).
- 122.Hung, C. C., Yang, Y. H., Kuo, P. F., & Hsu, K. C., Protein hydrolysates from tuna cooking juice inhibit cell growth and induce apoptosis of human breast cancer cell line MCF-7. *Journal of Functional Foods*, **11**, 563-570 (2014).
- 123. Tonon, R. V., dos Santos, B. A., Couto, C. C., Mellinger-Silva, C., Brígida, A. I. S., & Cabral, L. M., Coupling of ultrafiltration and enzymatic hydrolysis aiming at valorizing shrimp wastewater. *Food chemistry*, **198**, 20-27 (2016).
- 124. Mahdabi, M., & HosseiniShekarabi, S. P., A comparative study on some functional and antioxidant properties of kilka meat, fishmeal, and stickwater protein

hydrolysates. Journal of Aquatic Food Product Technology, **27**(7), 844-858 (2018).

- 125. Pan, X., Zhao, Y. Q., Hu, F. Y., & Wang, B., Preparation and identification of antioxidant peptides from protein hydrolysate of skate (*Raja porosa*) cartilage. *Journal of functional foods*, **25**, 220-230 (2016).
- 126. Taheri, A., Farvin, K. S., Jacobsen, C., & Baron, C. P., Antioxidant activities and functional properties of protein and peptide fractions isolated from salted herring brine. *Food chemistry*, **142**, 318-326 (2014).
- 127. Gringer, N., Hosseini, S. V., Svendsen, T., Undeland, I., Christensen, M. L., & Baron, C. P., Recovery of biomolecules from marinated herring (*Clupeaharengus*) brine using ultrafiltration through ceramic membranes. *LWT-Food Science and Technology*, **63**(1), 423-429 (2015).
- 128. Tremblay, A., Corcuff, R., Goulet, C., Godefroy, S. B., Doyen, A., & Beaulieu, L., Valorization of snow crab (*Chionoecetesopilio*) cooking effluents for food applications. *Journal of the Science of Food and Agriculture*, **100**(1), 384-393 (2020).
- 129. Amado, I. R., Vázquez, J. A., González, M. P., & Murado, M. A., Production of antihypertensive and antioxidant activities by enzymatic hydrolysis of protein concentrates recovered by ultrafiltration from cuttlefish processing wastewaters. *Biochemical engineering journal*, **76**, 43-54 (2013).
- 130. Huang, Y. B., Wang, X. F., Wang, H. Y., Liu, Y., & Chen, Y., Studies on mechanism of action of anticancer peptides by modulation of hydrophobicity within a defined structural framework. *Molecular cancer therapeutics*, **10**(3), 416-426 (2011).
- Bechtel, P. J., Properties of stickwater from fish processing byproducts. *Journal of Aquatic Food Product Technology*, 14(2), 25-38 (2005).
- 132. Garcia-Sifuentes, C., Pacheco-Aguilar, R., Lugo-Sánchez, M., Garcia-Sánchez, G., Ramirez-Suarez, J. C., & Garcia-Carreno, F., Properties of recovered solids from stickwater treated by centrifugation and pH shift. *Food Chemistry*, **114**(1), 197-203 (2009).
- 133. Alkaya, E., & Demirer, G. N., Minimizing and adding value to seafood processing

wastes. *Food and Bioproducts Processing*, **100**, 195-202 (2016).

- 134. Monteiro, A., Paquincha, D., Martins, F., Queirós, R. P., Saraiva, J. A., Švarc-Gajić, J., ... & Carvalho, A. P., Liquid by-products from fish canning industry as sustainable sources of ω3 lipids. *Journal of environmental management*, **219**, 9-17 (2018).
- 135. Jarrault, C., Dornier, M., Labatut, M. L., Giampaoli, P., &Lameloise, M. L., Coupling nanofiltration and osmotic evaporation for the recovery of a natural flavouring concentrate from shrimp cooking juice. *Innovative Food Science & Emerging Technologies*, **43**, 182-190 (2017).
- 136. de Farias Silva, C. E., Barbera, E., &Bertucco, A., Biorefinery as a promising approach to promote ethanol industry from microalgae and cyanobacteria. In *Bioethanol production from food crops*, 343-359 (2019).
- 137. Khoo, K. S., Lee, S. Y., Ooi, C. W., Fu, X., Miao, X., Ling, T. C., & Show, P. L., Recent advances in biorefinery of astaxanthin from *Haematococcus pluvialis*. *Bioresource technology*, **288**, 121606 (2019).
- 138. Mitra, M., & Mishra, S., Multiproduct biorefinery from *Arthrospira* spp. towards zero waste: Current status and future trends. *Bioresource technology*, **291**, 121928 (2019).
- 139. Kratky, L., & Zamazal, P., Economic feasibility and sensitivity analysis of fish waste processing biorefinery. *Journal of Cleaner Production*, 243, 118677 (2020).
- 140. Abdollahi, M., Rezaei, M., Jafarpour, A., &Undeland, I., Sequential extraction of gelforming proteins, collagen and collagen hydrolysate from gutted silver carp (*Hypophthalmichthysmolitrix*), a biorefinery approach. *Food Chemistry*, **242**, 568-578 (2018).
- 141. Hülsey, M. J., Shell biorefinery: A comprehensive introduction. *Green Energy* & *Environment*, **3**(4), 318-327 (2018).